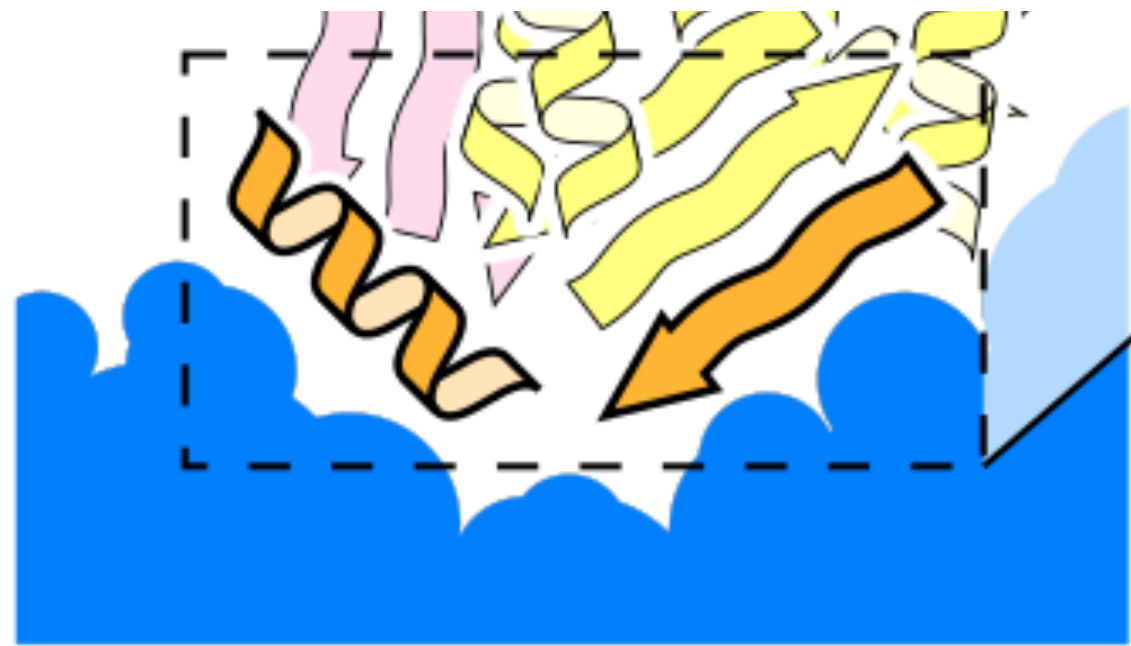


# Diffusion modeling of protein backbones for the motif-scaffolding problem

**Brian Trippe & Jason Yim**

# Computational Protein Design Workflow

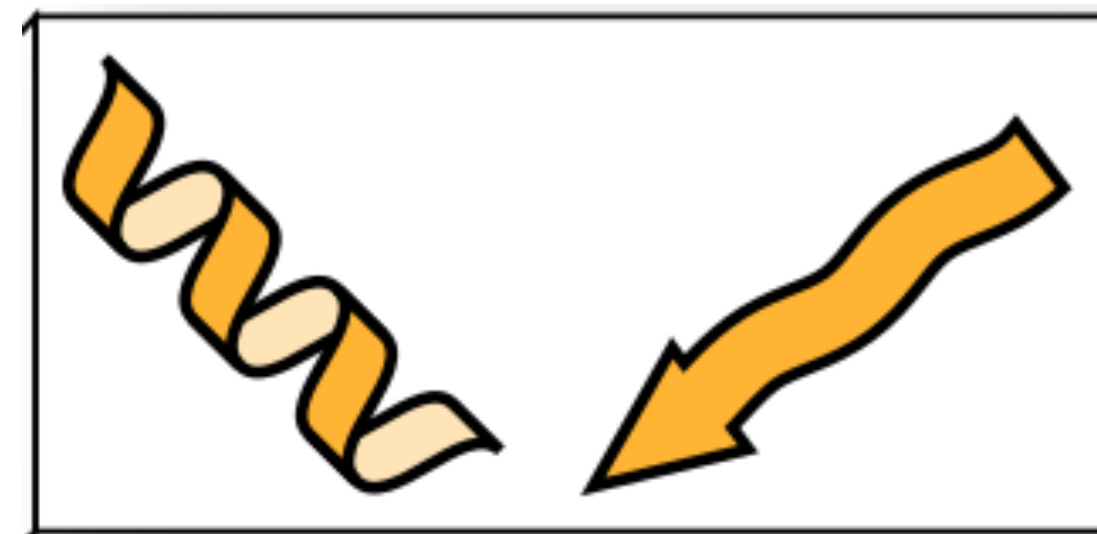
Desired Function  
(e.g. binding)



Motif  
Identification



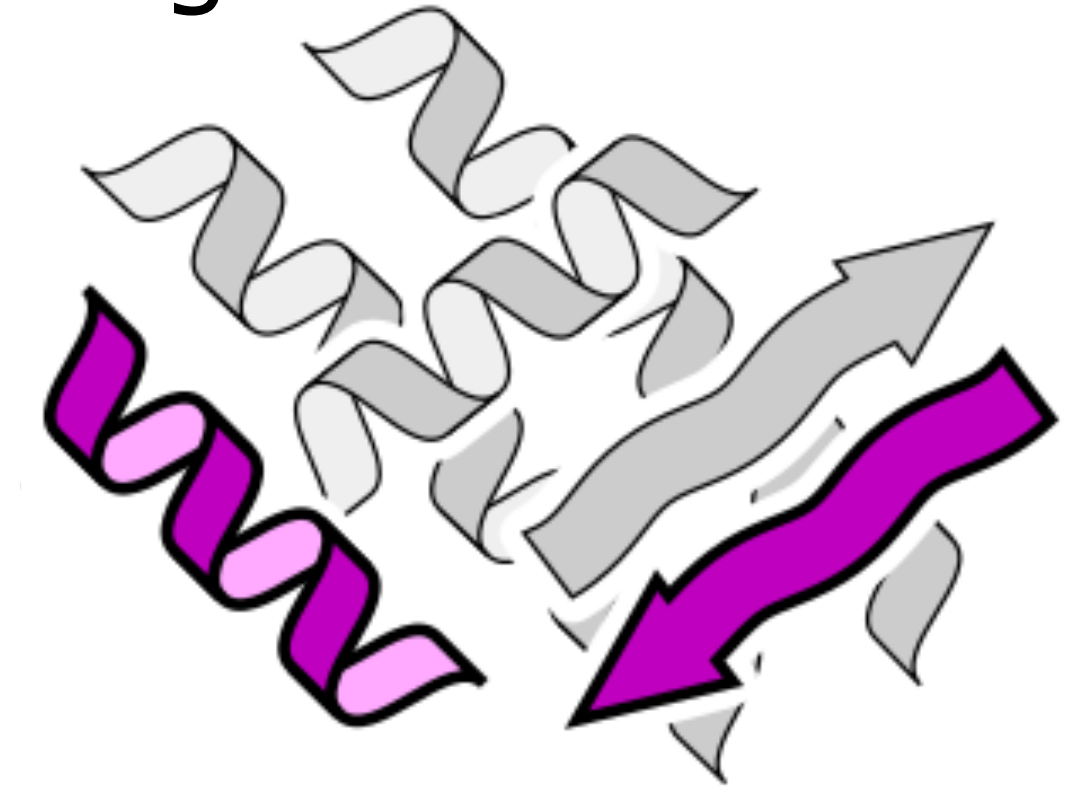
Functional "**Motif**"



Motif-Scaffolding

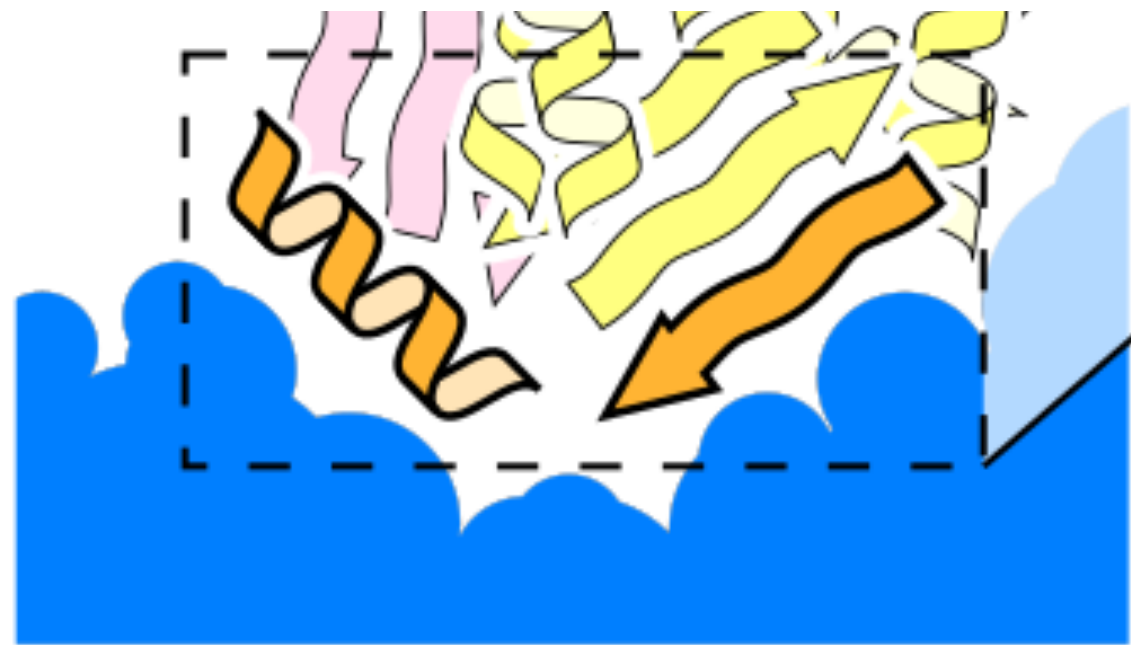


Designed "**Scaffold**"



# Computational Protein Design Workflow

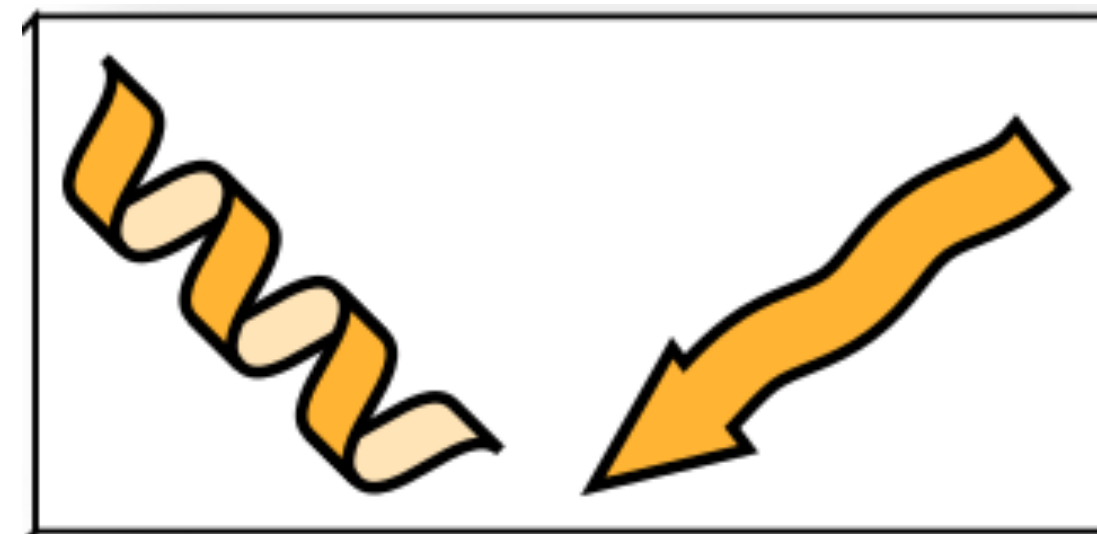
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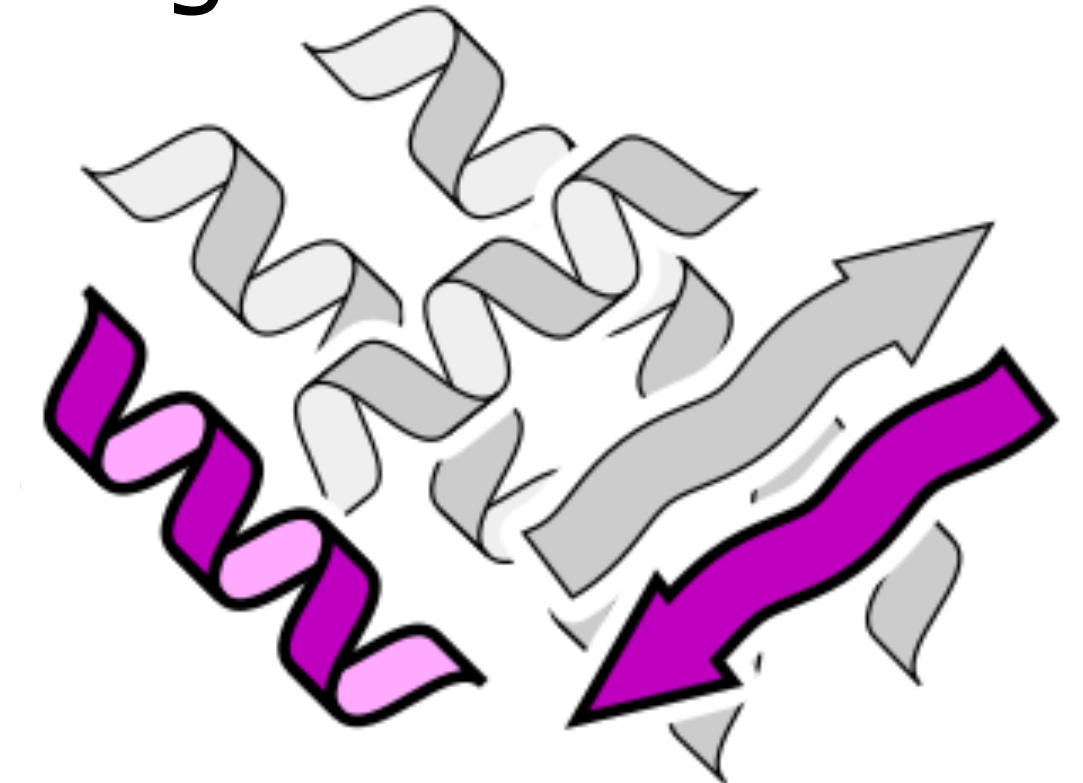
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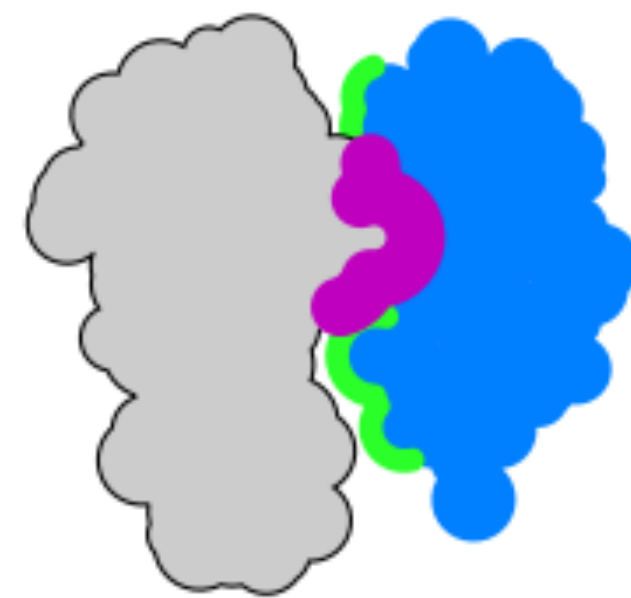
Motif-Scaffolding



Designed "**Scaffold**"



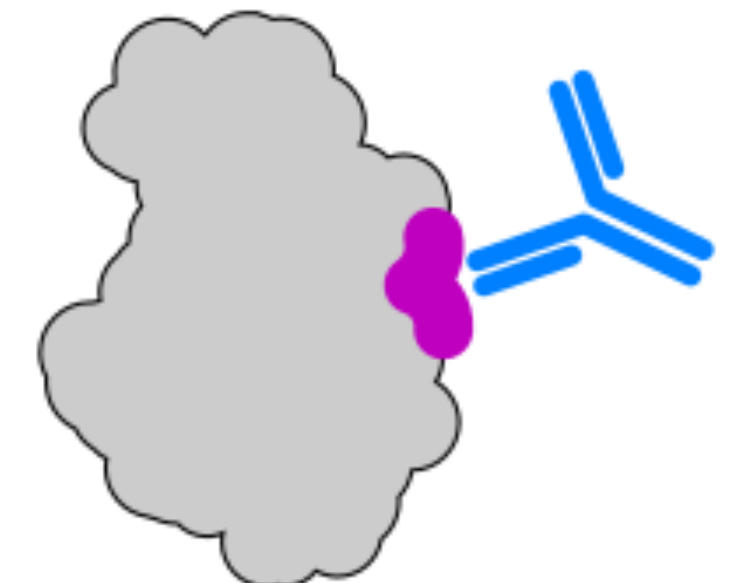
Motifs can have various  
functions and sources



**Protein interaction  
interfaces**  
(via fragment docking)



**Catalytic & metal-  
binding sites**  
(quantum chemistry)



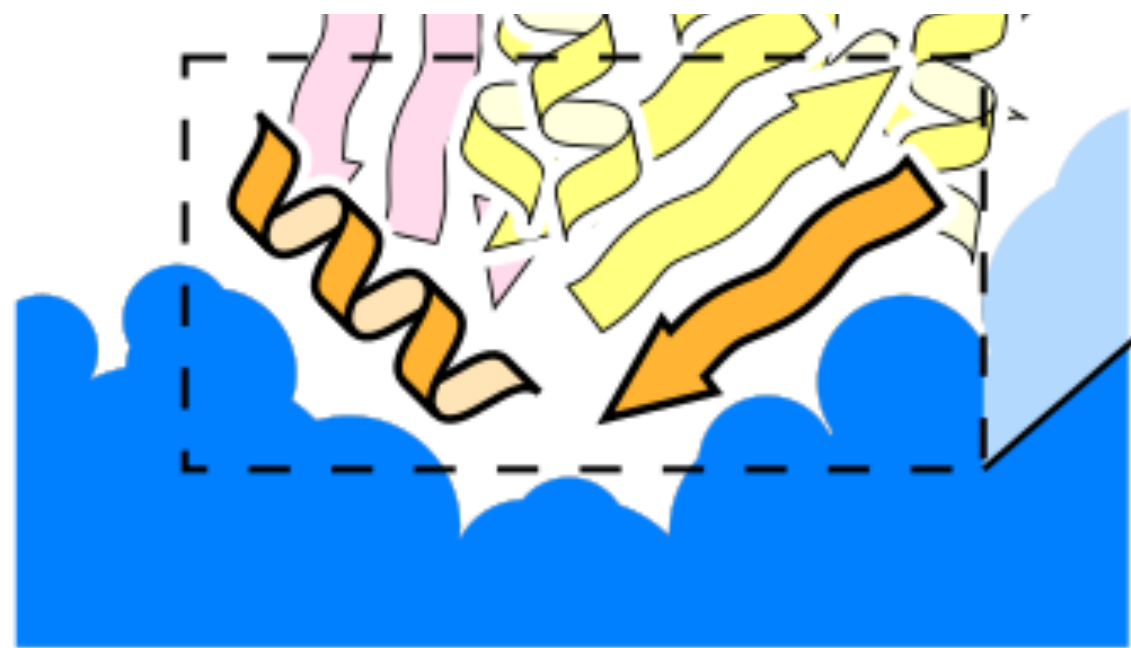
**Epitope  
presentation**  
(Native interface)

[Figure credit: Doug  
Tischer & David Juergens]

# Computational Protein Design Workflow

This talk

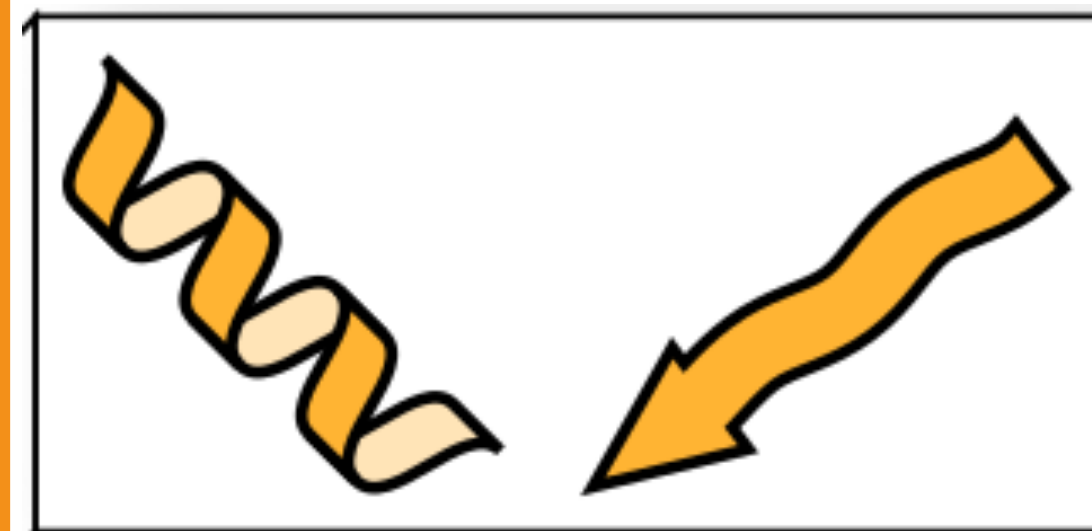
Desired Function  
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Motif  
Identification



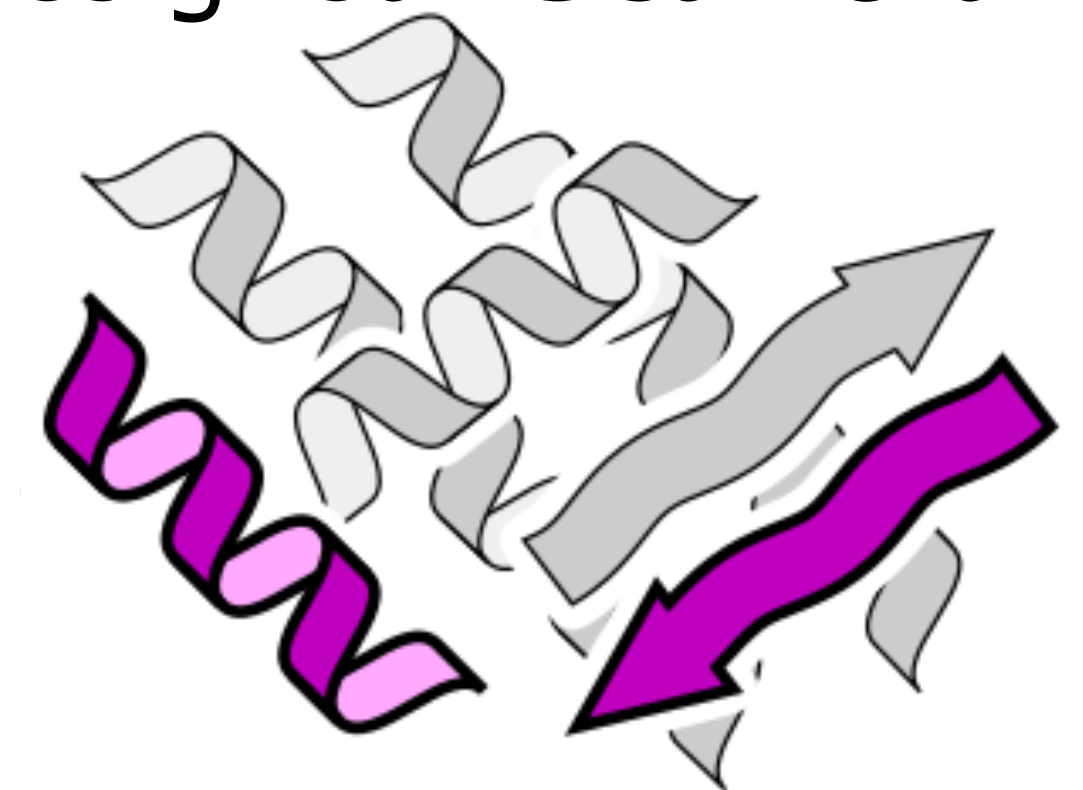
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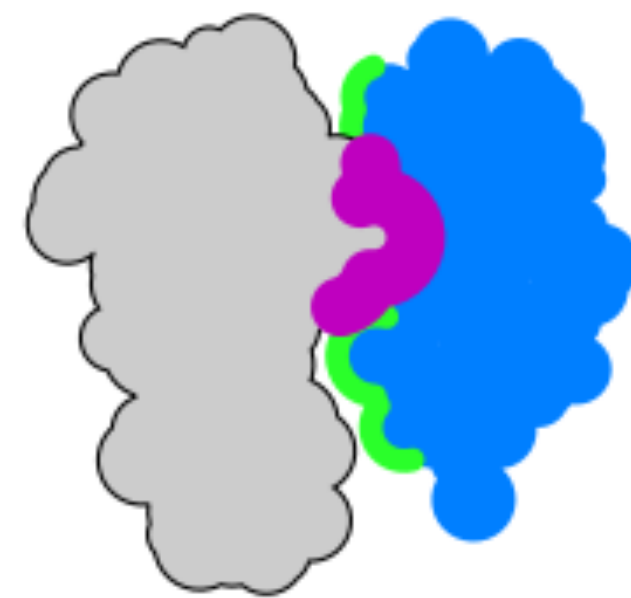
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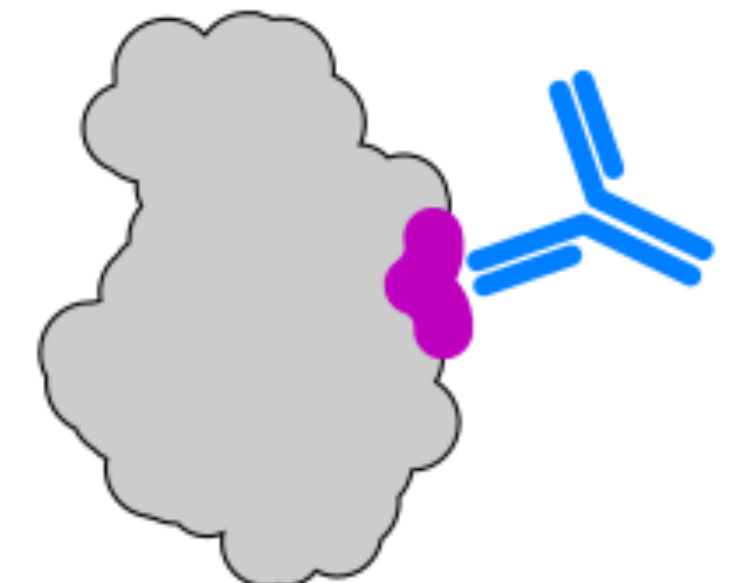
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**Post-AlphaFold, protein design is  
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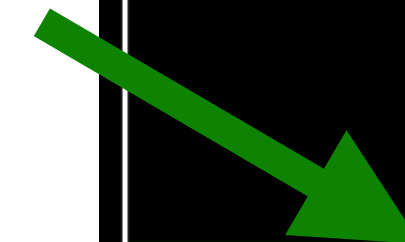
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**Post-AlphaFold, protein design is  
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- ~~Naive guessing?~~  $\sim 20^{100}$  sequences!

All Structures (x)

Structures  
with motif





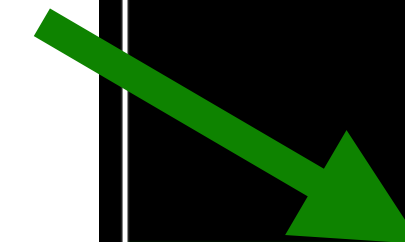
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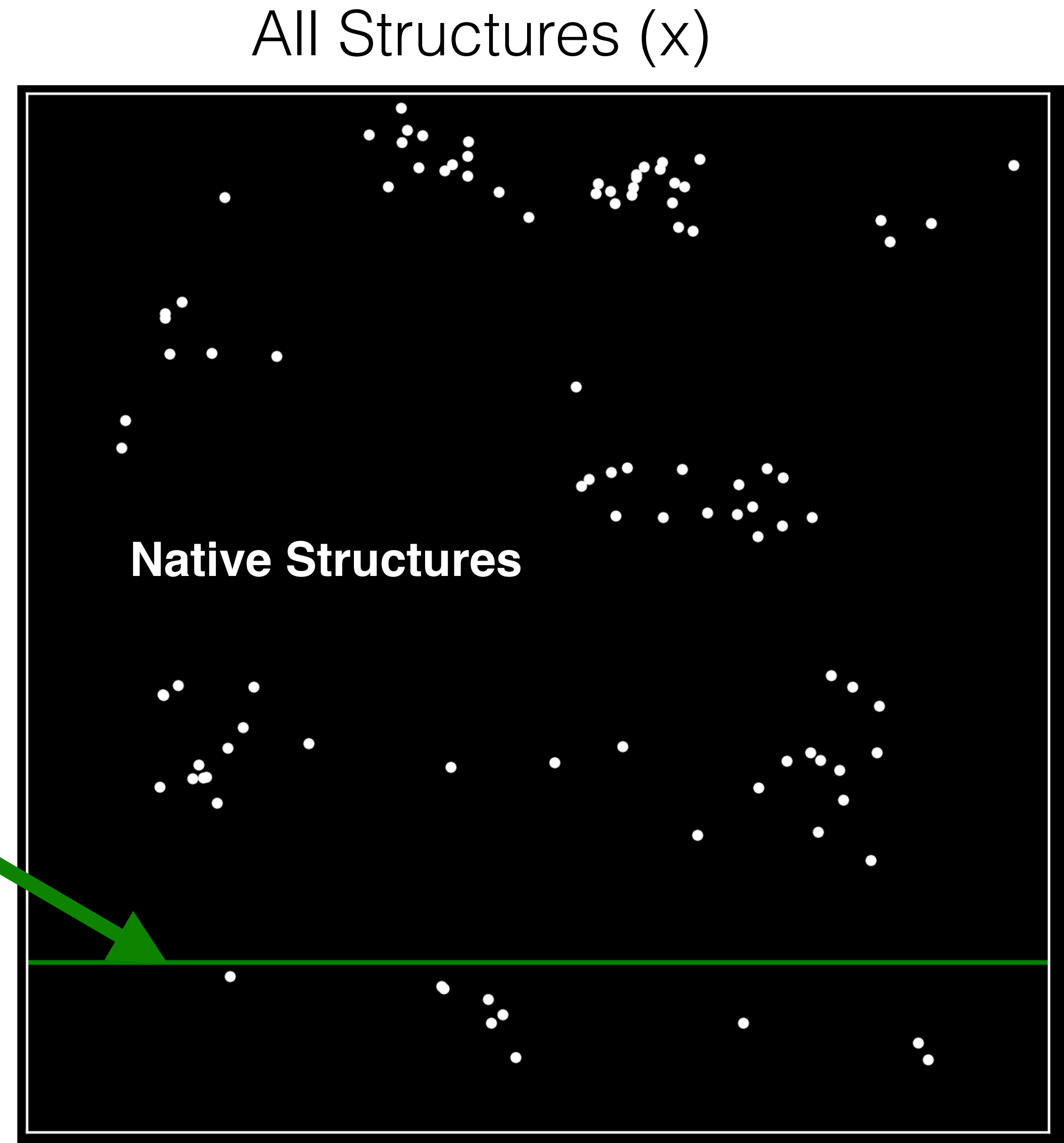


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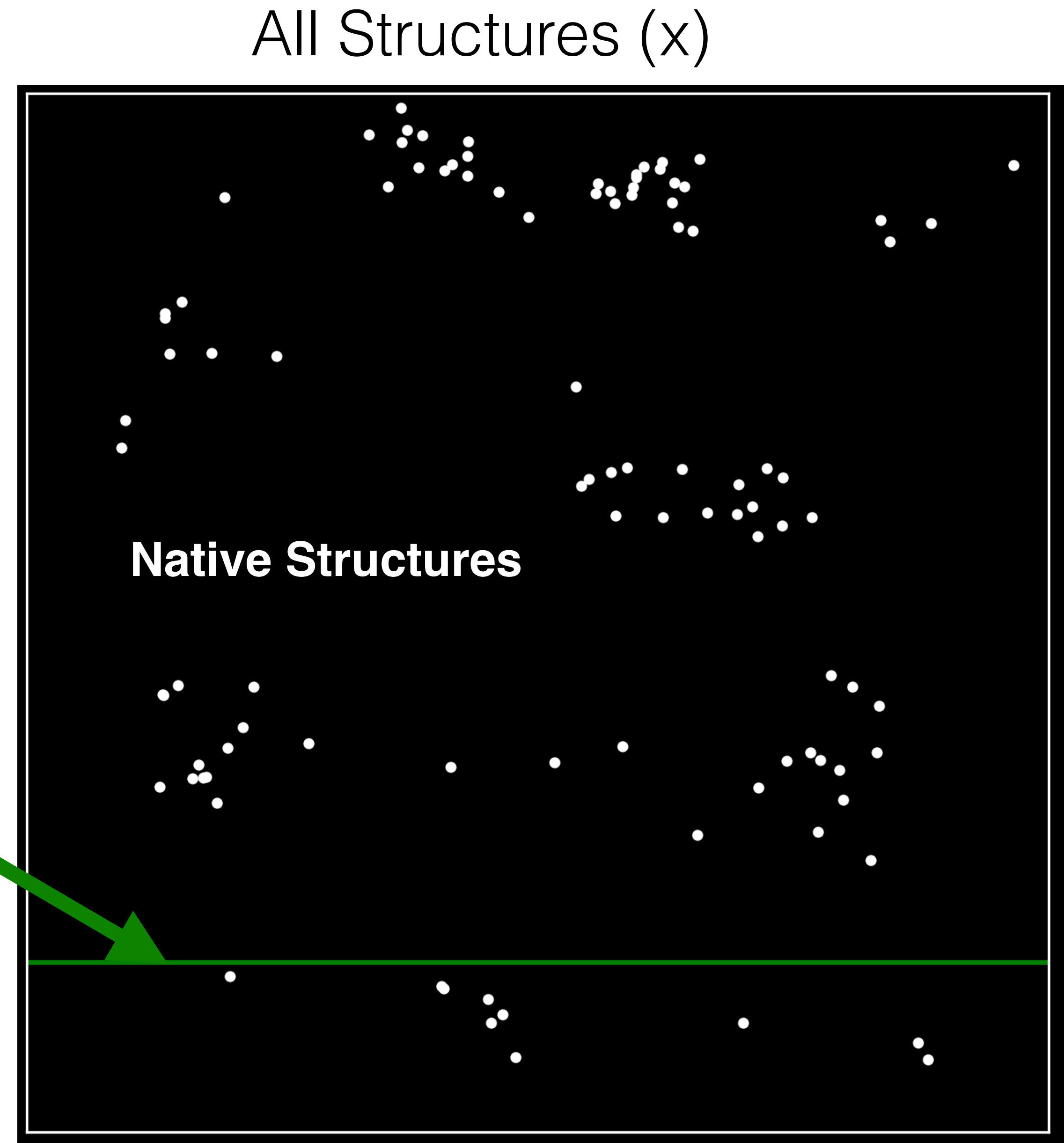


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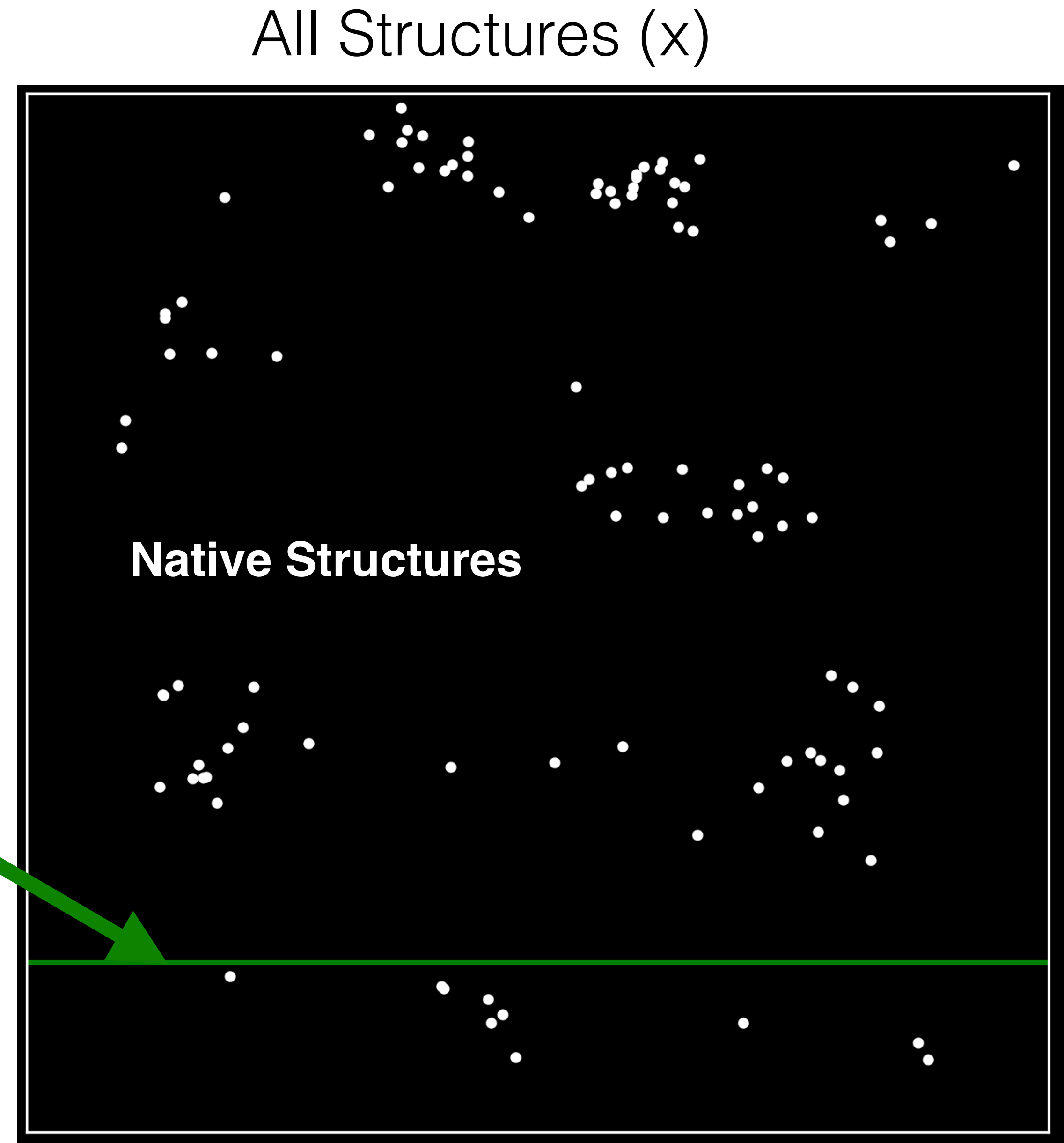


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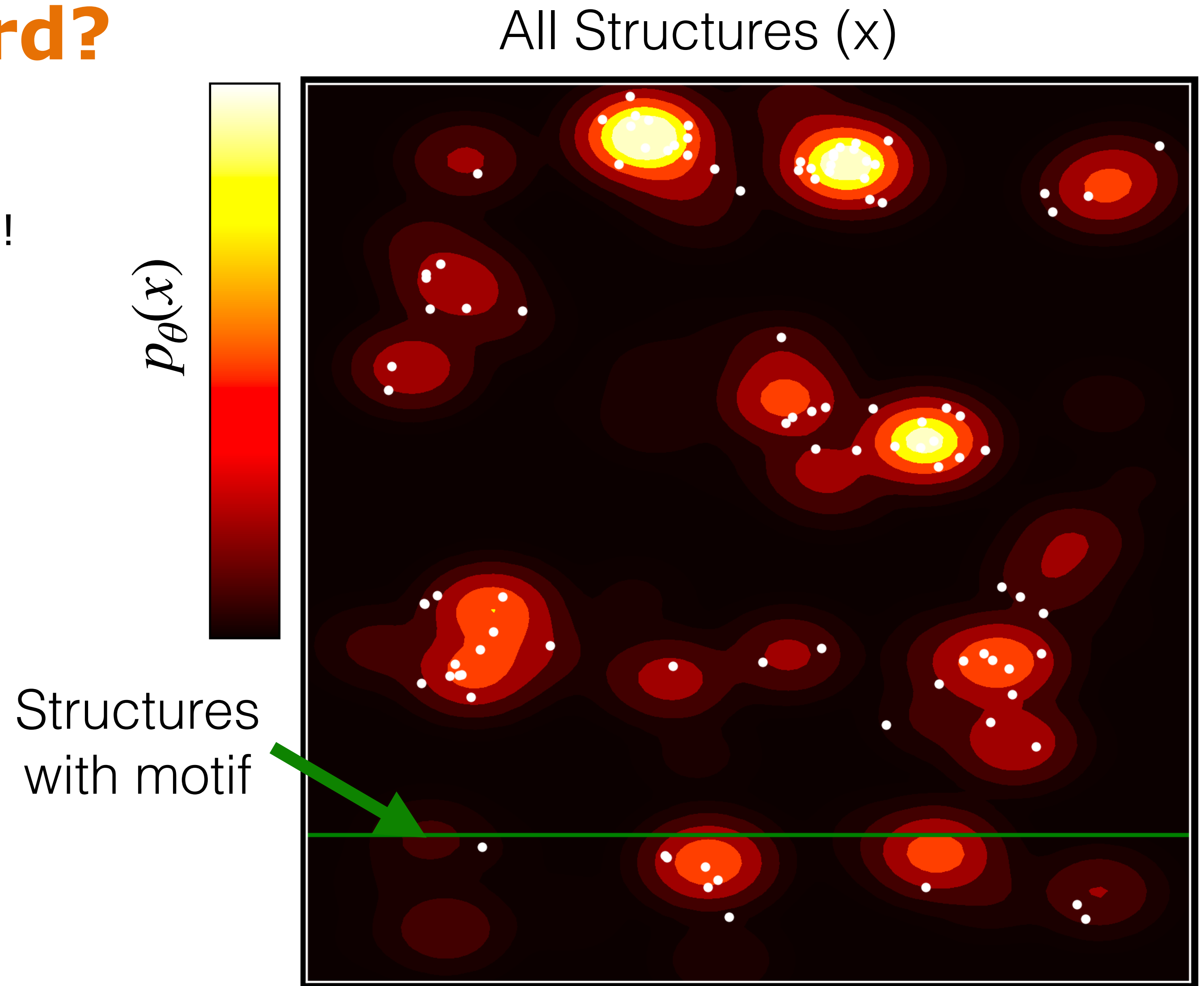




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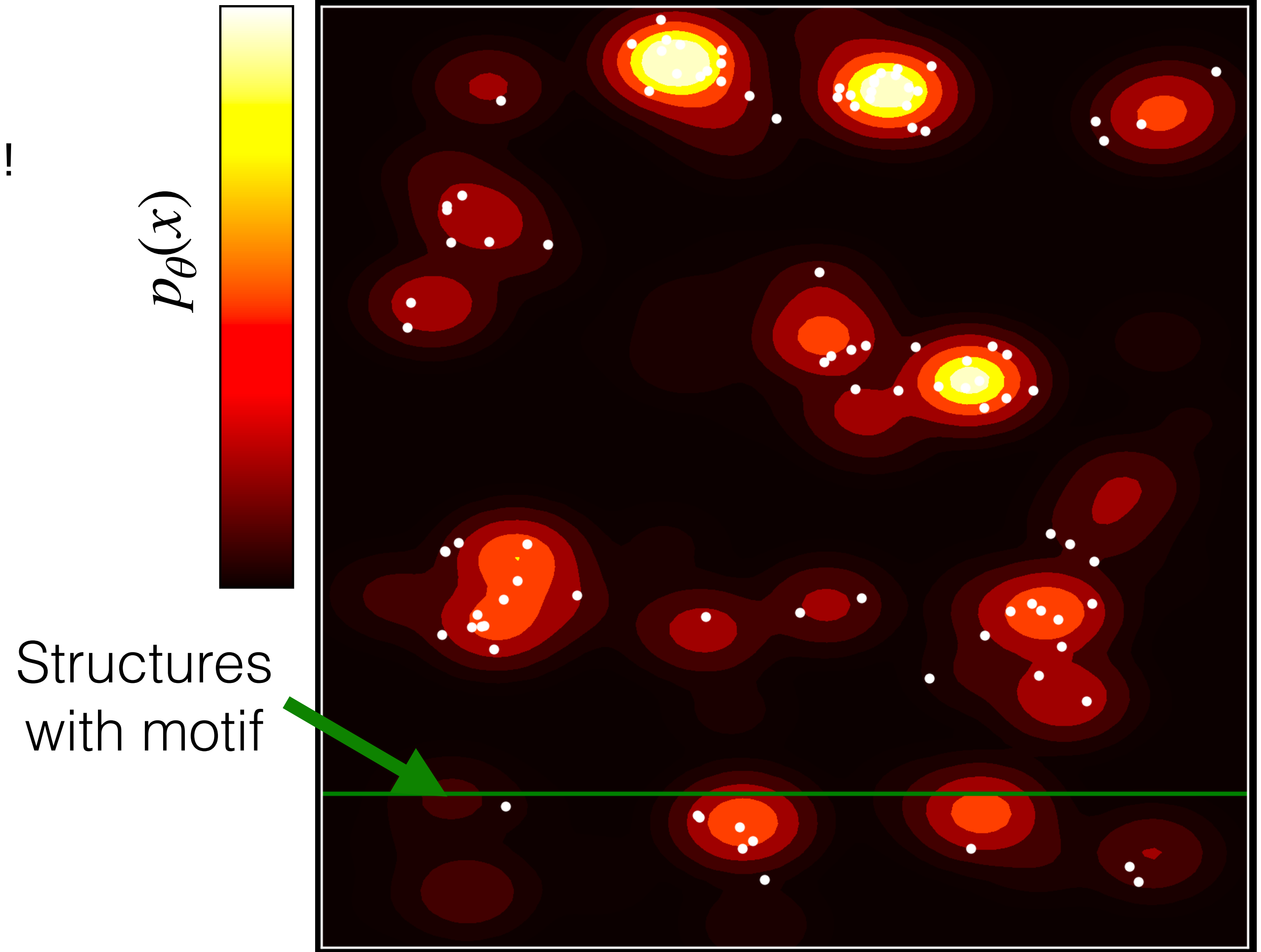
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**Motif**

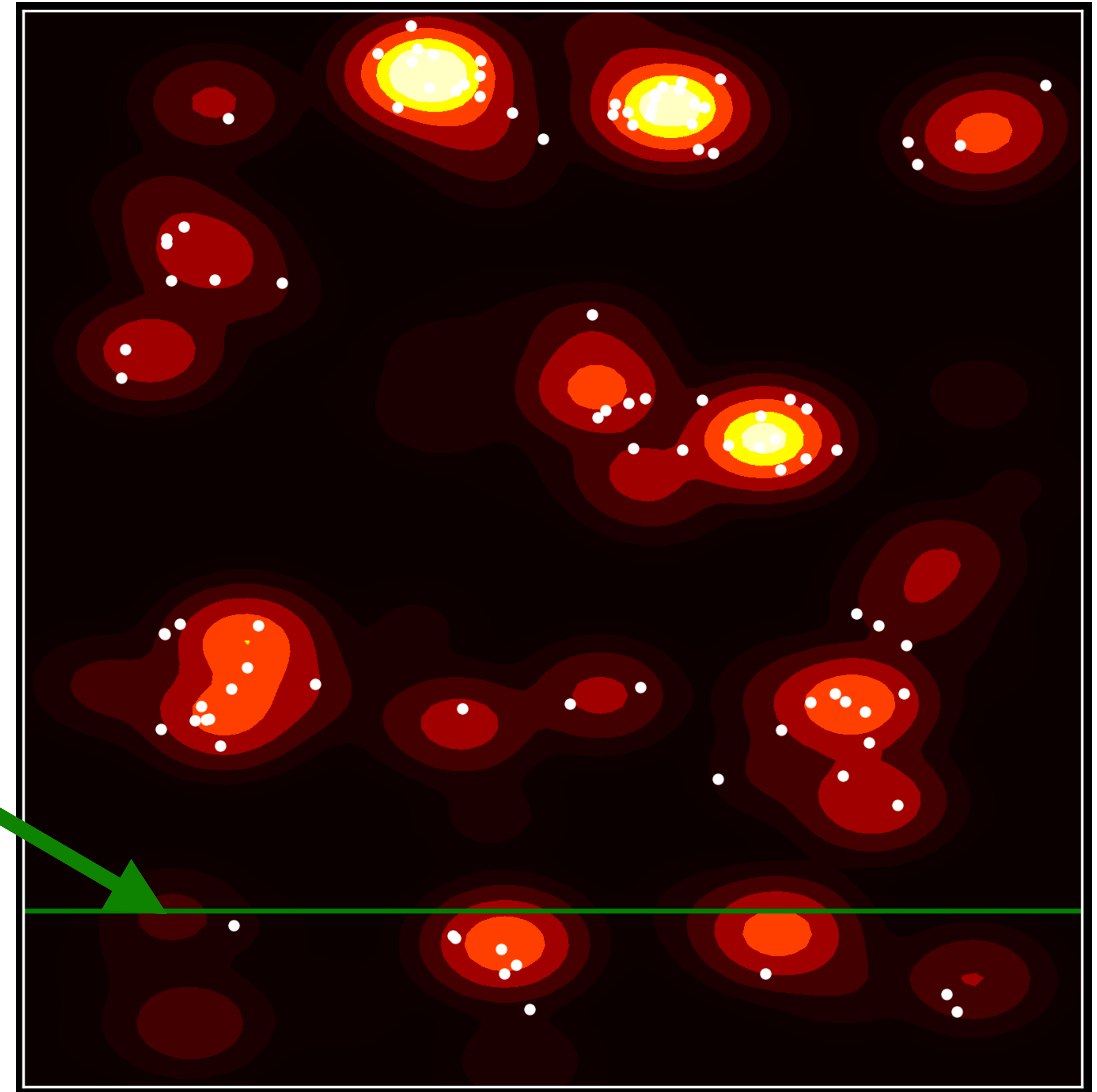
**Scaffold**

- Draw  $x_S \sim p_{\theta}(x_S \mid x_M)$

$p_{\theta}(x)$

Structures with motif

All Structures (x)



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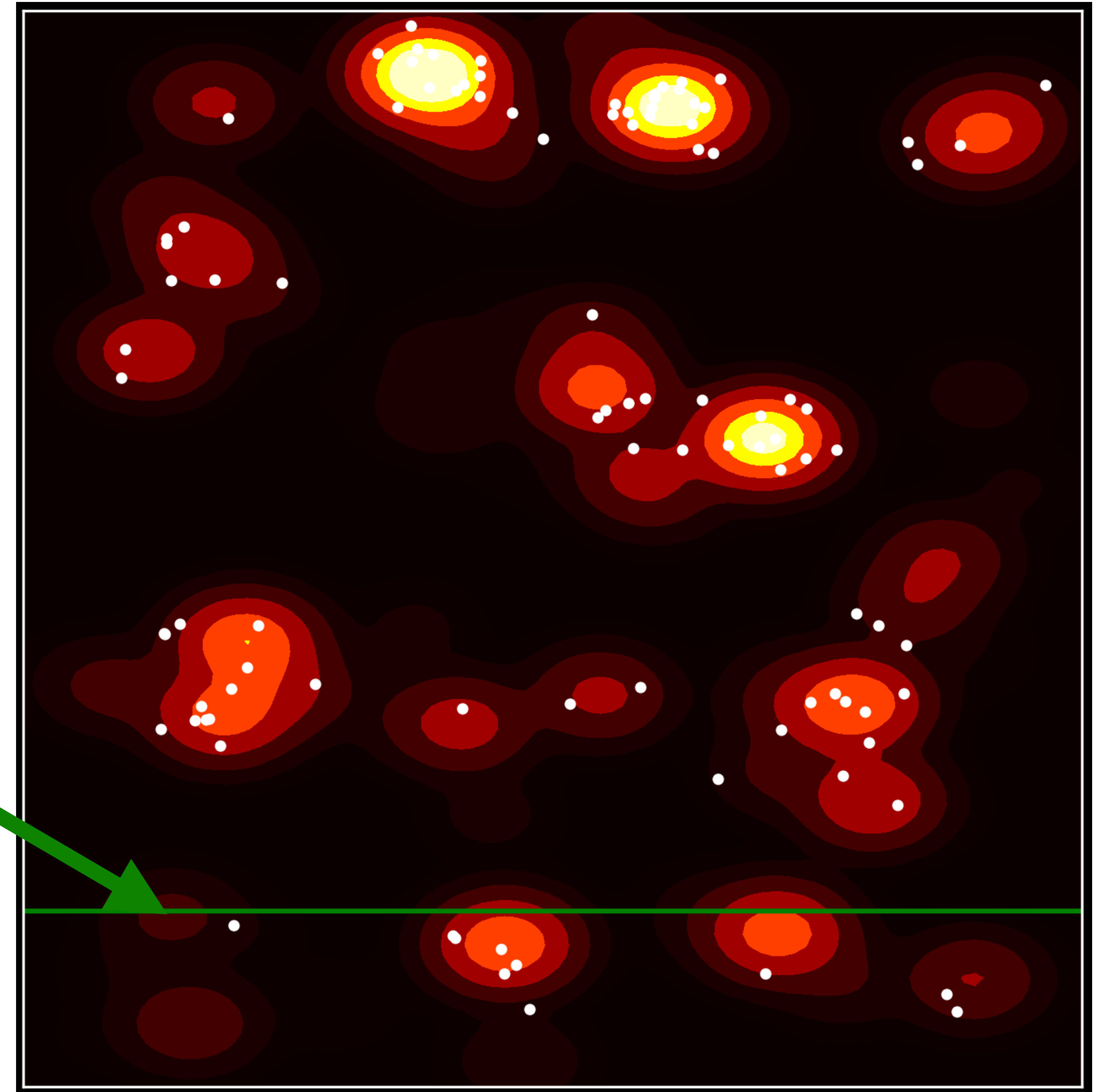
**Scaffold**

- Draw  $x_S \sim p_{\theta}(x_S | x_M)$

$p_{\theta}(x)$

Structures with motif

All Structures (x)



**Key Tool:** Diffusion generative models & sequential Monte Carlo



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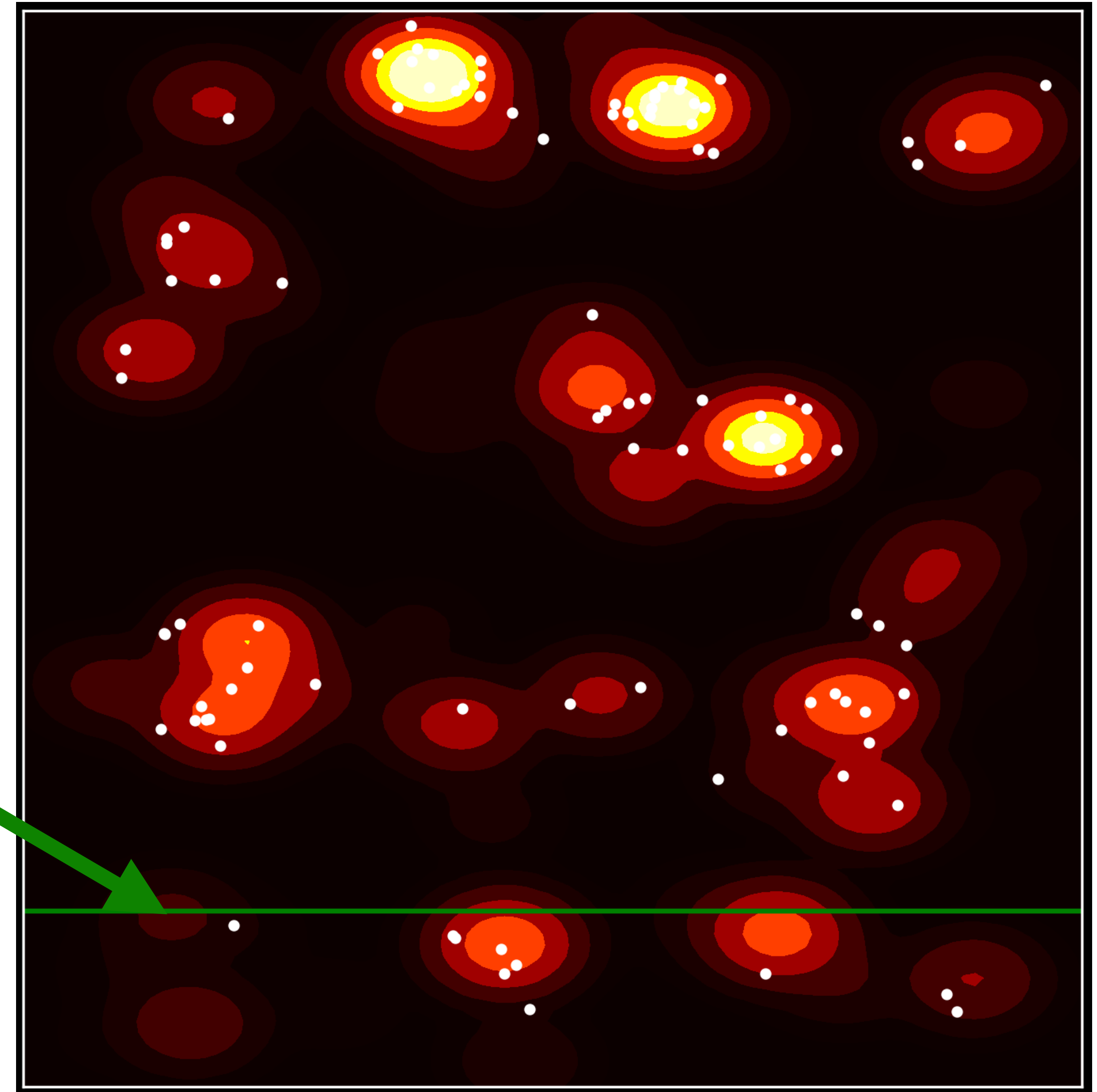
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**Motif**      **Scaffold**
  - Draw  $x_S \sim p_{\theta}(x_S | x_M)$

Structures with motif

$p_{\theta}(x)$

All Structures (x)



**Key Tool:** Diffusion generative models & sequential Monte Carlo

**We show:** Methods with potential to build long, diverse scaffolds

# Roadmap



# Roadmap

## Learning $p_{\theta}(x)$ [ProtDiff]

- Diffusion generative modeling background
- Adapting diffusion for protein backbones
- Model performance and limitations

# Roadmap

## **Learning** $p_{\theta}(x)$ [**ProtDiff**]

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- Why conditional sampling vs. “naive” in-painting?
- Sequential Monte Carlo for exact sampling in the large-compute limit



# Roadmap

## **Learning $p_{\theta}(x)$ [ProtDiff]**

- Diffusion generative modeling background
- Adapting diffusion for protein backbones
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- Why conditional sampling vs. “naive” in-painting?
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## **Limitations, Related Work, and Future directions**

# Diffusion models on protein backbones

Figures/slides borrowed from:

- CVPR 2022 Tutorial: Denoising Diffusion-based Generative Modeling: Foundations and Applications



# State-of-the-art

## DALL·E 2

*“a propaganda poster depicting a cat dressed as french emperor napoleon holding a piece of cheese”*



## IMAGEN

*“A photo of a raccoon wearing an astronaut helmet, looking out of the window at night.”*





# Denoising diffusion probabilistic models

## General idea

Data



Noise

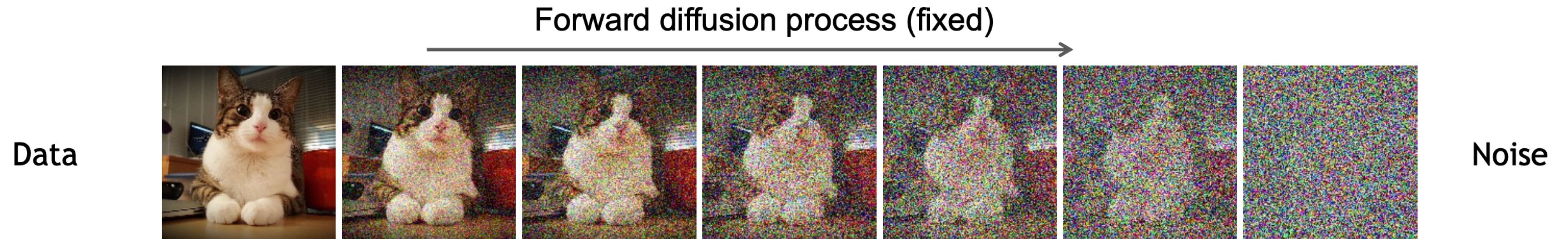
[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

## General idea

- Forward *diffusion* process gradually adds noise to input data.

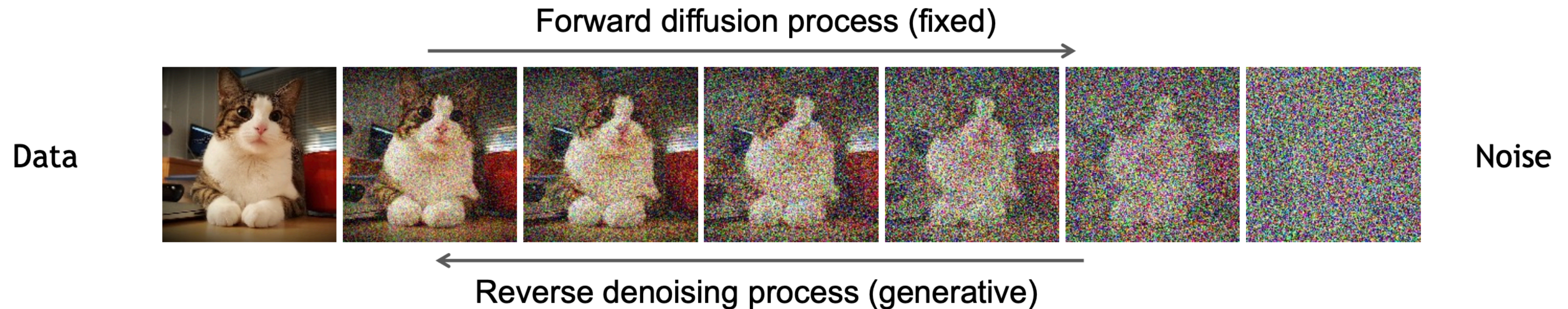




# Denoising diffusion probabilistic models

## General idea

- Forward *diffusion* process gradually adds noise to input data.
- Reverse *denoising* process generates data by removing noise.





# Denoising diffusion probabilistic models

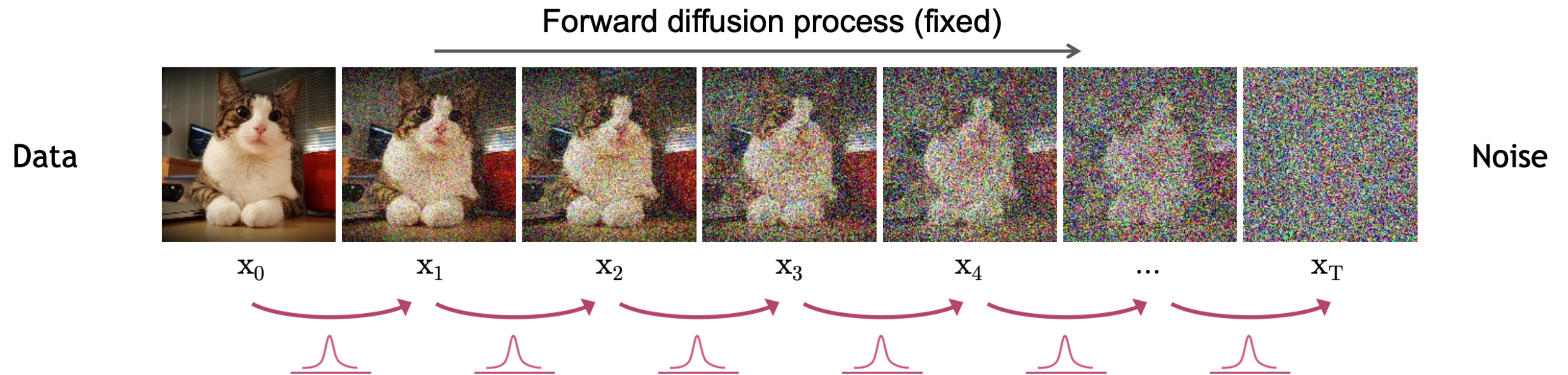
## Forward diffusion





# Denoising diffusion probabilistic models

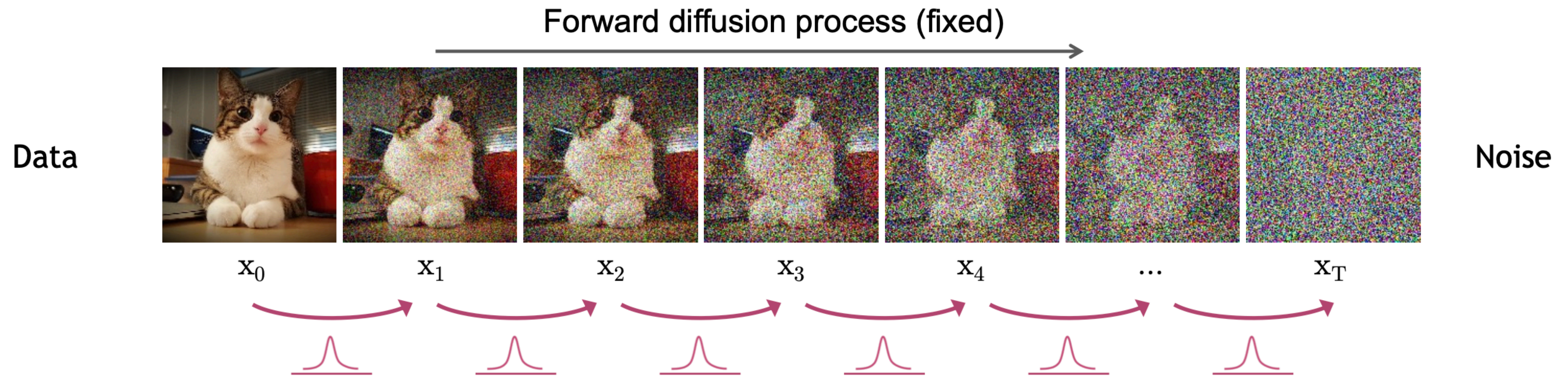
## Forward diffusion





# Denoising diffusion probabilistic models

## Forward diffusion



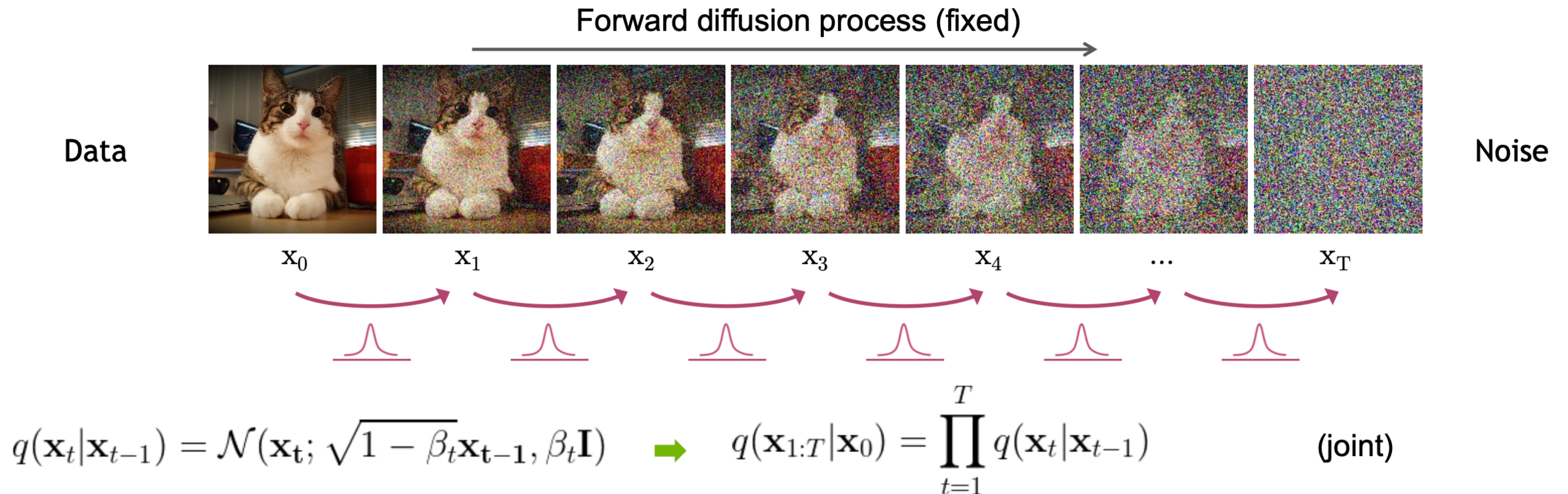
$$q(\mathbf{x}_t | \mathbf{x}_{t-1}) = \mathcal{N}(\mathbf{x}_t; \sqrt{1 - \beta_t} \mathbf{x}_{t-1}, \beta_t \mathbf{I})$$

- $\beta_t$  how much noise is added on each step
- [Ho et al]:  $\beta_0 = 0.0001$ ,  $\beta_T = 0.2$ ,  $\beta_{t-1} < \beta_t$
- $(1 - \beta_t)$  is how much signal is kept.



# Denoising diffusion probabilistic models

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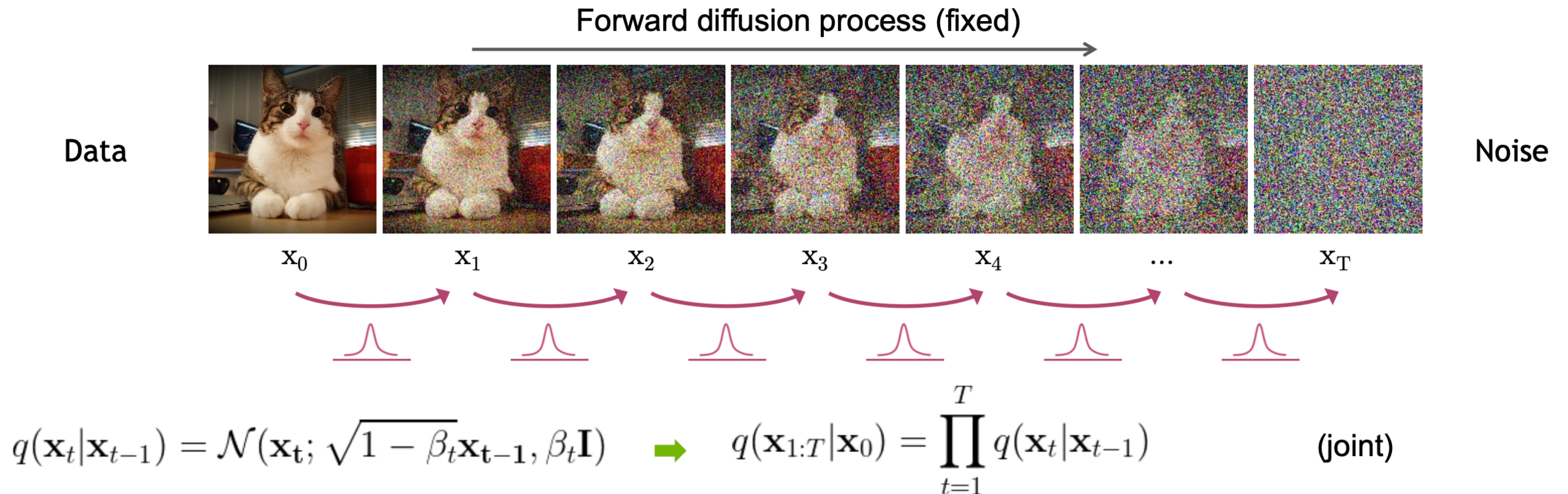


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- $(1 - \beta_t)$  is how much signal is kept.

- Training requires sampling every  $x_t$
- But this is expensive for large  $t$  :  $q(x_t | x_{t-1}) \cdot q(x_{t-1} | x_{t-2}) \dots q(x_1 | x_0)$

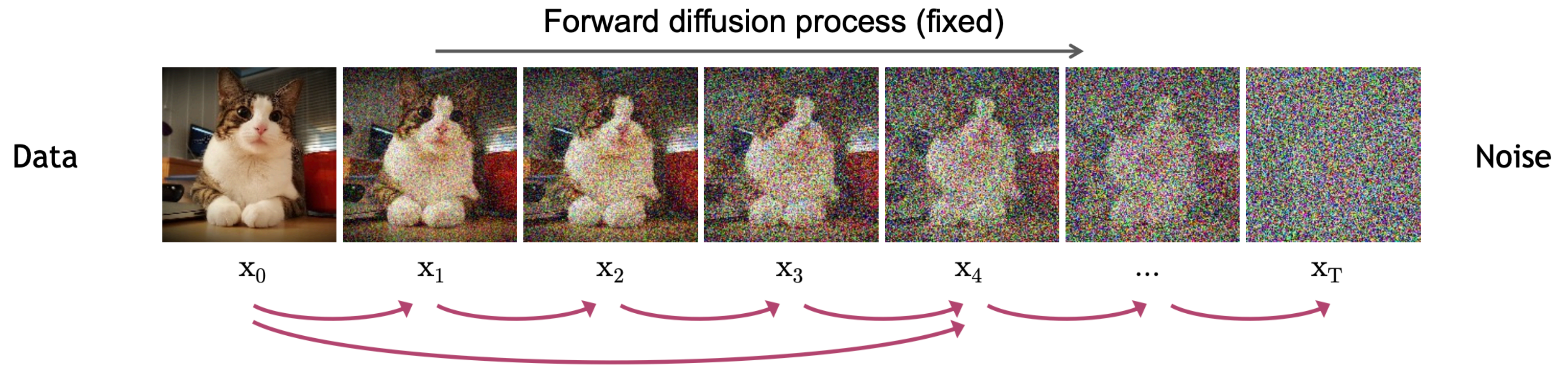
[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

## Marginal forward distribution

- Desirable during training to sample  $q(x_t)$  for any  $t$  in  $O(1)$  instead of  $O(T)$

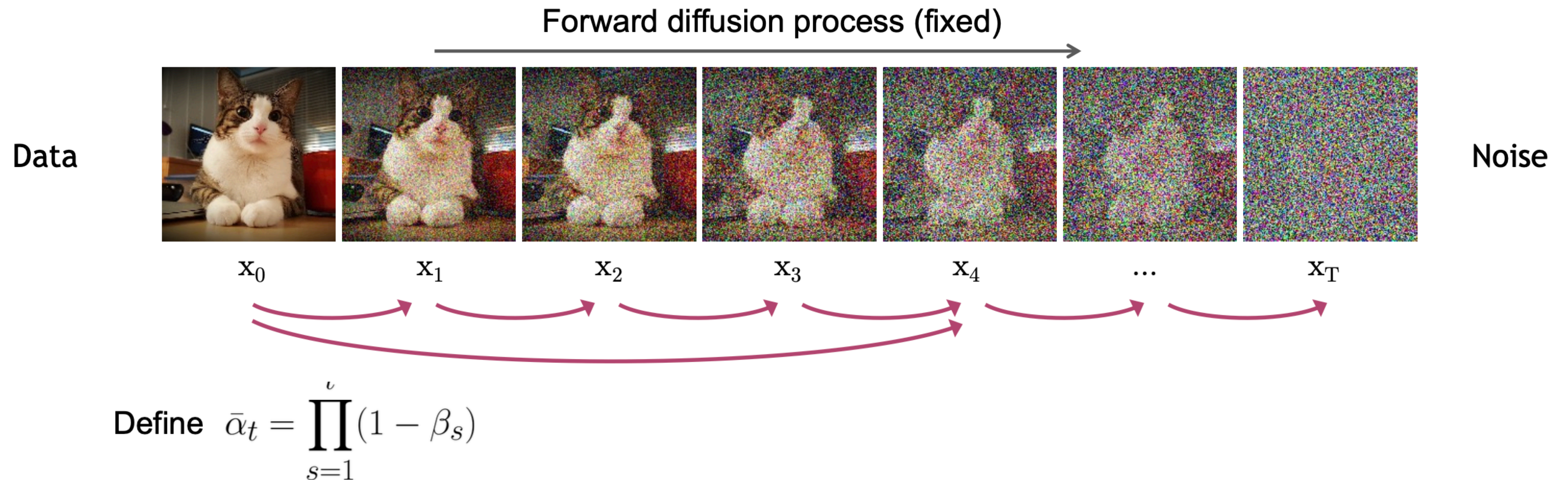




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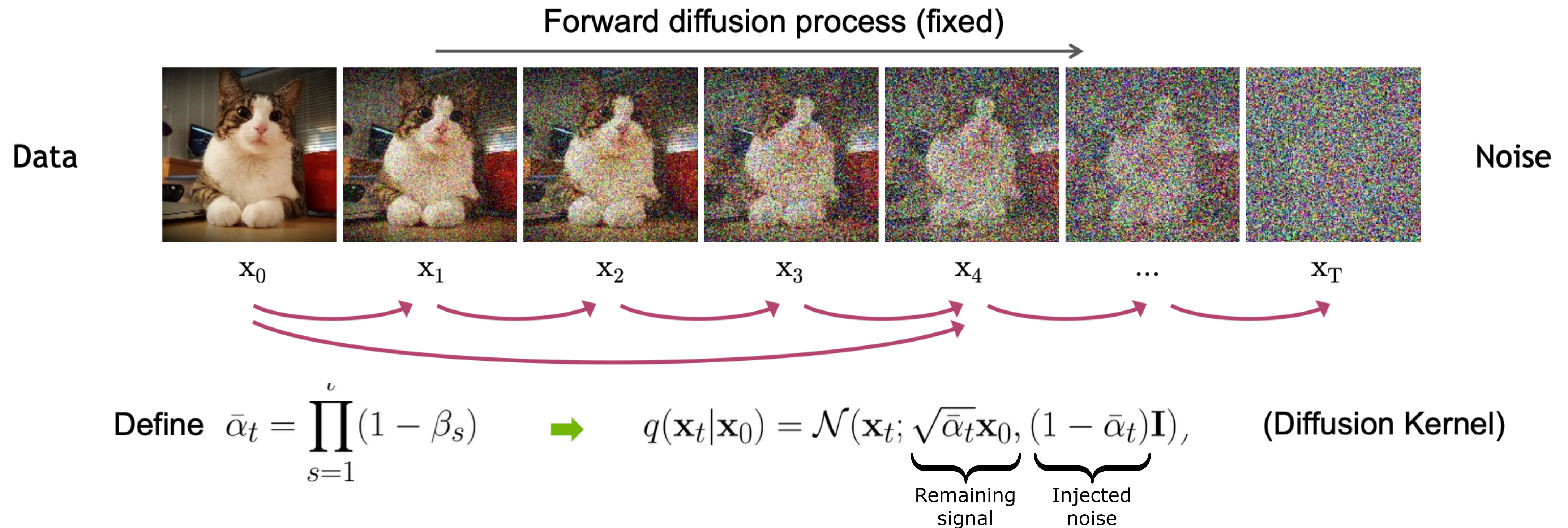




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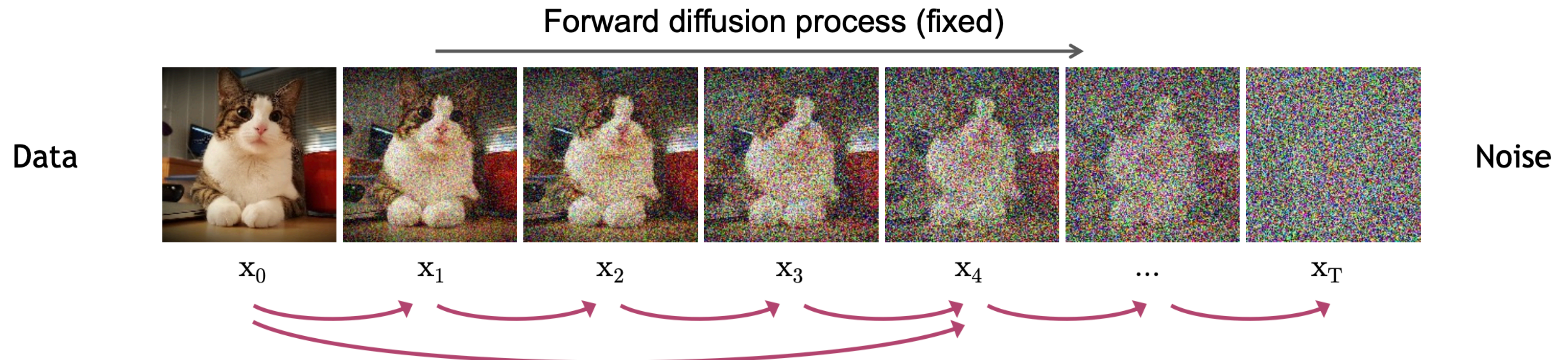




# Denoising diffusion probabilistic models

## Marginal forward distribution

- Desirable during training to sample  $q(x_t)$  for any  $t$  in  $O(1)$  instead of  $O(T)$



Define  $\bar{\alpha}_t = \prod_{s=1}^t (1 - \beta_s)$  ➔  $q(\mathbf{x}_t | \mathbf{x}_0) = \mathcal{N}(\mathbf{x}_t; \underbrace{\sqrt{\bar{\alpha}_t} \mathbf{x}_0}_{\text{Remaining signal}}, \underbrace{(1 - \bar{\alpha}_t) \mathbf{I}}_{\text{Injected noise}}),$  (Diffusion Kernel)

$$\underbrace{q(\mathbf{x}_t)}_{\text{Diffused data dist.}} = \int \underbrace{q(\mathbf{x}_0, \mathbf{x}_t)}_{\text{Joint dist.}} d\mathbf{x}_0 = \int \underbrace{q(\mathbf{x}_0)}_{\text{Input data dist.}} \underbrace{q(\mathbf{x}_t | \mathbf{x}_0)}_{\text{Diffusion kernel}} d\mathbf{x}_0$$

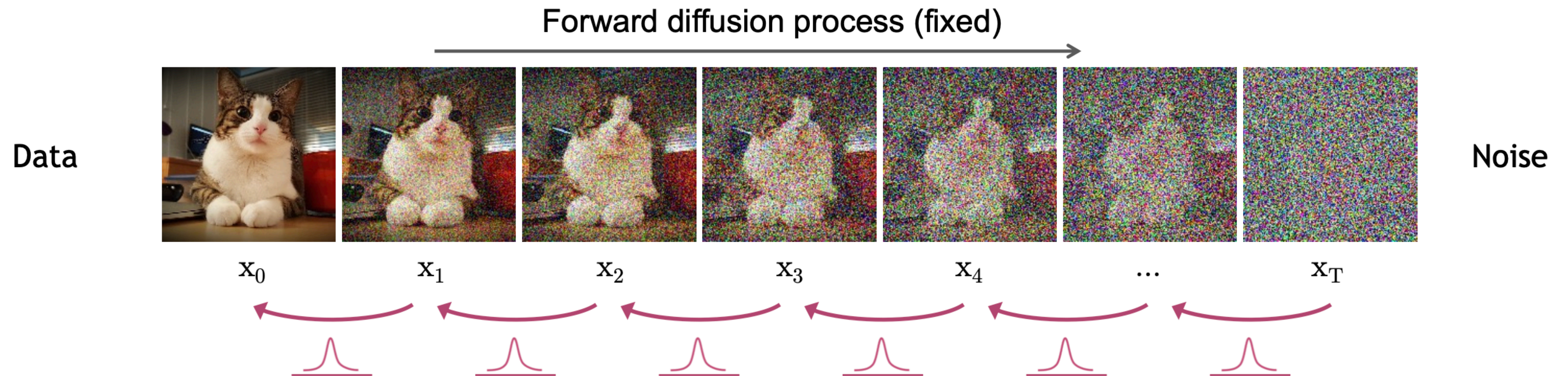
For sampling:  $\mathbf{x}_t = \sqrt{\bar{\alpha}_t} \mathbf{x}_0 + \sqrt{(1 - \bar{\alpha}_t)} \epsilon$  where  $\epsilon \sim \mathcal{N}(\mathbf{0}, \mathbf{I})$

[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

## Reverse denoising process

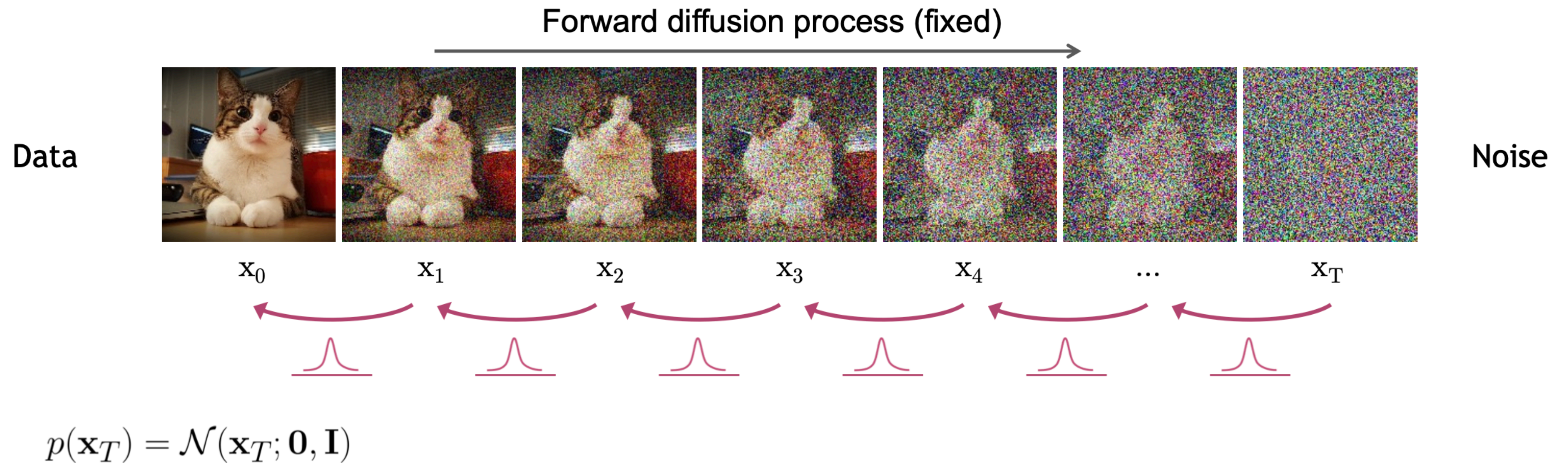


[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

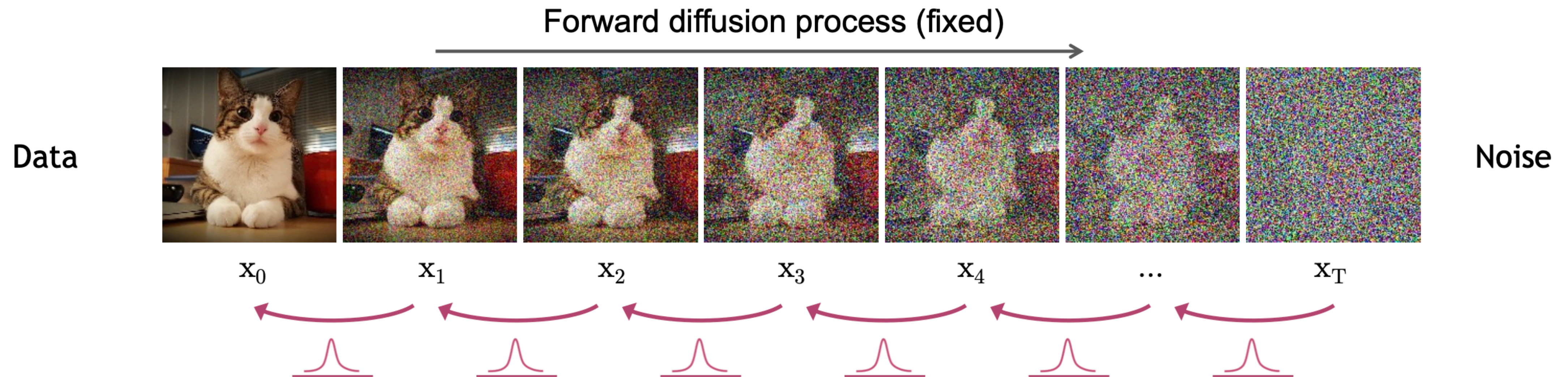
## Reverse denoising process





# Denoising diffusion probabilistic models

## Reverse denoising process



$$p(\mathbf{x}_T) = \mathcal{N}(\mathbf{x}_T; \mathbf{0}, \mathbf{I})$$

Goal:  $p_{\theta}(\mathbf{x}_{t-1}|\mathbf{x}_t) \approx q(\mathbf{x}_t|\mathbf{x}_{t-1})$

$$p_{\theta}(\mathbf{x}_{t-1}|\mathbf{x}_t) = \mathcal{N}(\mathbf{x}_{t-1}; \underbrace{\mu_{\theta}(\mathbf{x}_t, t)}_{\text{Trainable network}}, \sigma_t^2 \mathbf{I})$$

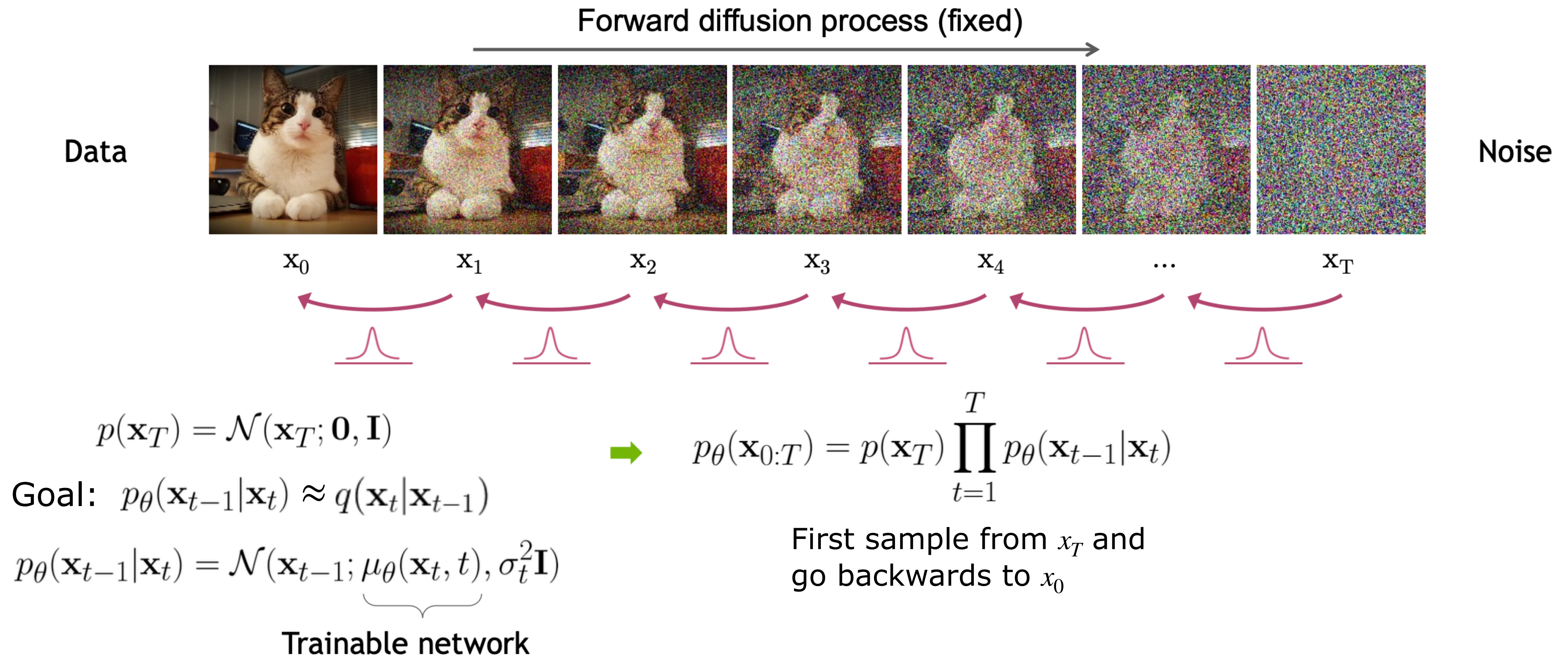
Trainable network

[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

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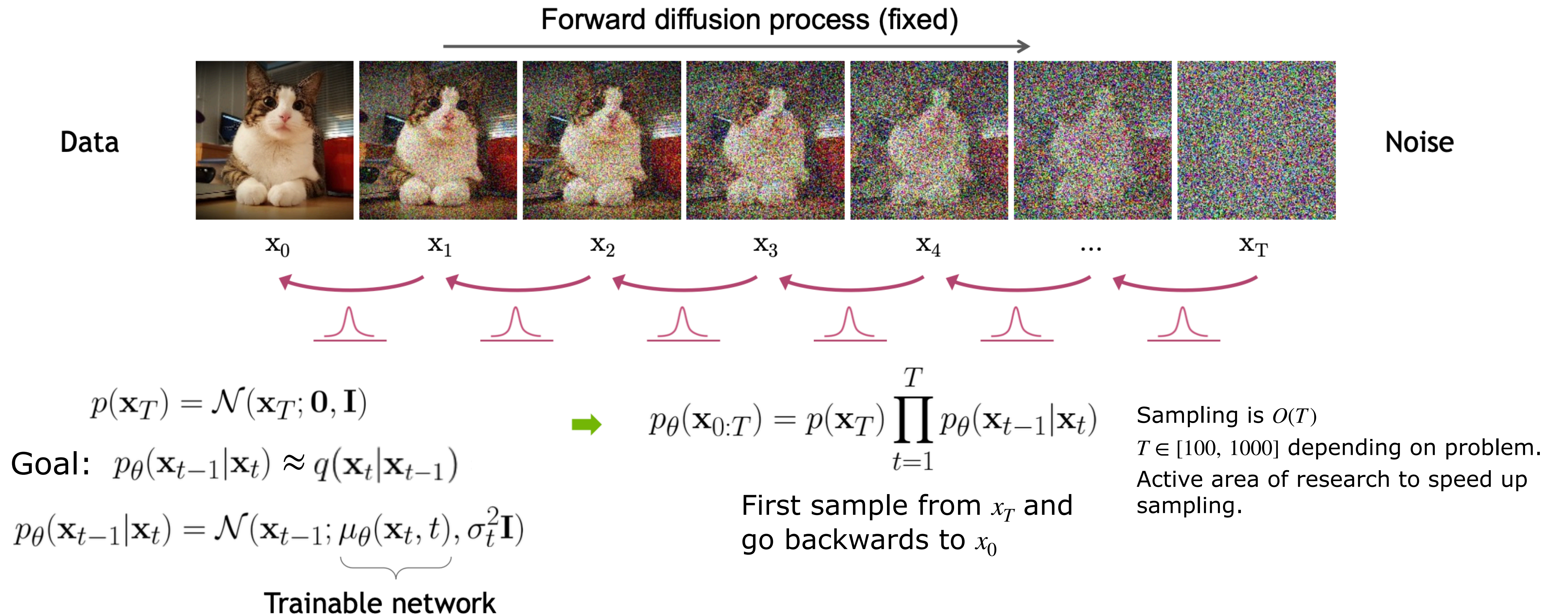


[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

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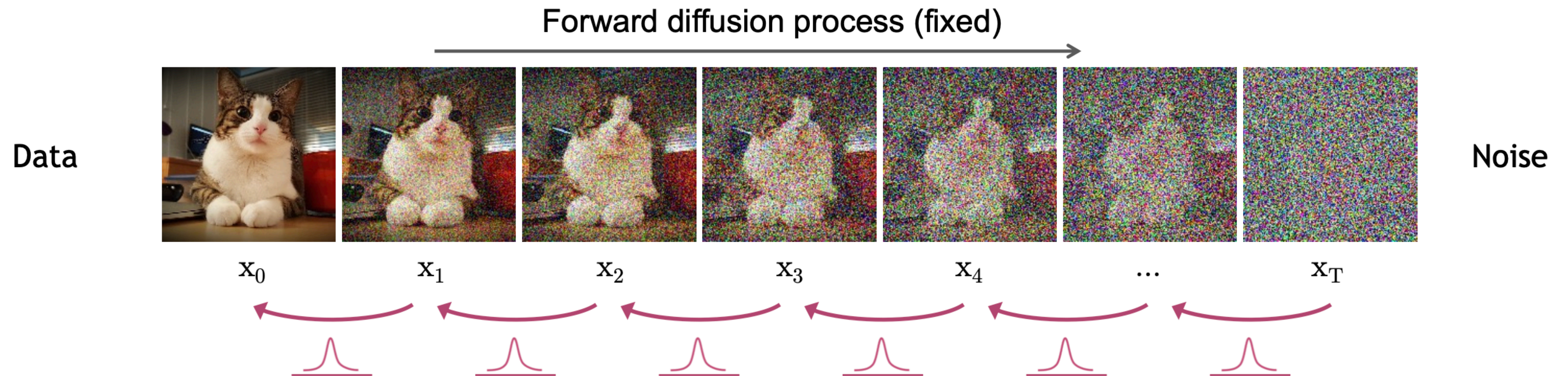


[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

## Optimization

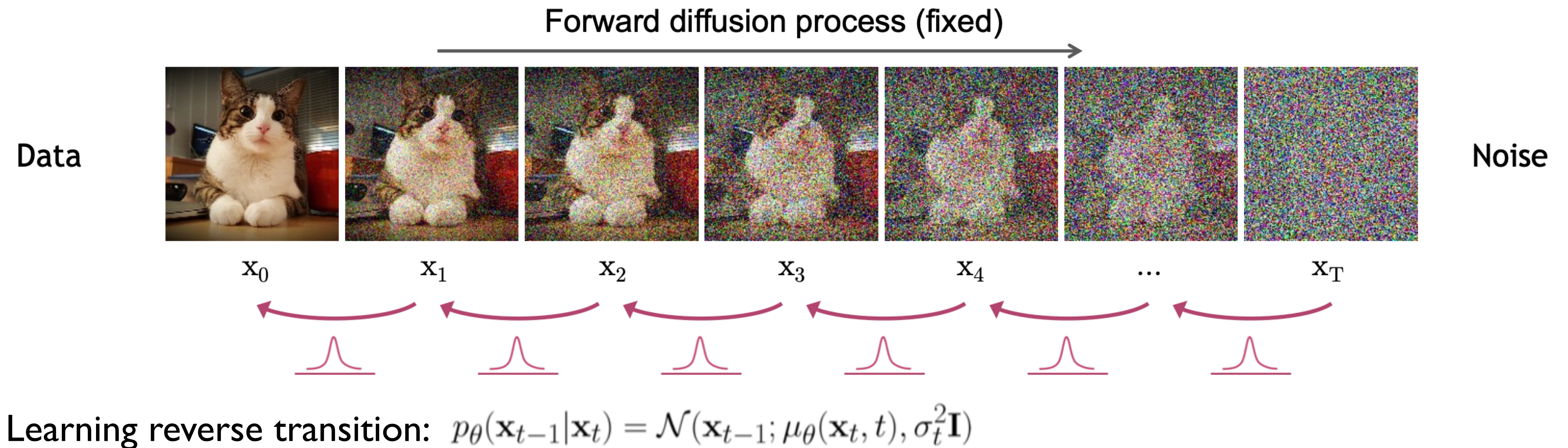


[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

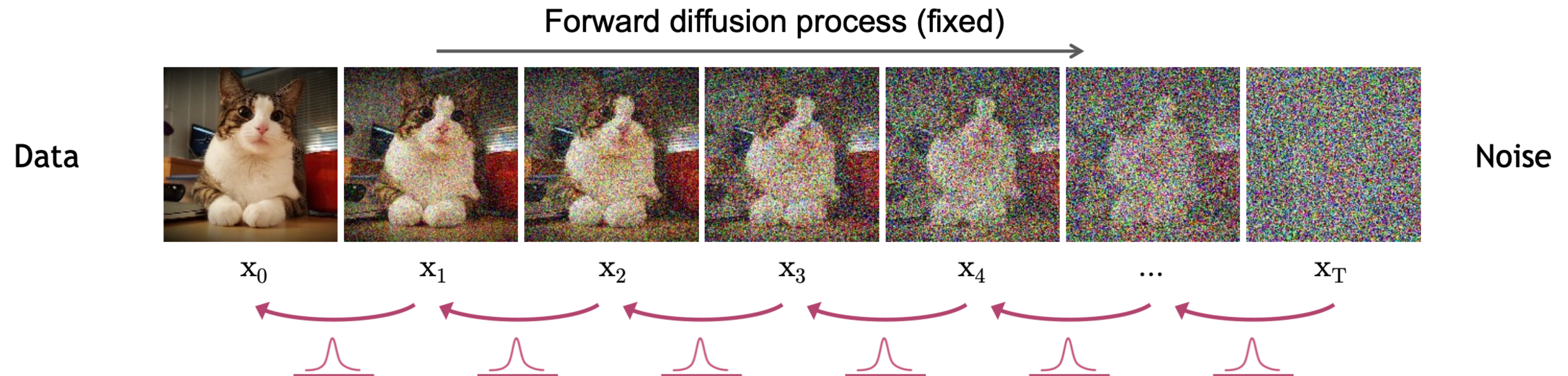
## Optimization





# Denoising diffusion probabilistic models

## Optimization



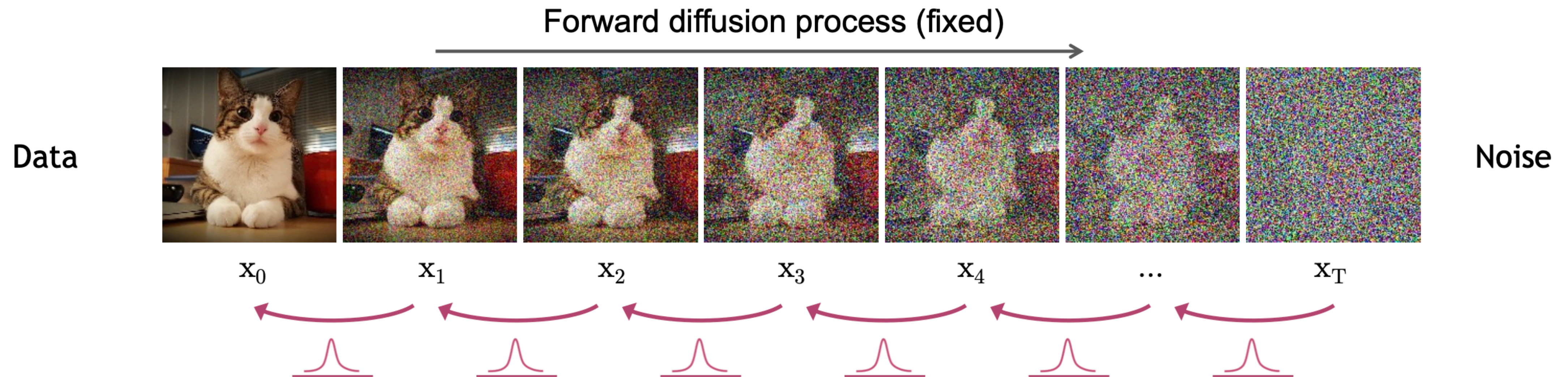
Learning reverse transition:  $p_{\theta}(\mathbf{x}_{t-1}|\mathbf{x}_t) = \mathcal{N}(\mathbf{x}_{t-1}; \mu_{\theta}(\mathbf{x}_t, t), \sigma_t^2 \mathbf{I})$

Recall  $\mathbf{x}_t = \sqrt{\bar{\alpha}_t} \mathbf{x}_0 + \sqrt{(1 - \bar{\alpha}_t)} \epsilon$



# Denoising diffusion probabilistic models

## Optimization



Learning reverse transition:  $p_{\theta}(\mathbf{x}_{t-1}|\mathbf{x}_t) = \mathcal{N}(\mathbf{x}_{t-1}; \mu_{\theta}(\mathbf{x}_t, t), \sigma_t^2 \mathbf{I})$

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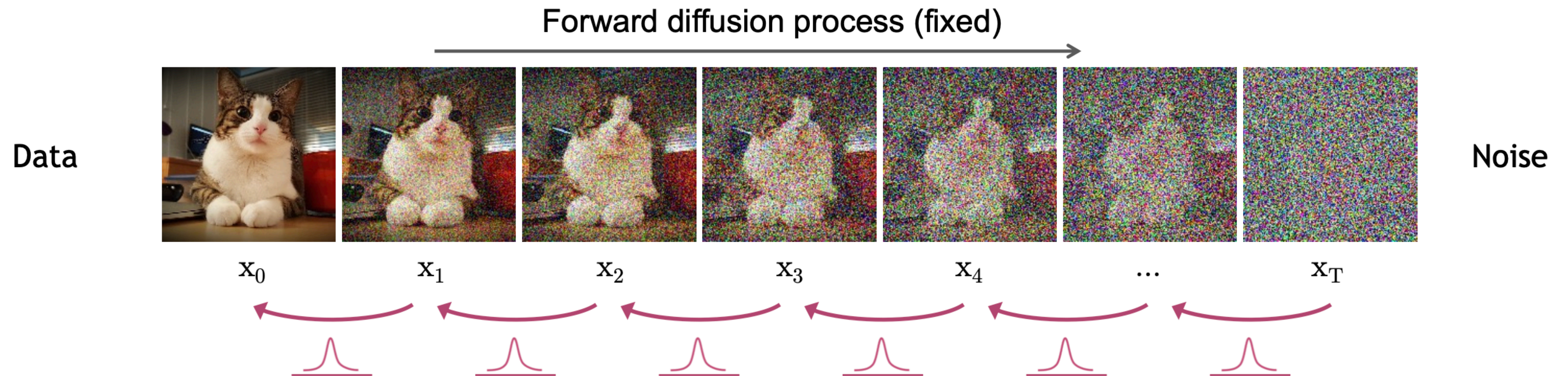
Related to  $x_0$ :  $\mu_{\theta}(\mathbf{x}_t, t) = \frac{1}{\sqrt{1 - \beta_t}} \left( \mathbf{x}_t - \frac{\beta_t}{\sqrt{1 - \bar{\alpha}_t}} \epsilon_{\theta}(\mathbf{x}_t, t) \right)$

[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

## Optimization



Learning reverse transition:  $p_{\theta}(\mathbf{x}_{t-1}|\mathbf{x}_t) = \mathcal{N}(\mathbf{x}_{t-1}; \mu_{\theta}(\mathbf{x}_t, t), \sigma_t^2 \mathbf{I})$

Reparameterization from [Ho et al]:  
 $L = \mathbb{E}[\|\epsilon - \epsilon_{\theta}\|^2], \quad \epsilon \sim \mathcal{N}(0, 1)$

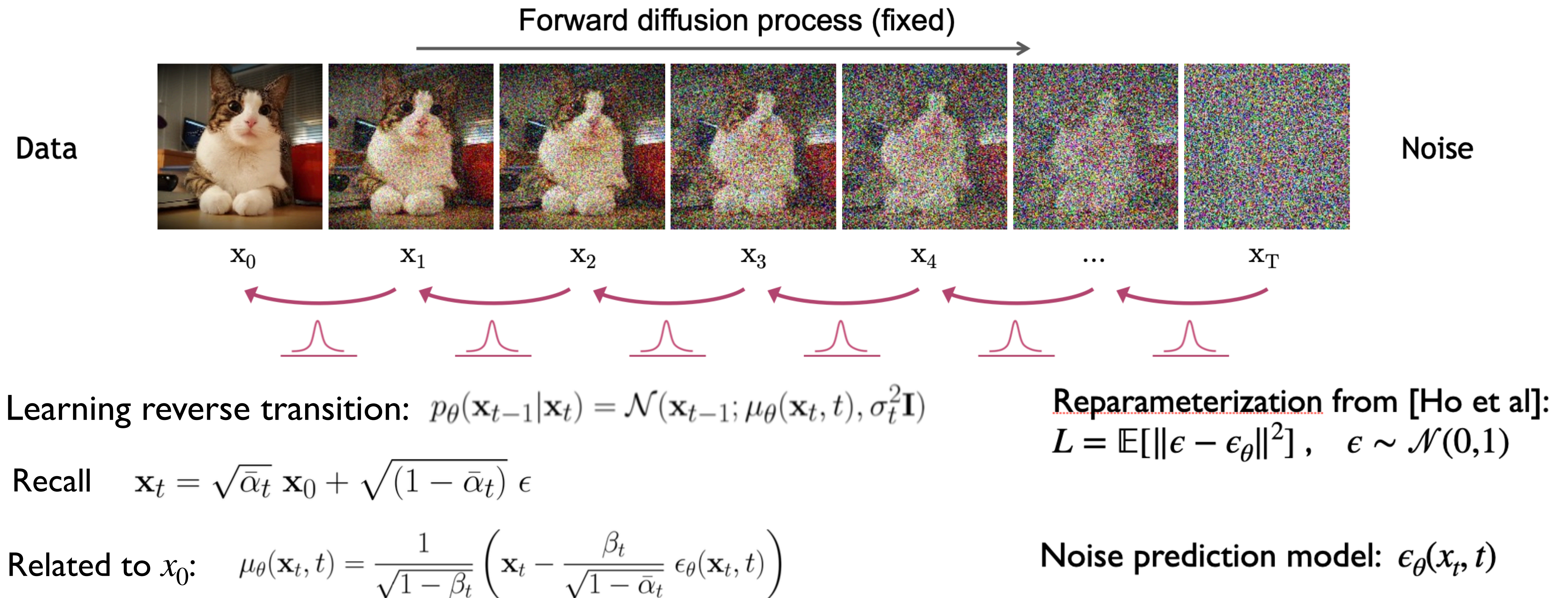
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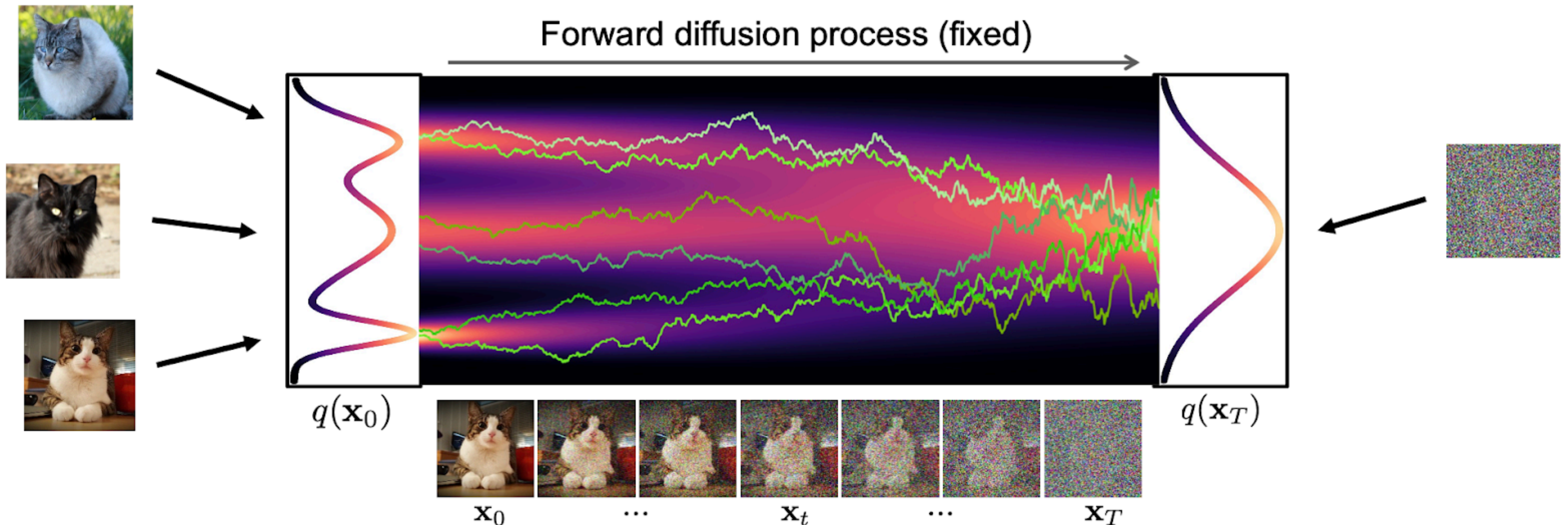
[Figure from CVPR tutorial]



# Denoising diffusion probabilistic models

## Intuition

- No encoder (replaced with forward diffusion) only need to train decoder (compared to VAEs).
- Decoder is easy to train (compared to GANs) with simple loss that are easy to learn/predict.



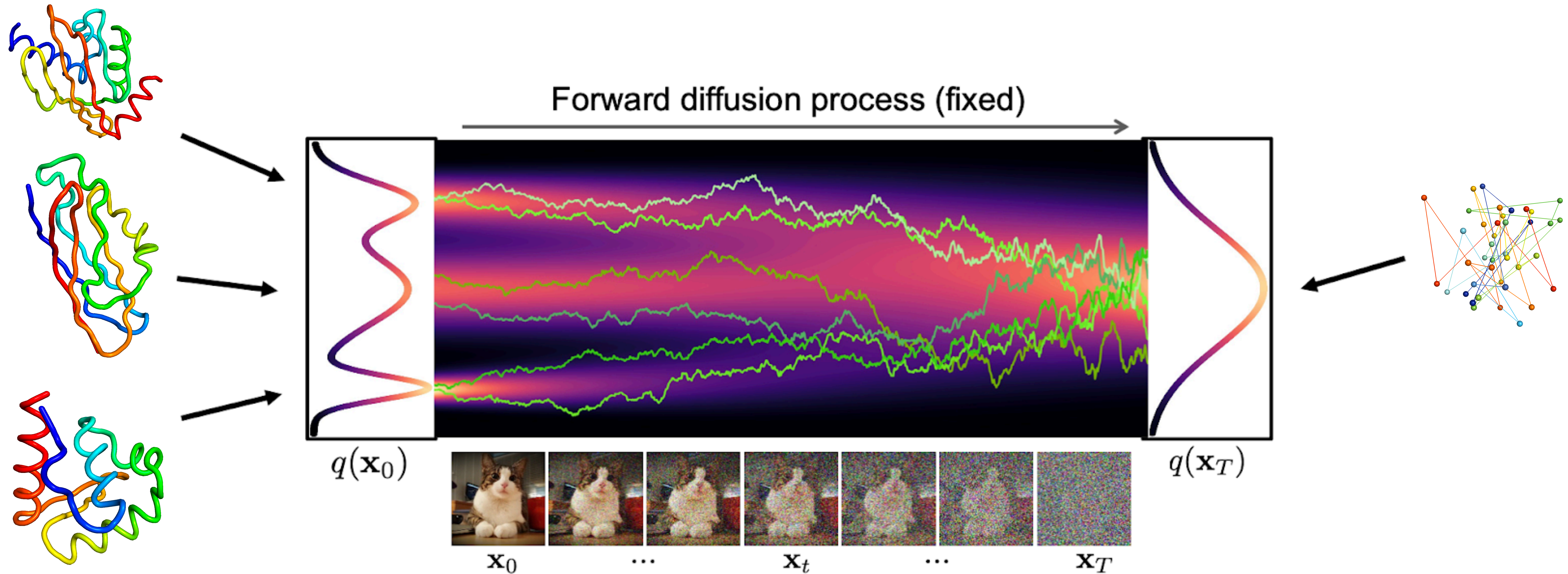
[Figure from CVPR tutorial]



# ProtDiff: diffusion on protein backbones

## Benefits of diffusion on proteins

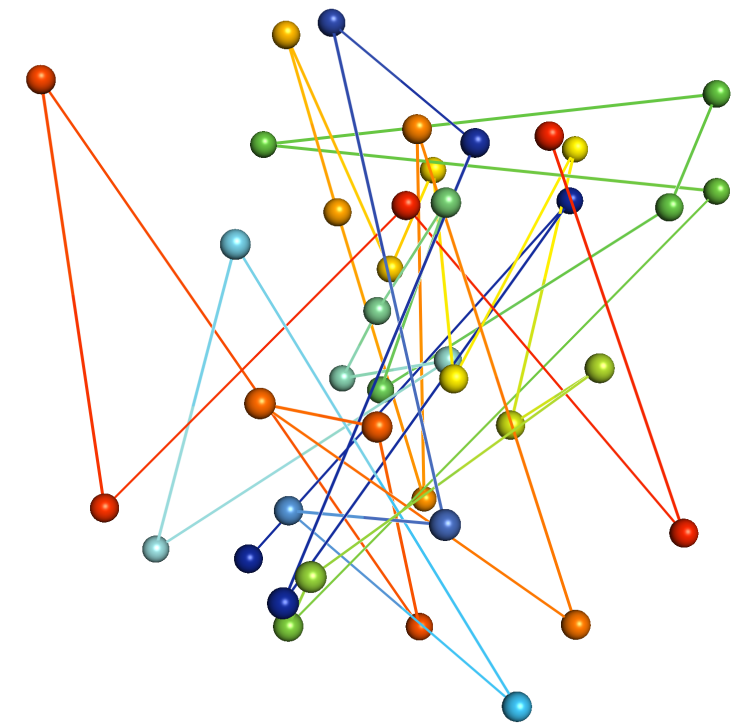
- Model directly in 3D space instead of distograms.
- Shown state-of-the-art success in generating small molecules [Hoogeboom et al.] and point clouds [Luo et al.]



[Figure from CVPR tutorial]



# ProtDiff: diffusion on protein backbones



random Gaussian  
point cloud

=

$x_T$

...

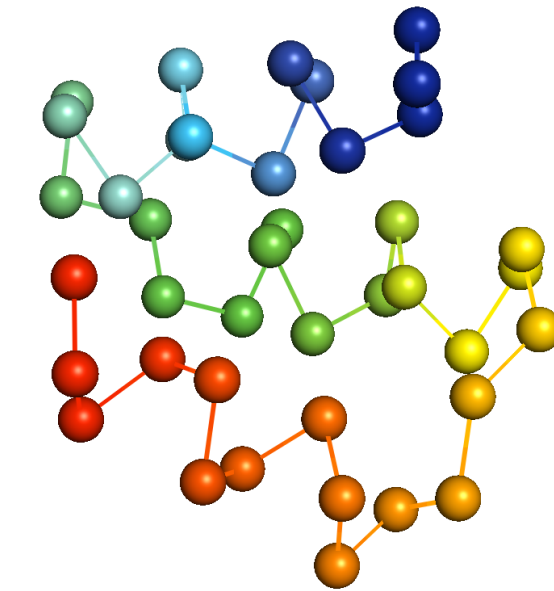
$x_t$

$x_{t-1}$

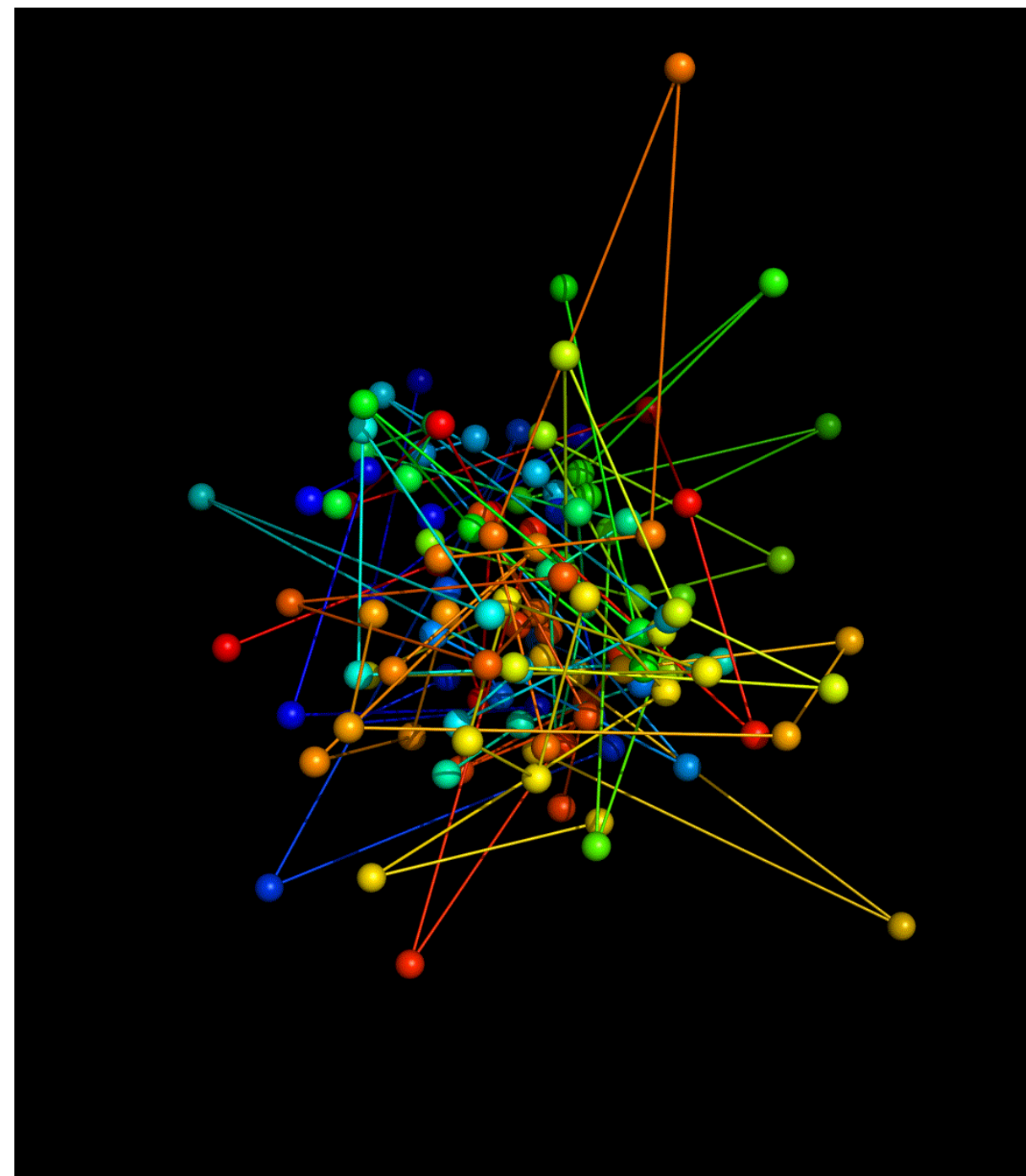
...

$x_0$

=



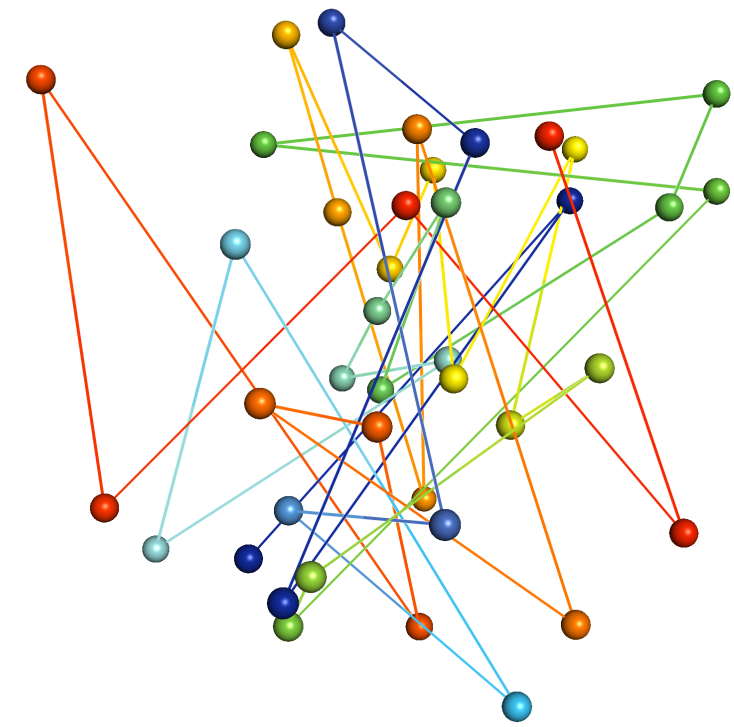
3D C $\alpha$  coordinates  
of a protein



## Future works:

- Incorporate SE(3) rigid bodies.
- Incorporate sequence.

# ProtDiff: diffusion on protein backbones



random Gaussian  
point cloud

=

$x_T$

...

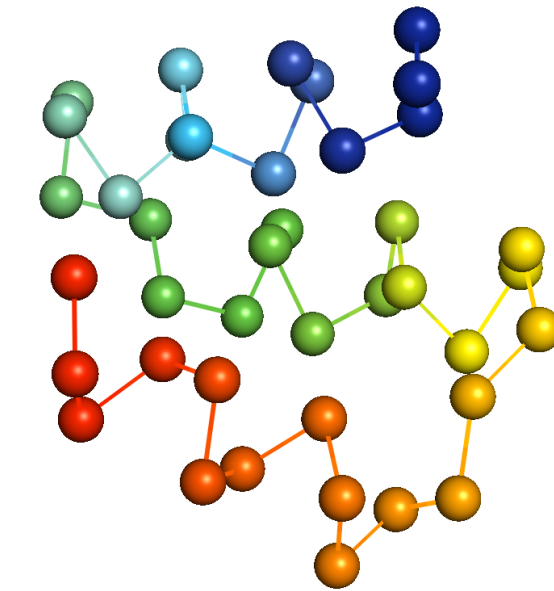
$x_t$

$x_{t-1}$

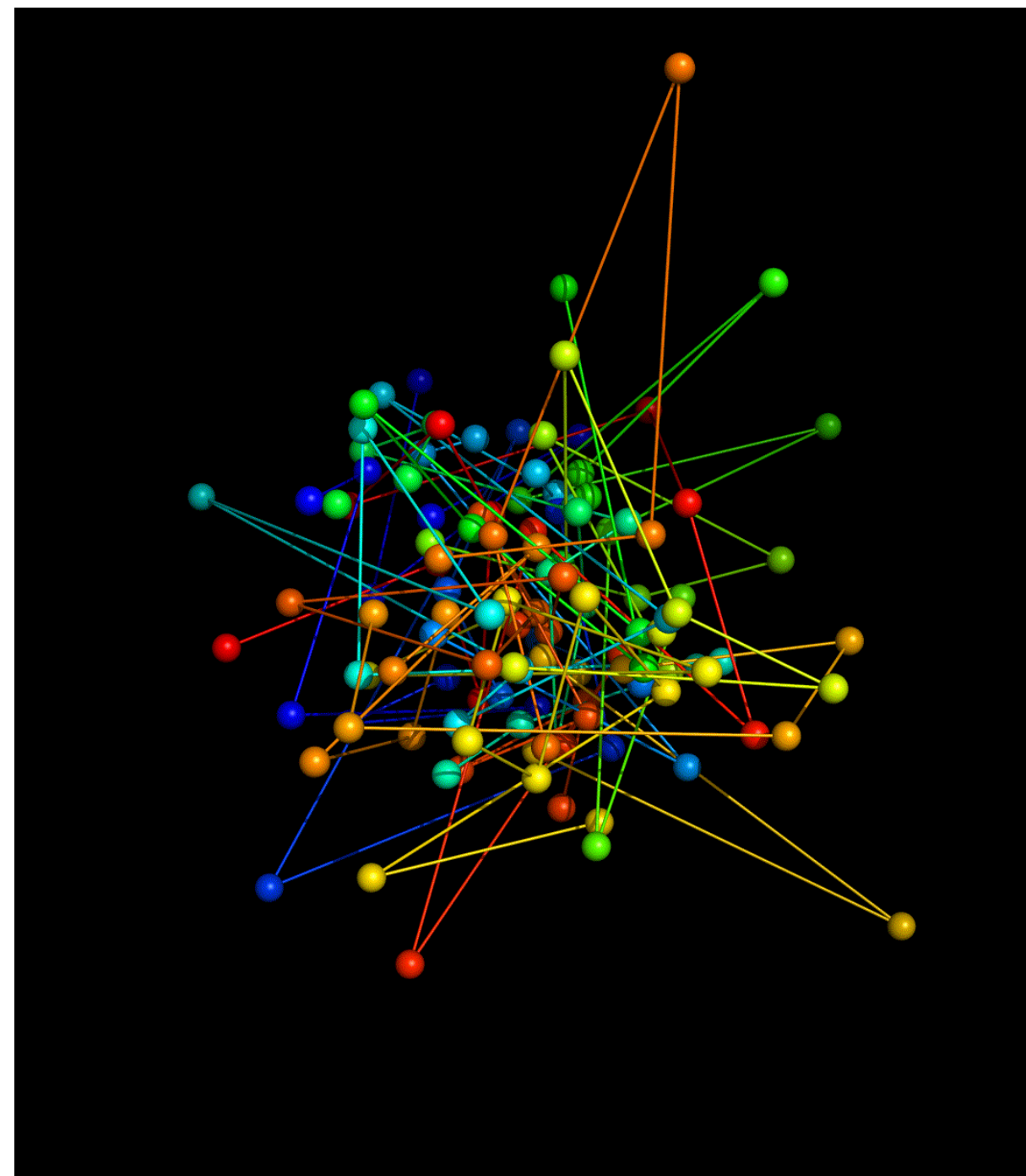
...

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=



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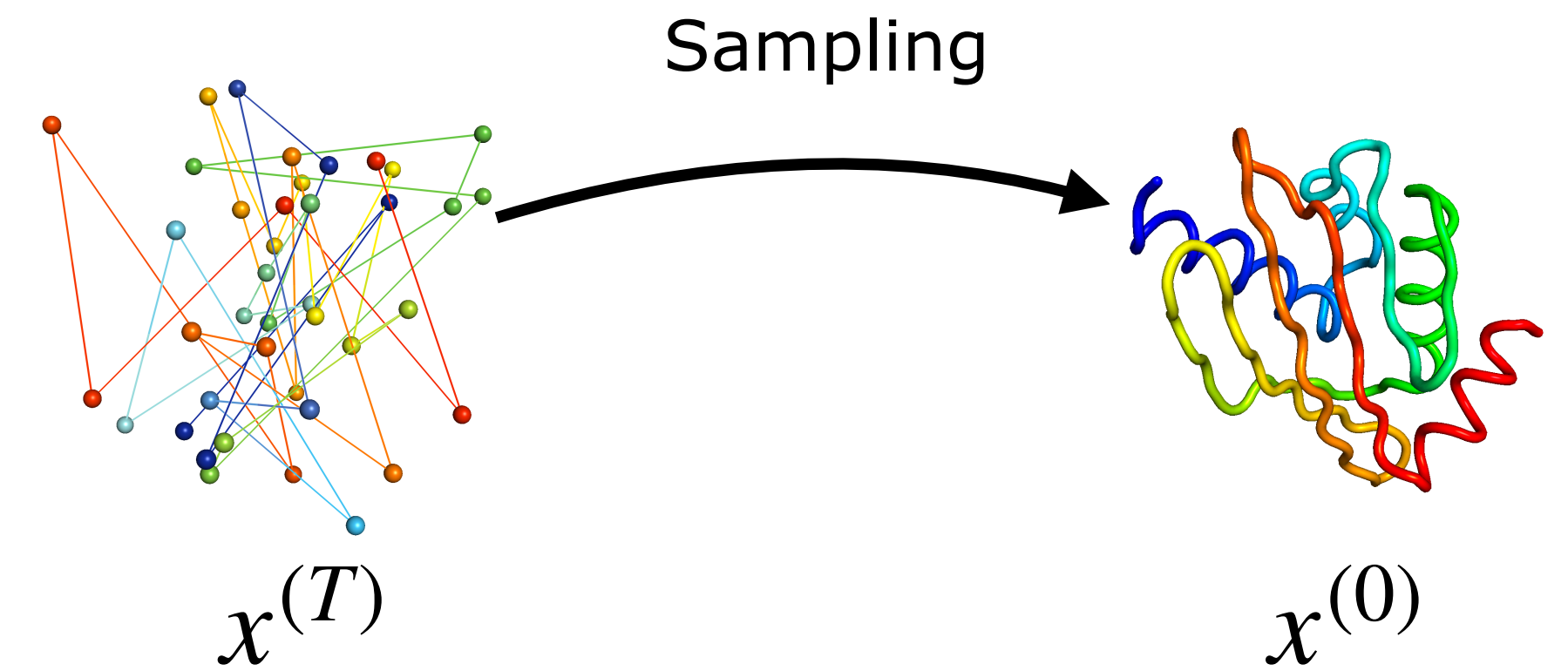
- Incorporate SE(3) rigid bodies.
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# Model details

▸ **Input:**

- $x_t \in \mathbb{R}^{N \times 3}$  : zero-centered 3D backbone point clouds.
  - Treats protein as a fully connected graph.
- $[s_1, \dots, s_N]$  where  $s_i$  is sequence position index of residue  $i$ 
  - Breaks permutation invariance.
- $t \in [1, \dots, T]$





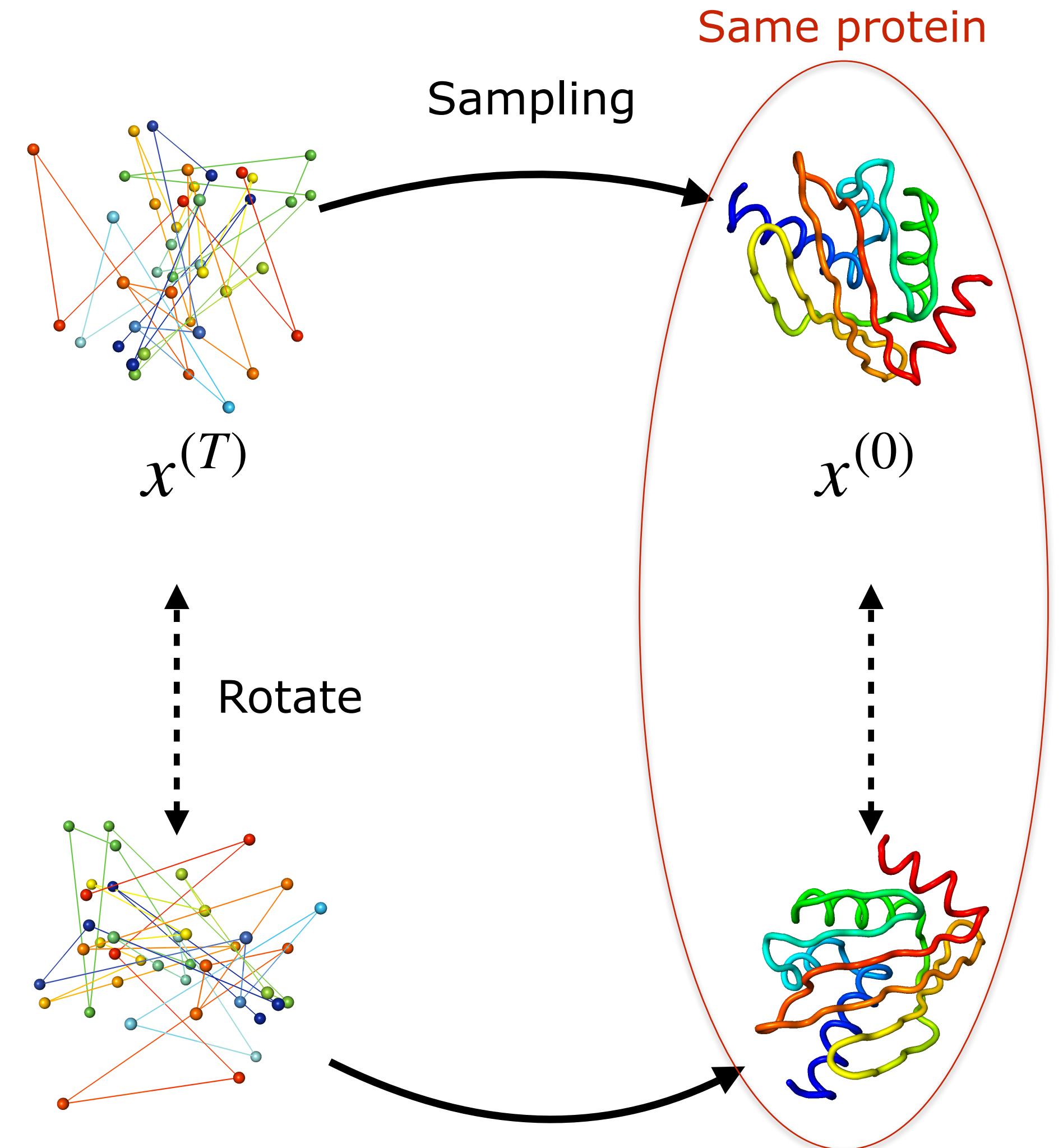
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## E(3) Equivariant diffusion model:

- Goal:  $p(x_0) = p(Rx_0)$ ,  $R \in SO(3)$
- Starting from **invariant** distribution with a **equivariant** noise prediction model leads to **equivariant** samples. [GeoDiff Xu et al.]
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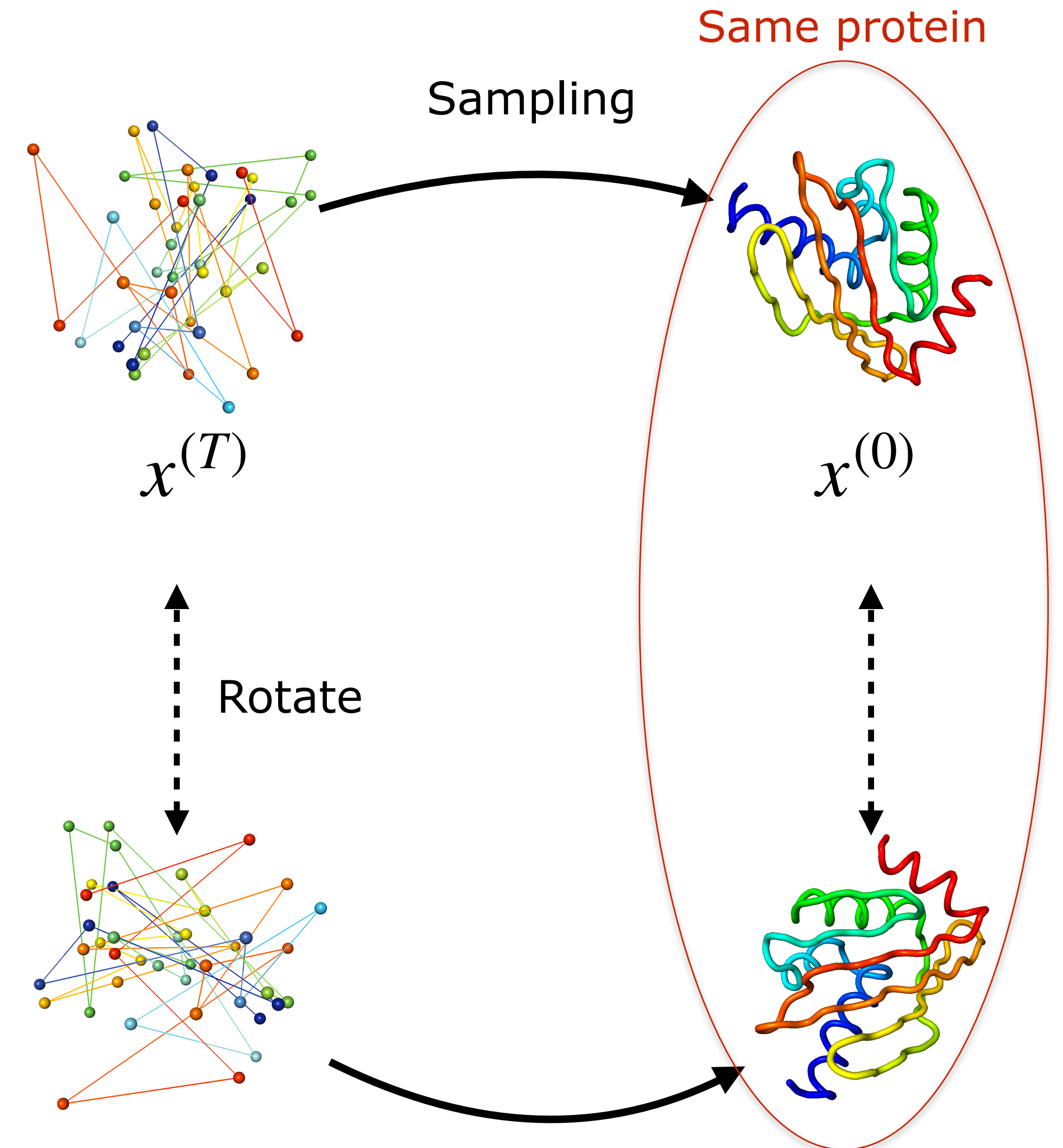
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## Neural network:

- E(3) equivariant graph neural network (EGNN) [Satorras et al.]
- Property that Euclidean transforms equally affect output.  
 $R \epsilon_\theta(x) = \epsilon_\theta(Rx) \rightarrow$  equivariant noise prediction

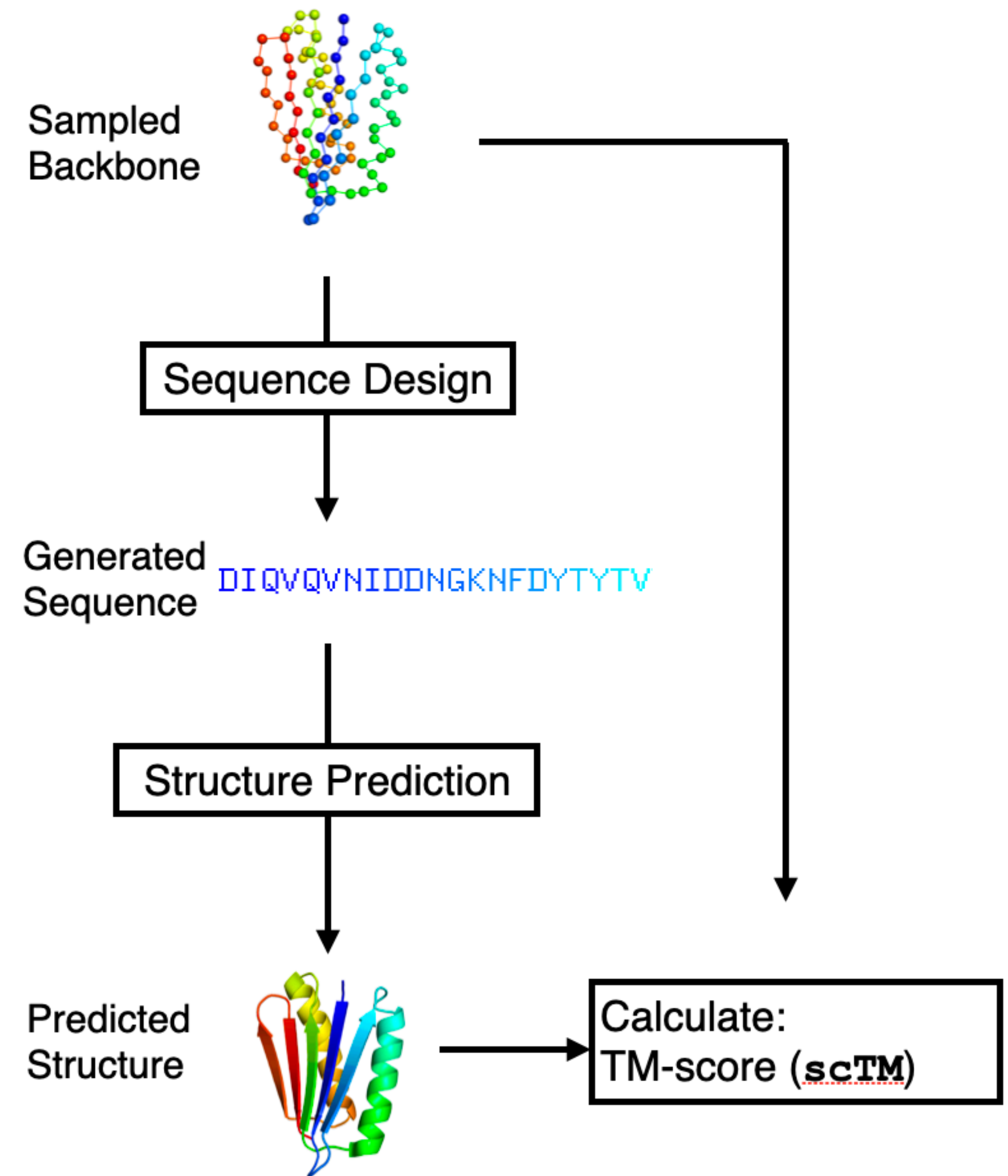




# Designability criterion

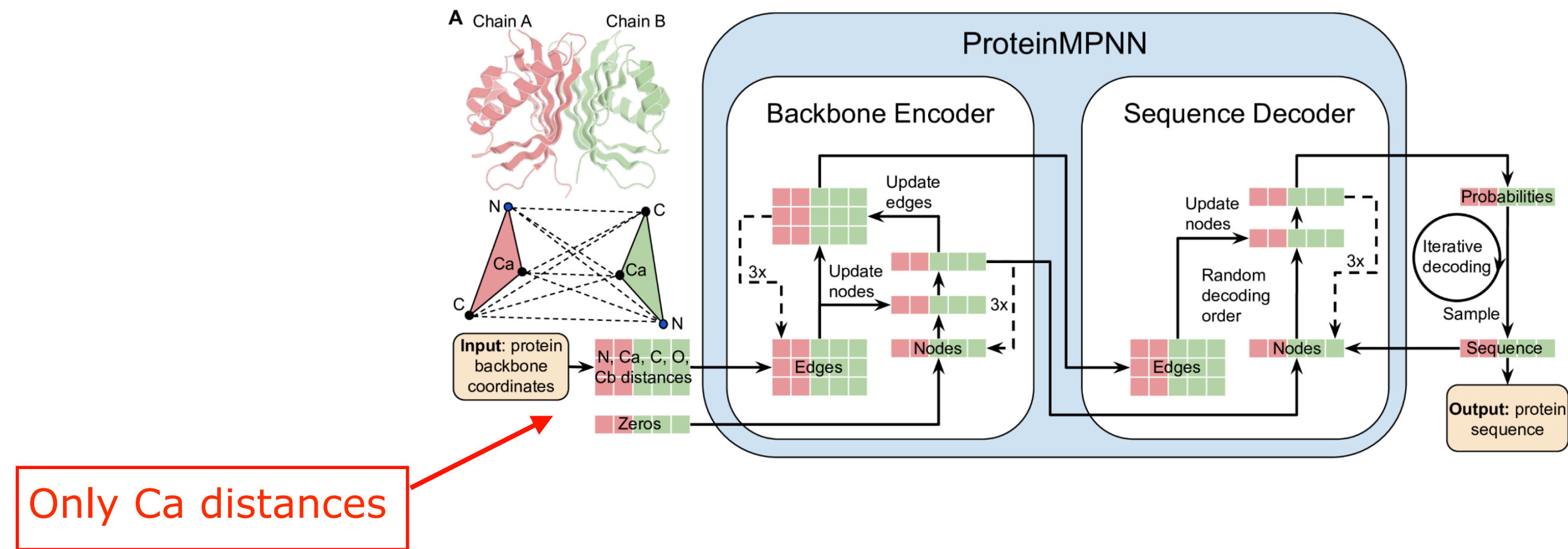
- A 3D structure is **designable** if a protein sequence can be found that folds into the same 3D structure.
- Use **sequence design** to search for likely sequences for a backbone.
- Use **structure prediction** (folding) to verify backbone samples can be designed from a protein sequence.
  - >0.5 TM-score indicates roughly same structure.

## Self-consistency evaluation



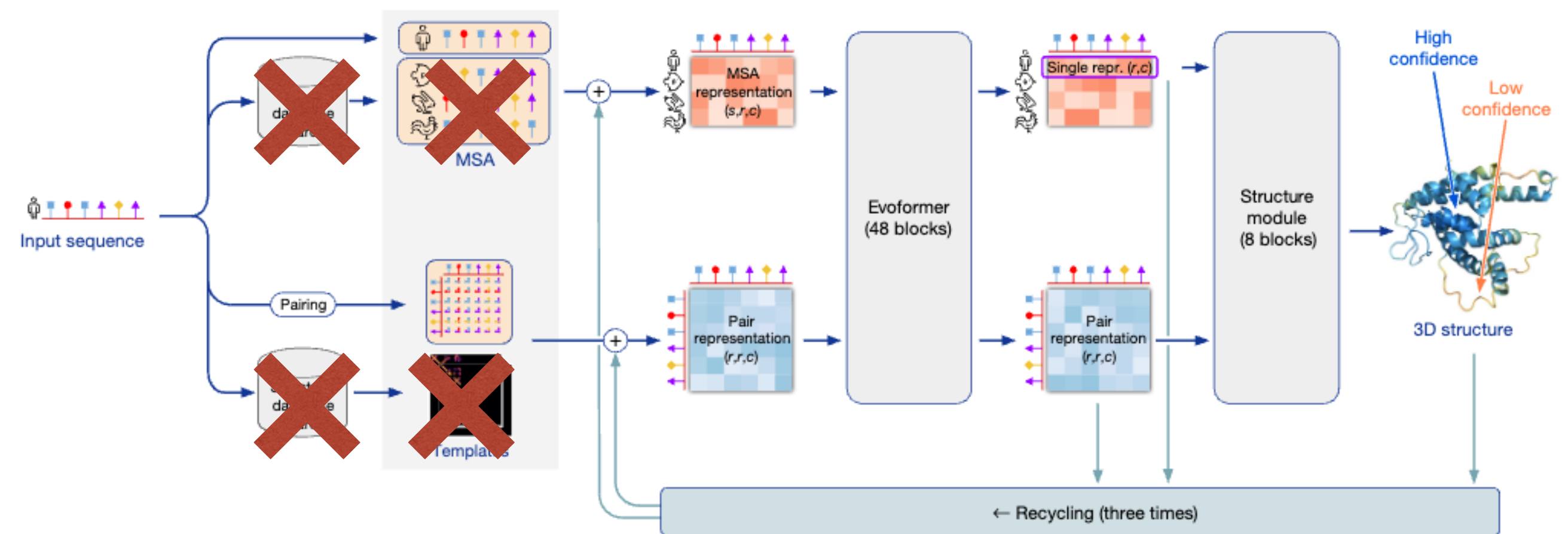
# Self-consistency components

- Sequence design: **ProteinMPNN**
  - State-of-the-art fixed backbone sequence design method.



[Dauparas et al]

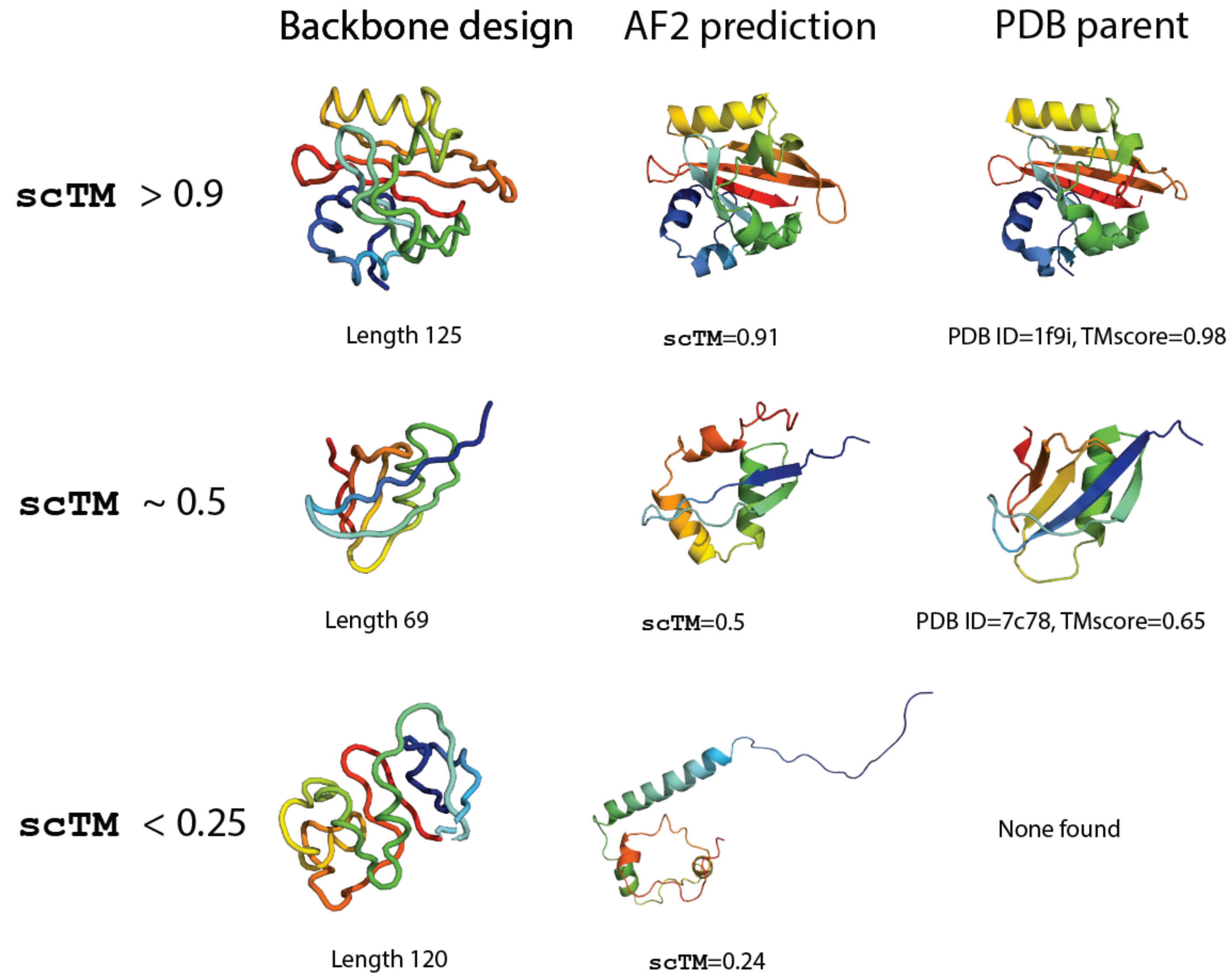
- Structure prediction: **AlphaFold2**
  - No MSA. No template. Allows for fast inference. Only include query sequence.
  - Use released CASP14 weights.



[Jumper et al]



# Characterization of self-consistency



# Unconditional sampling



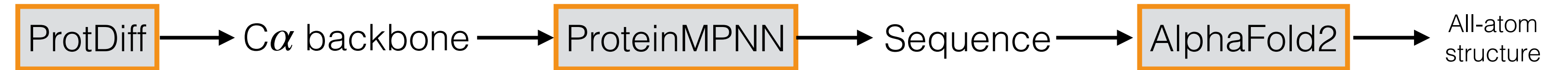
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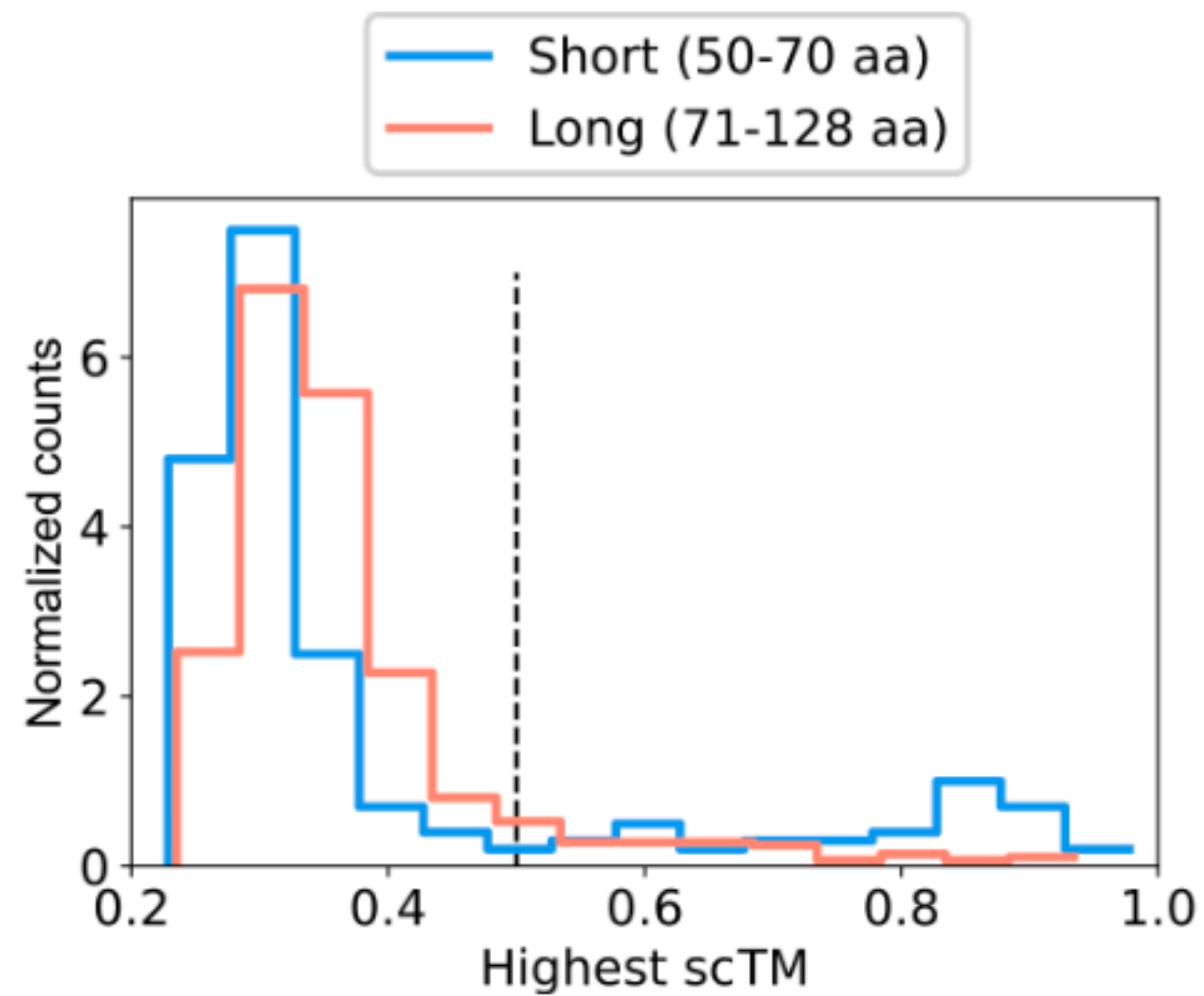
- Full sampling pipeline:





# Unconditional sampling

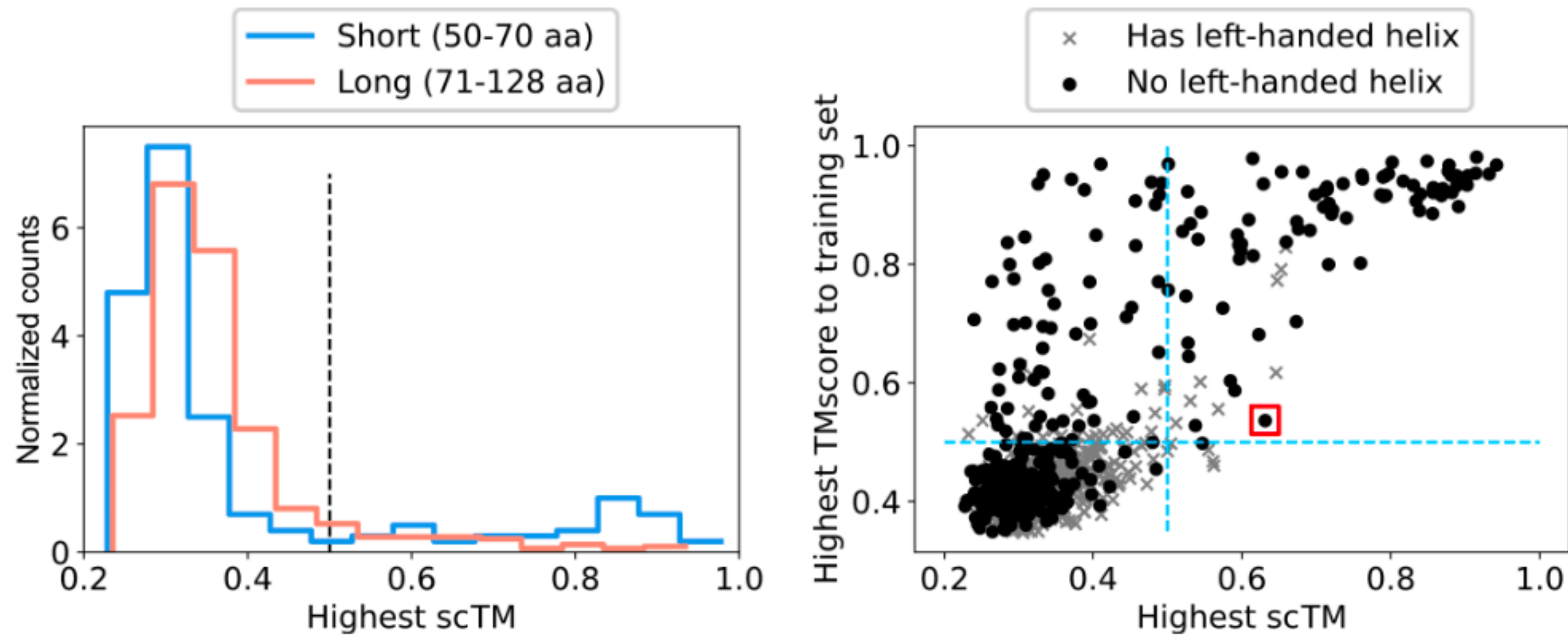
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- Compare the end structures through alignment (scTM)



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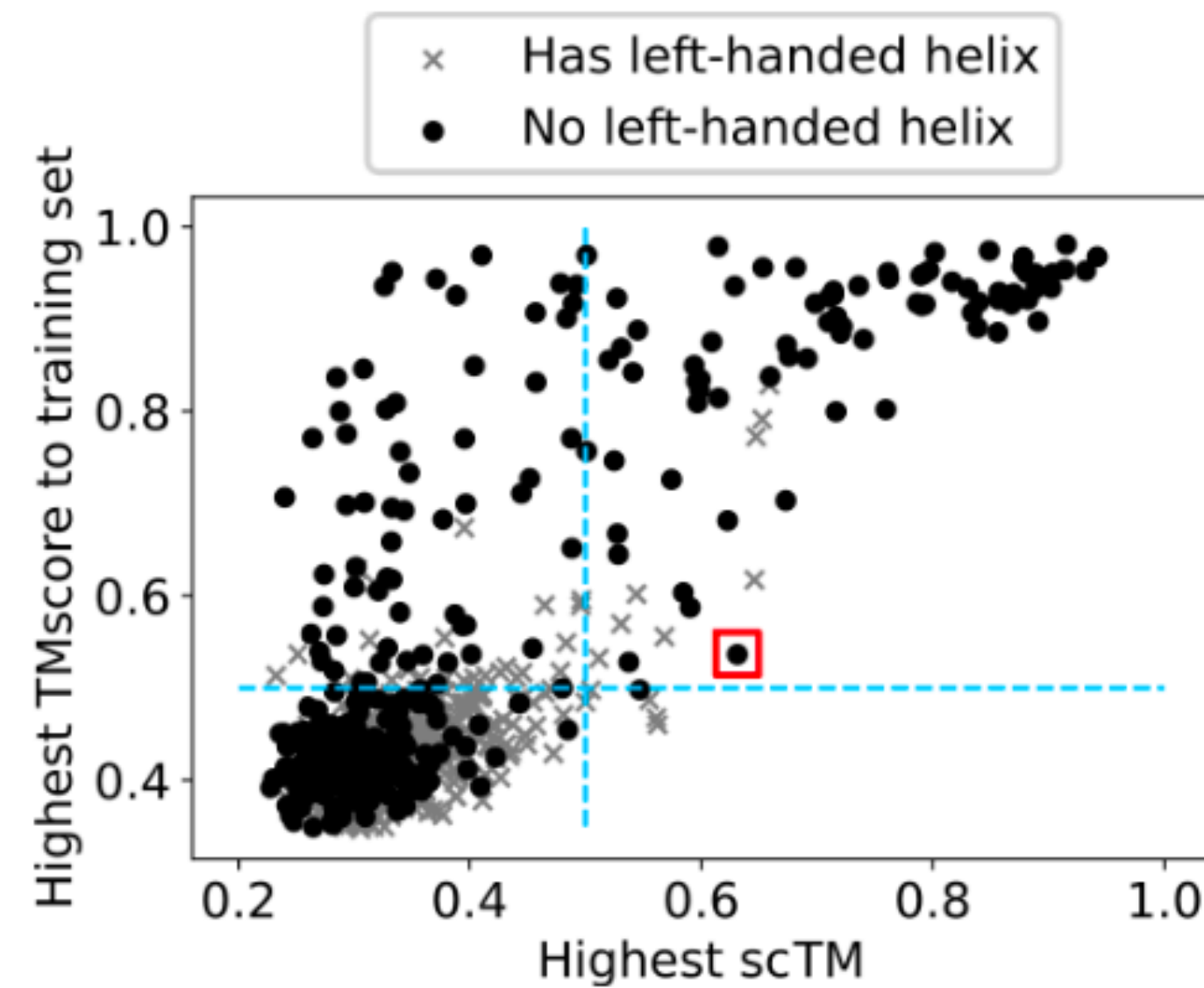
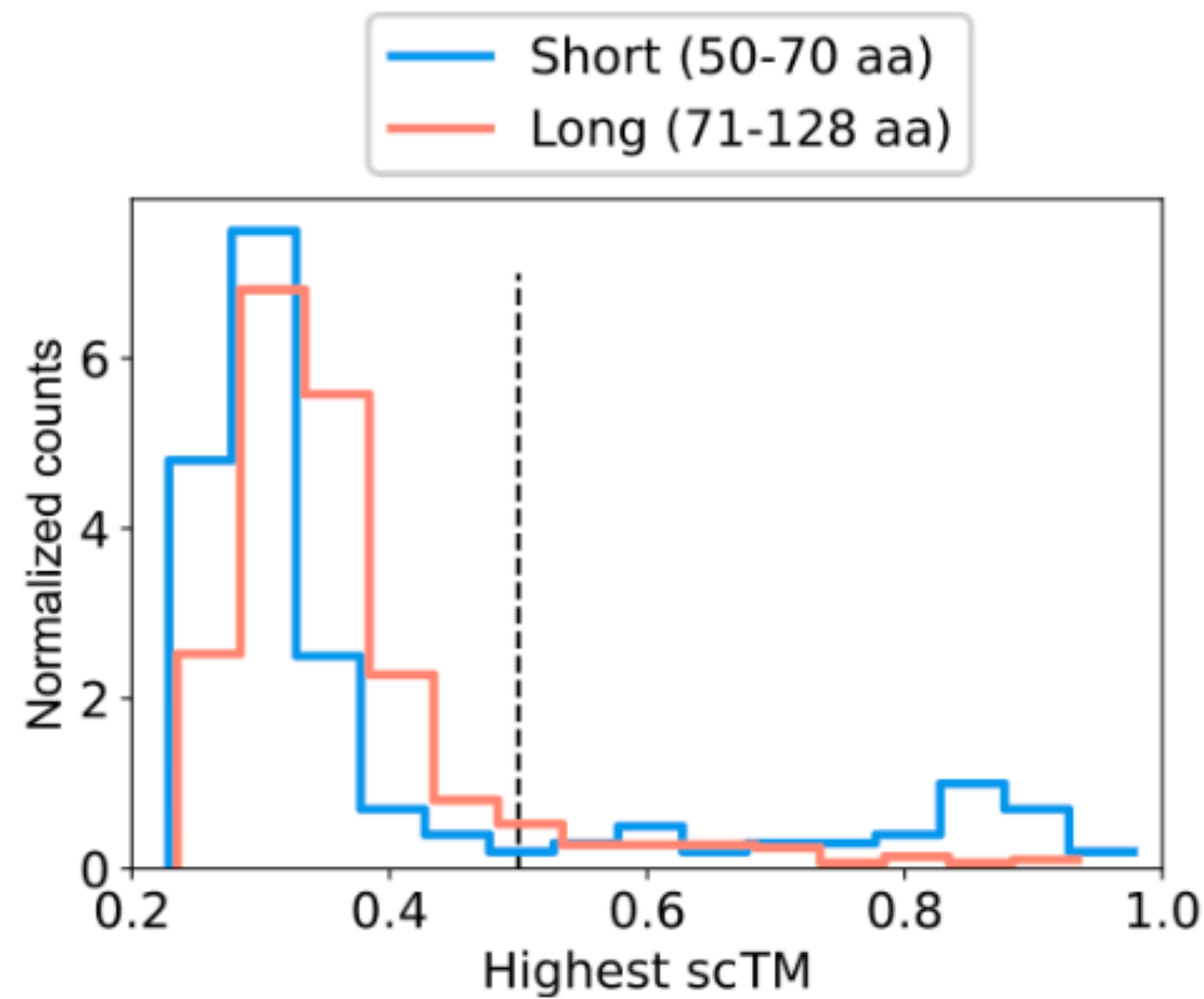


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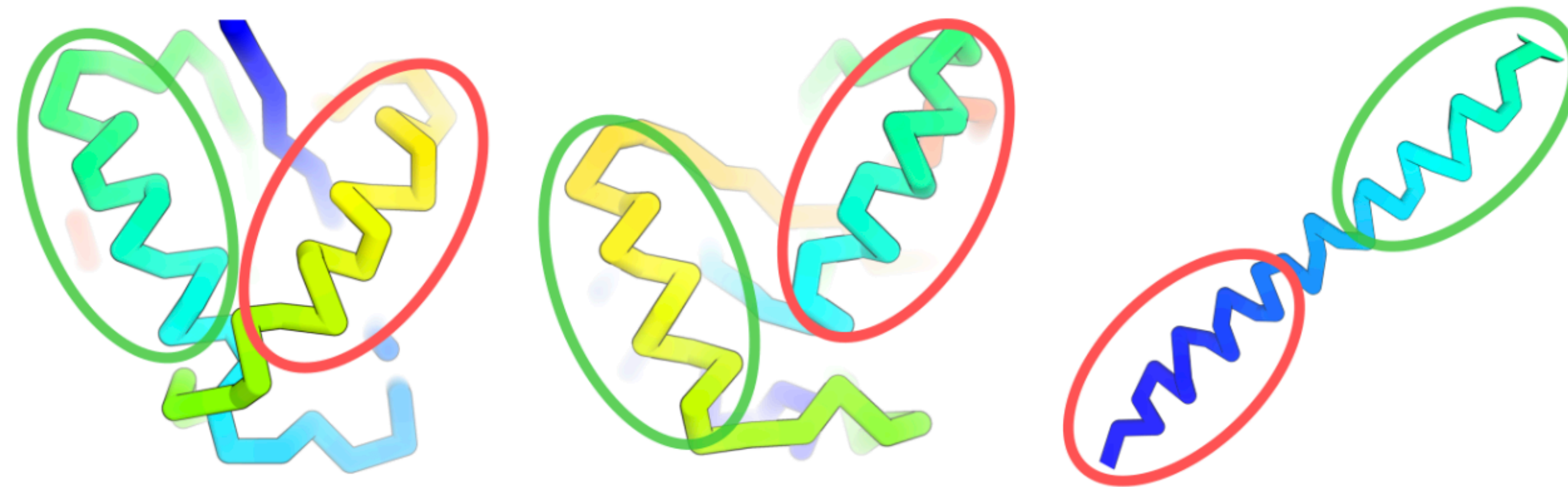
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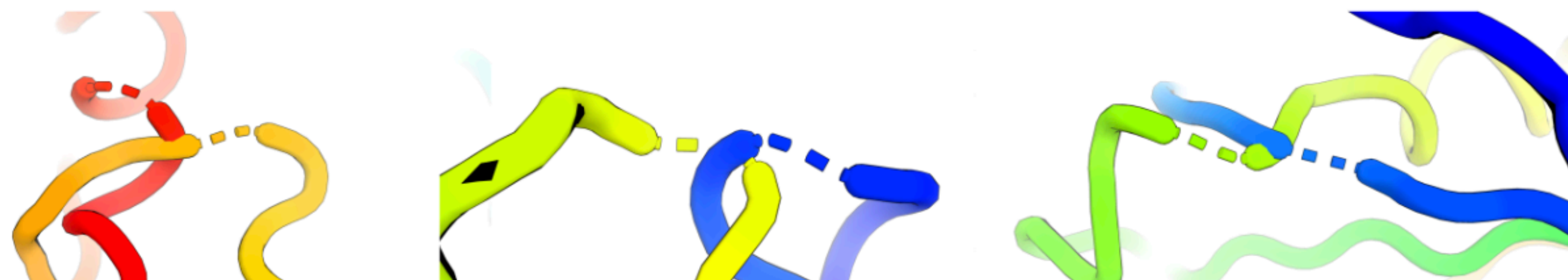
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# Specific failure mode

- The model can generate left-handed helices (in red)



- Chain breaks can occur





# Sample interpolations

- Noise interpolated samples



For samples with noise  $\epsilon^{(0:T)}$  and  $\tilde{\epsilon}^{(0:T)}$  we interpolate with noise set to  $\sqrt{\alpha}\epsilon^{(0:T)} + \sqrt{1-\alpha}\tilde{\epsilon}^{(0:T)}$

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# Roadmap

## Learning $p_{\theta}(x)$ [ProtDiff]

- Diffusion generative modeling background
- Adapting diffusion for protein backbones
- Model performance and limitations

## Sampling $x_S \sim p_{\theta}(x_S | x_M)$ [SMCDiff]

- Why conditional sampling vs. “naive” in-painting?
- Sequential Monte Carlo for exact sampling in the large-compute limit

## Limitations, Related Work, and Future directions

# Inpainting with Diffusion Models



“A girl hugging a corgi on a pedestal”



# Inpainting with Diffusion Models



“A girl hugging a corgi on a pedestal”

- Artifacts & edge effects in state-of-art inpainting





# Inpainting with Diffusion Models



“A girl hugging a corgi on a pedestal”

- ▶ For protein design, small errors break designability

- Artifacts & edge effects in state-of-art inpainting





# Inpainting with Diffusion Models



“A girl hugging a corgi on a pedestal”

- ▶ For protein design, small errors break designability
- ▶ Why does this work at all?

- Artifacts & edge effects in state-of-art inpainting





# Inpainting with Diffusion Models



“A girl hugging a corgi on a pedestal”

- For protein design, small errors break designability
- Why does this work at all?

**Hypothesis:** Inpainting approximates conditional sampling.  
Approximation error —> artifacts.

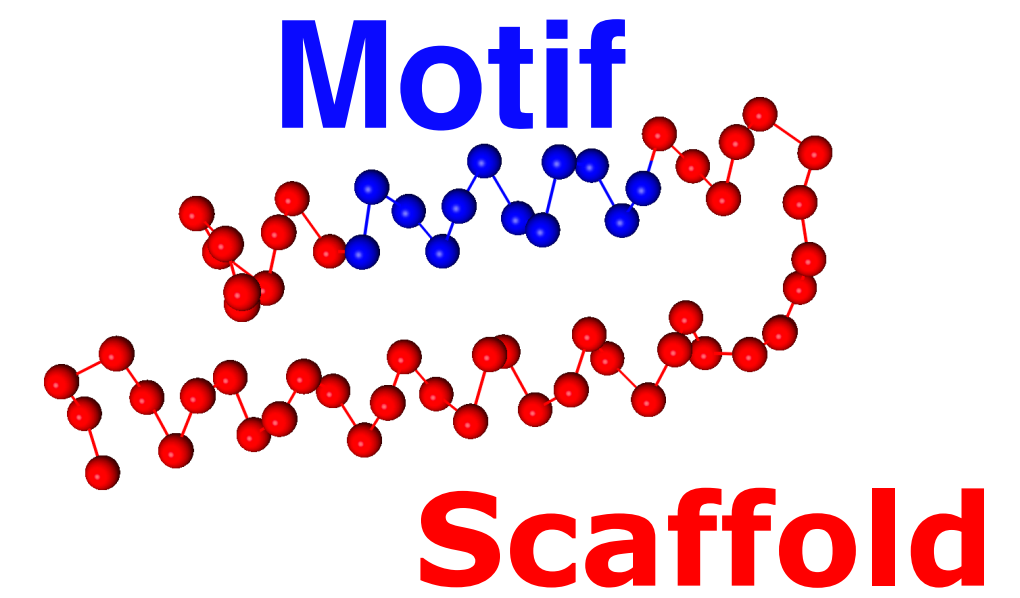
- Artifacts & edge effects in state-of-art inpainting





# Conditional Sampling to Fix Inpainting?

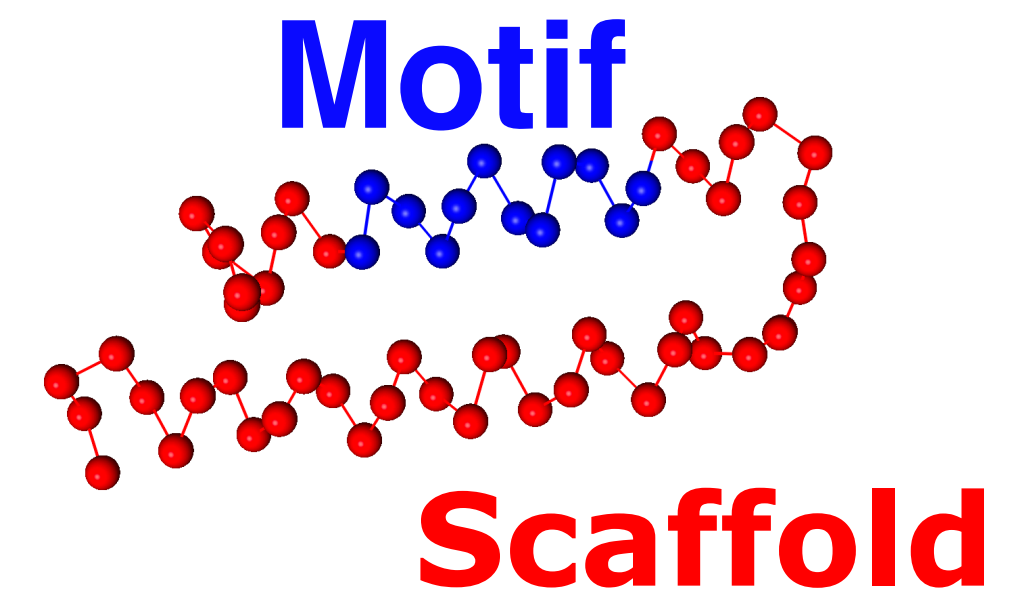
- Partition structure as  $x = [x_M, x_S]$



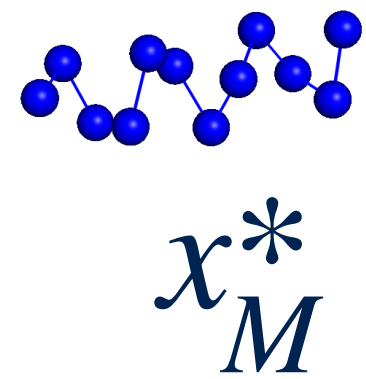
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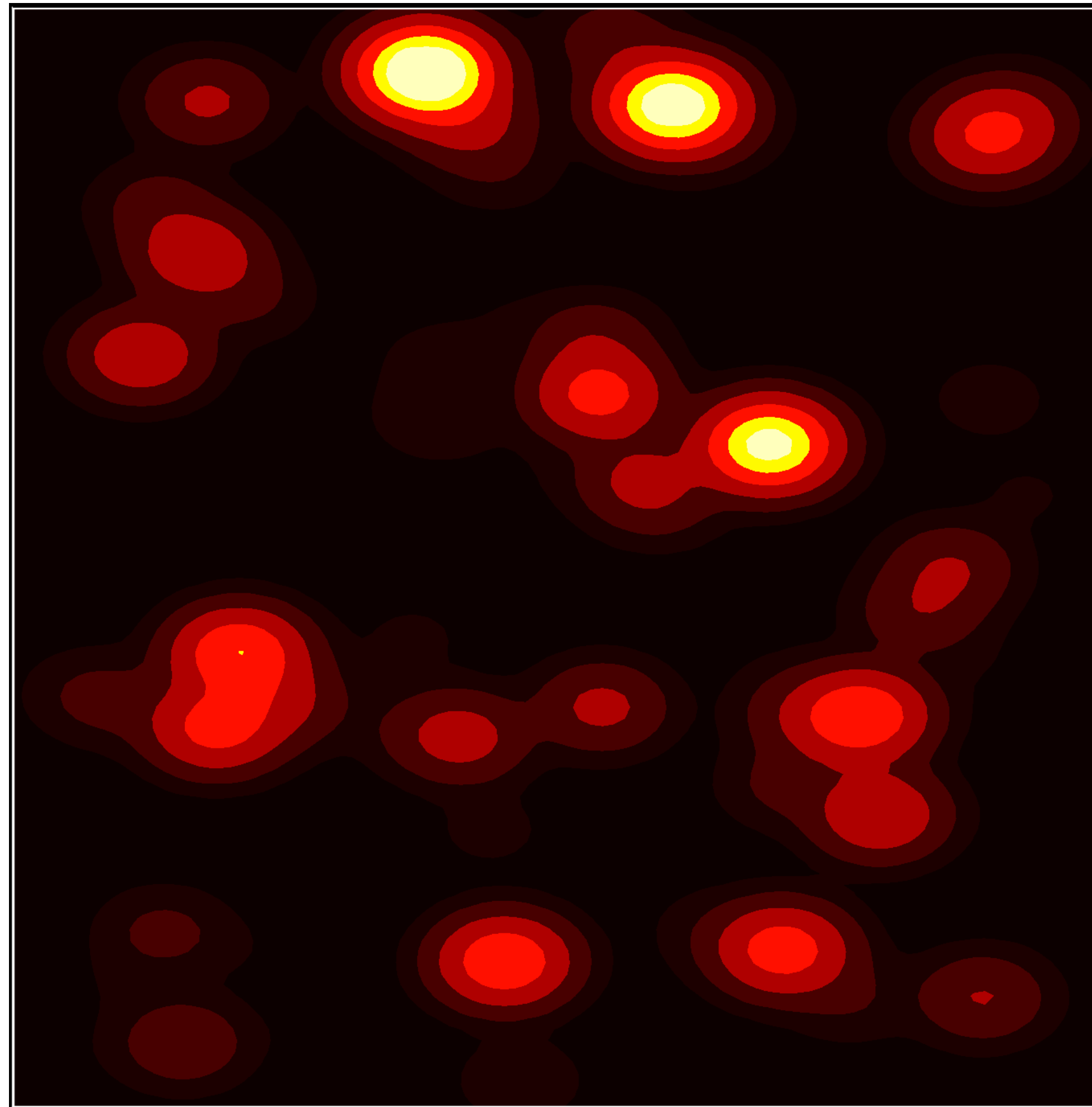
Structure space



**Motif**



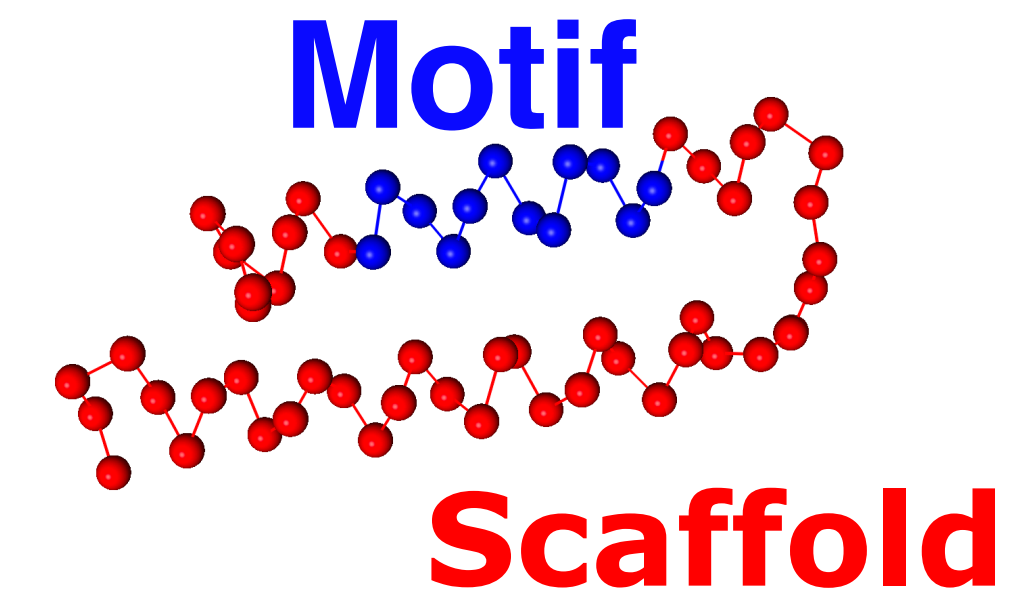
**Scaffold**





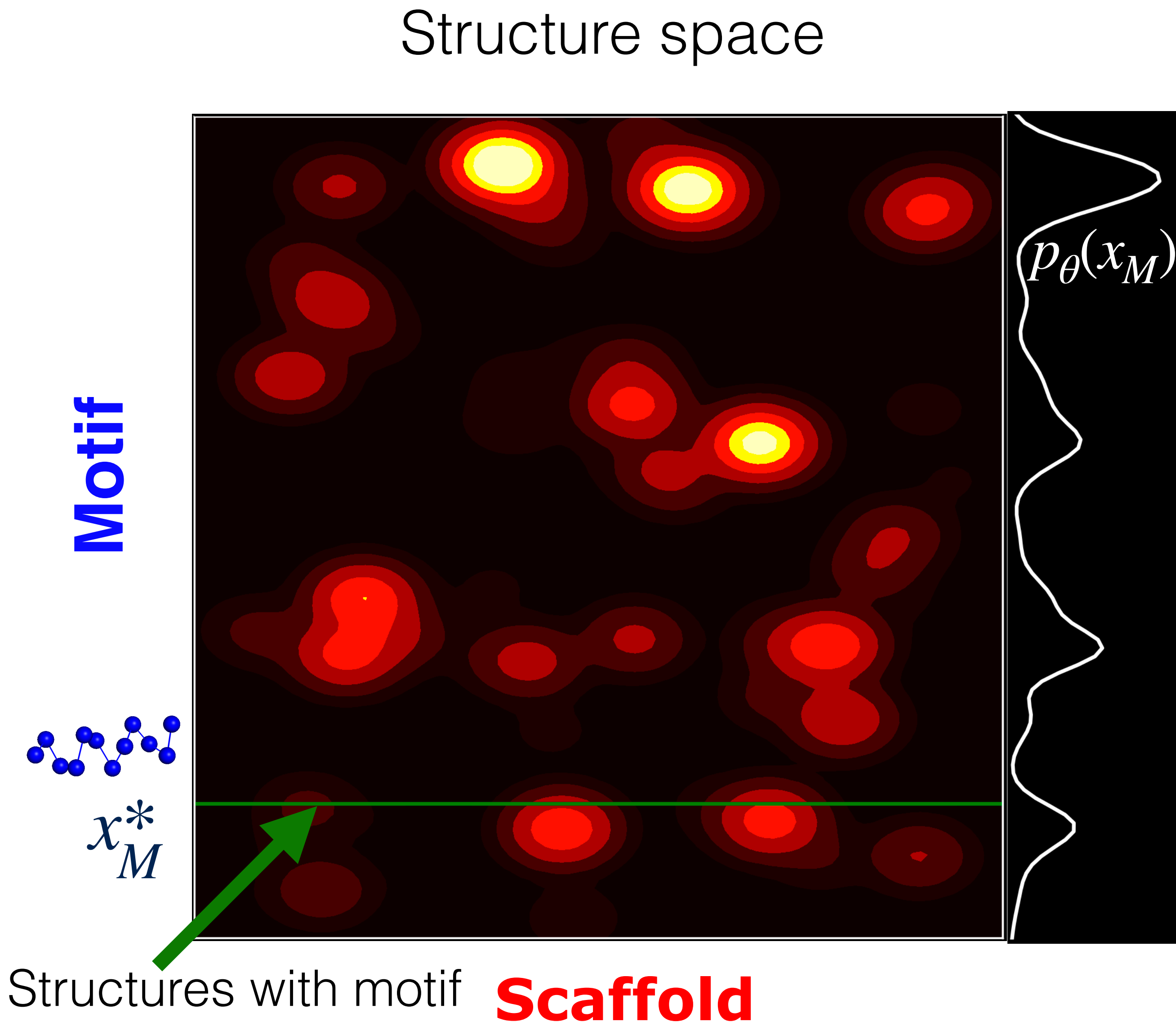
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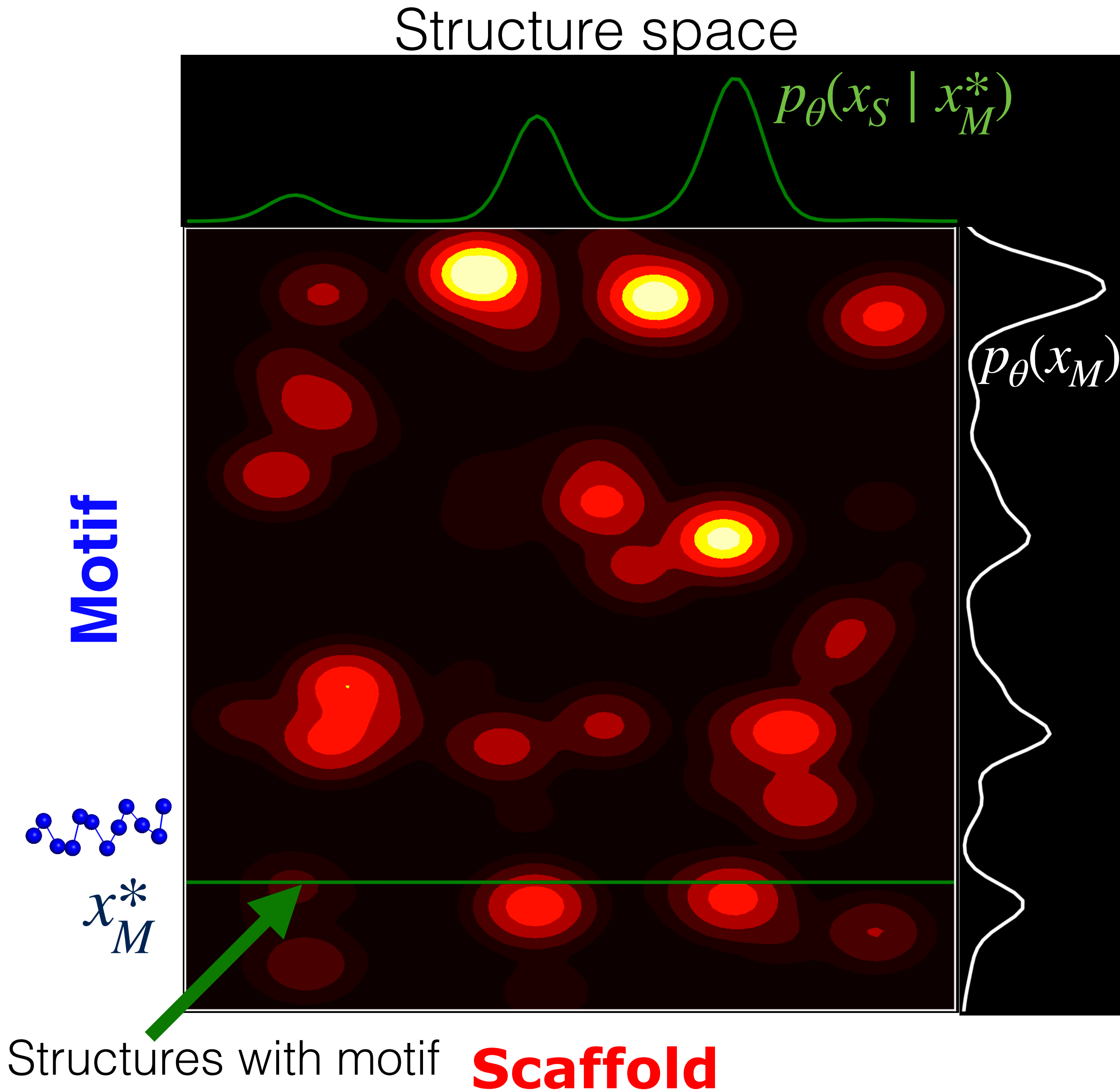


**Our rationale:** Assume

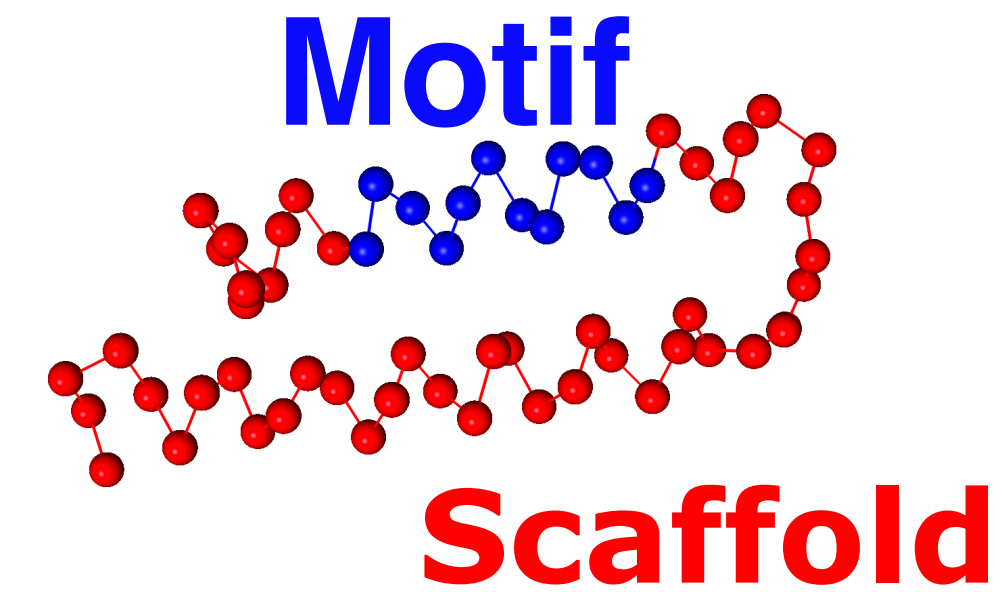
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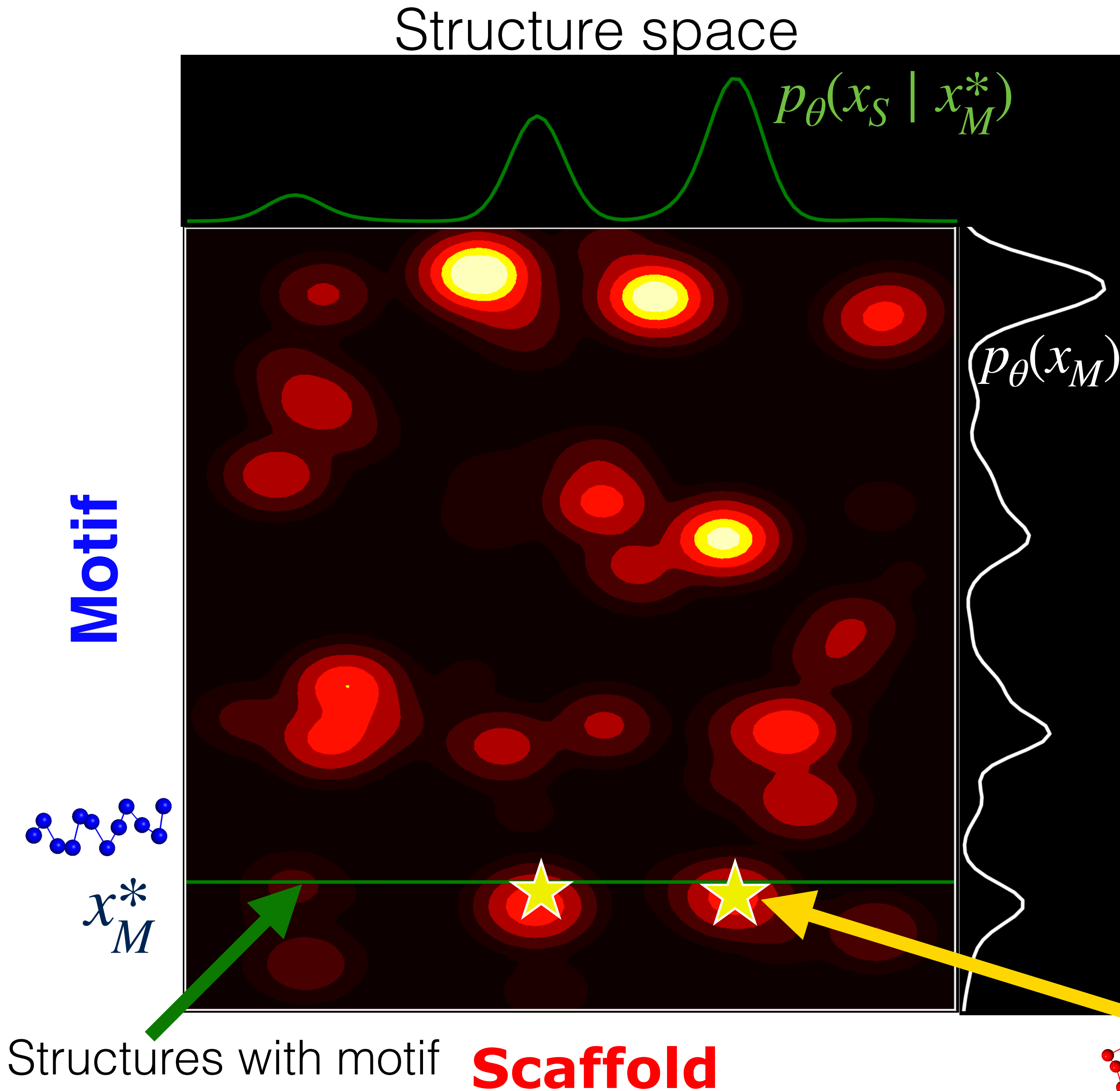


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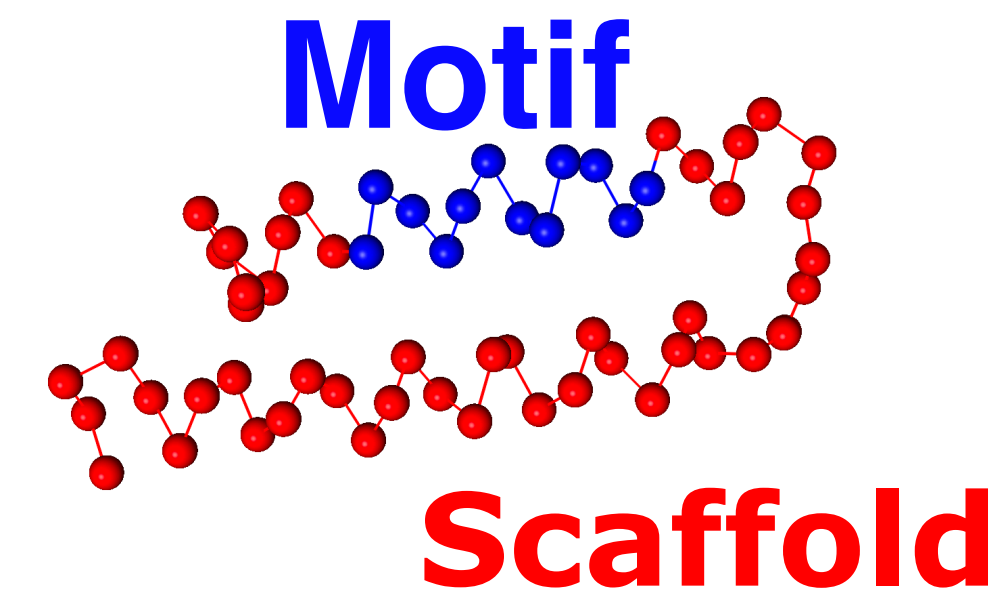
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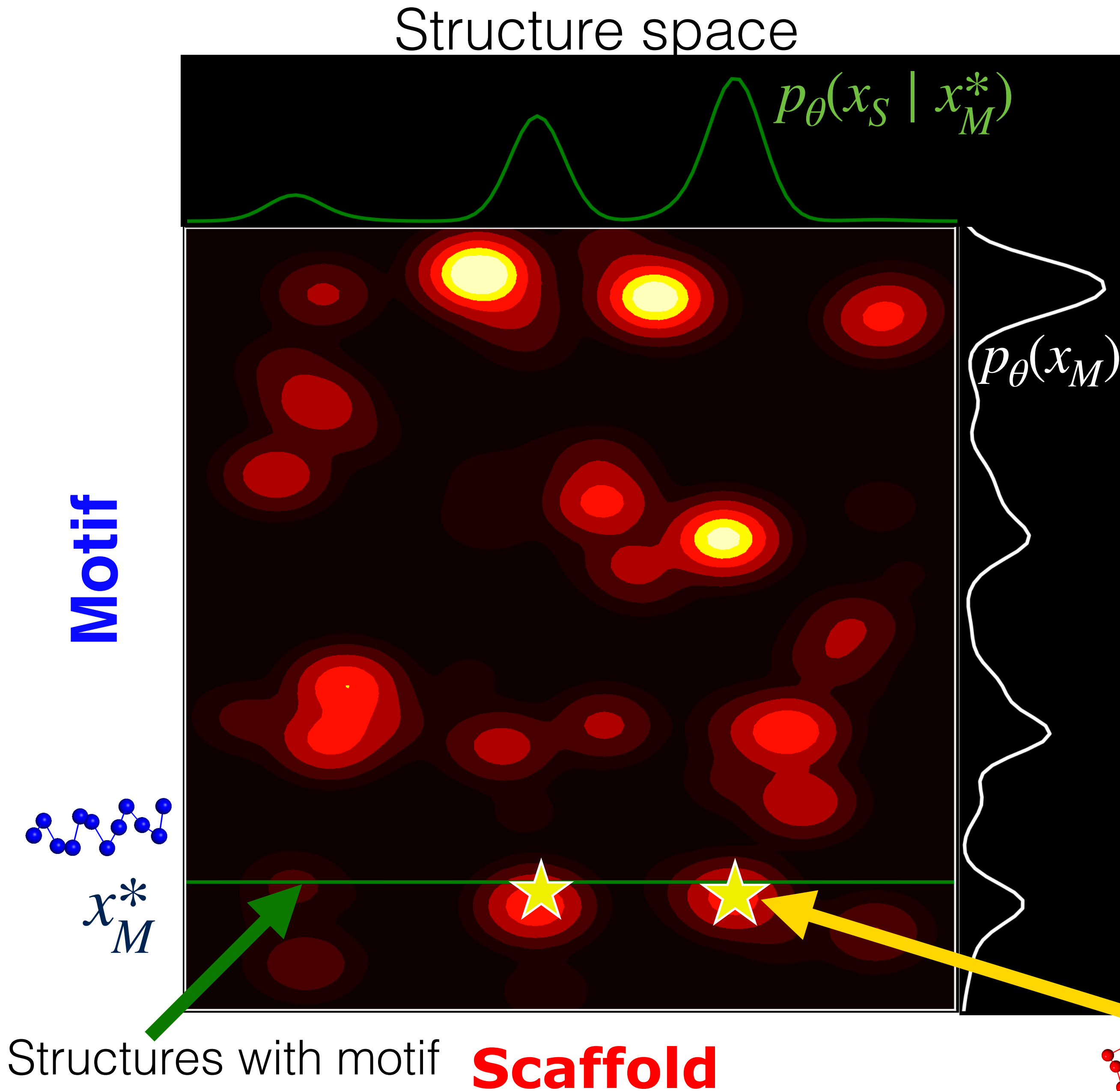
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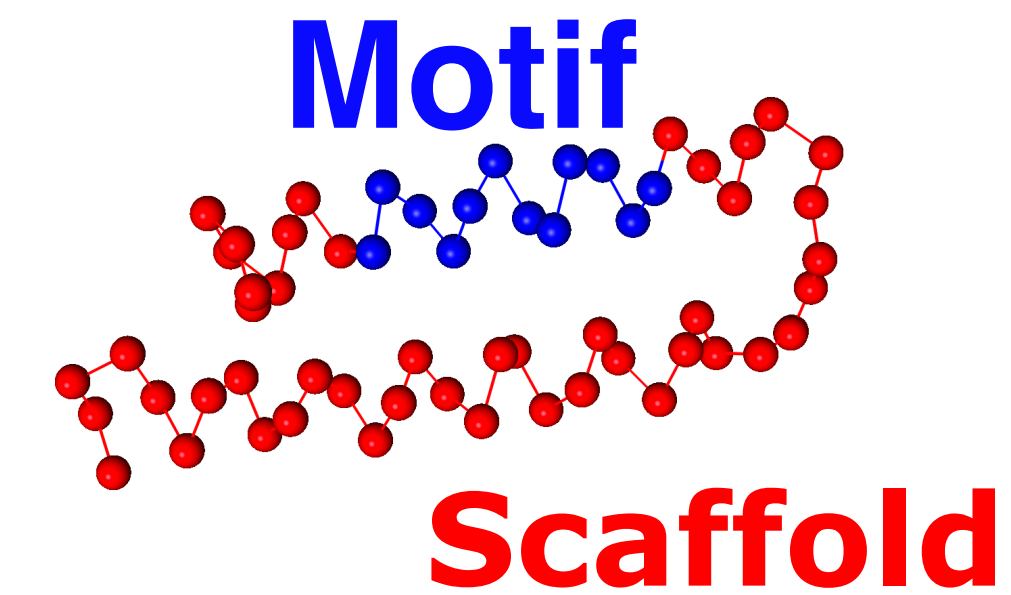
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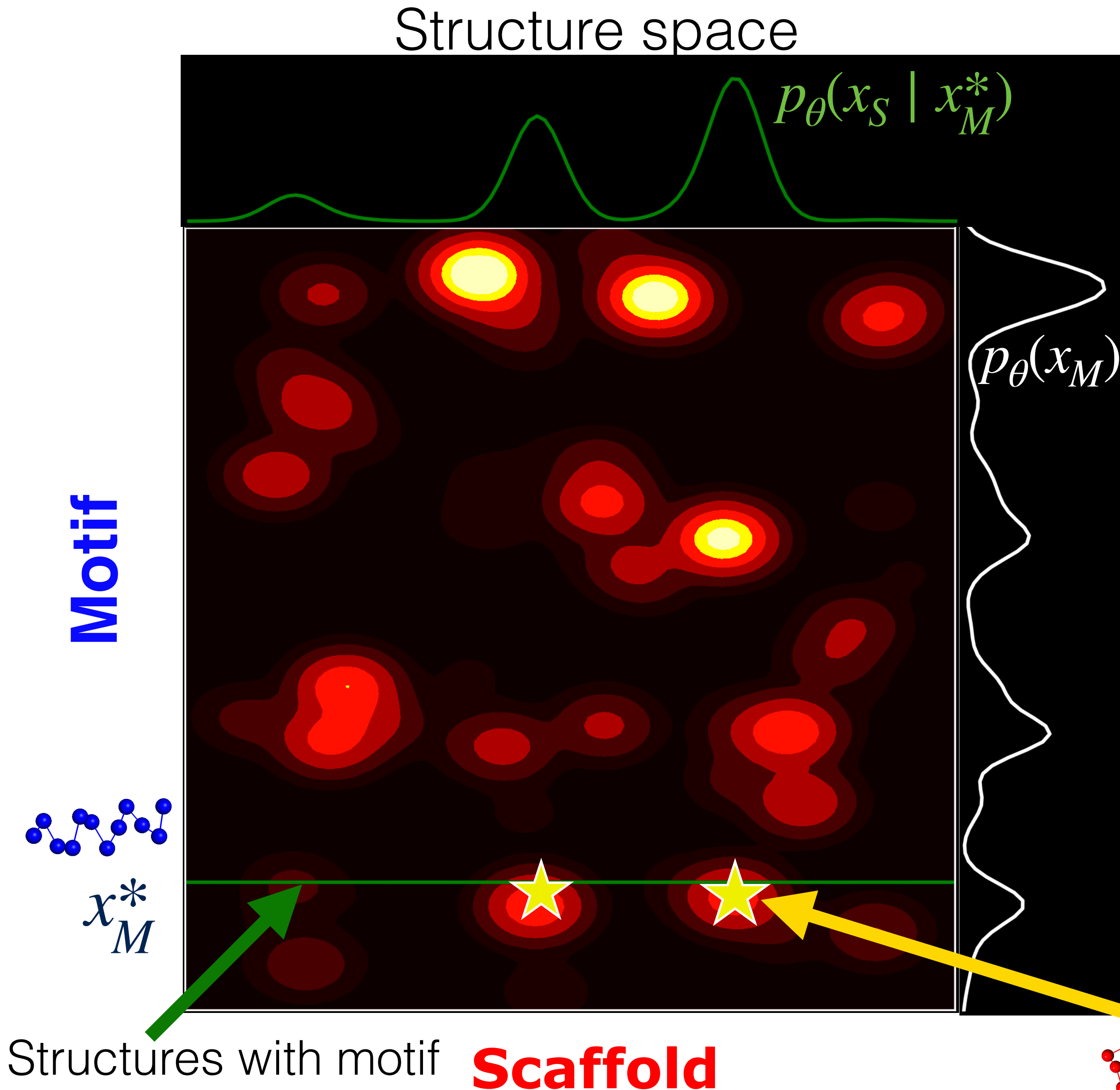
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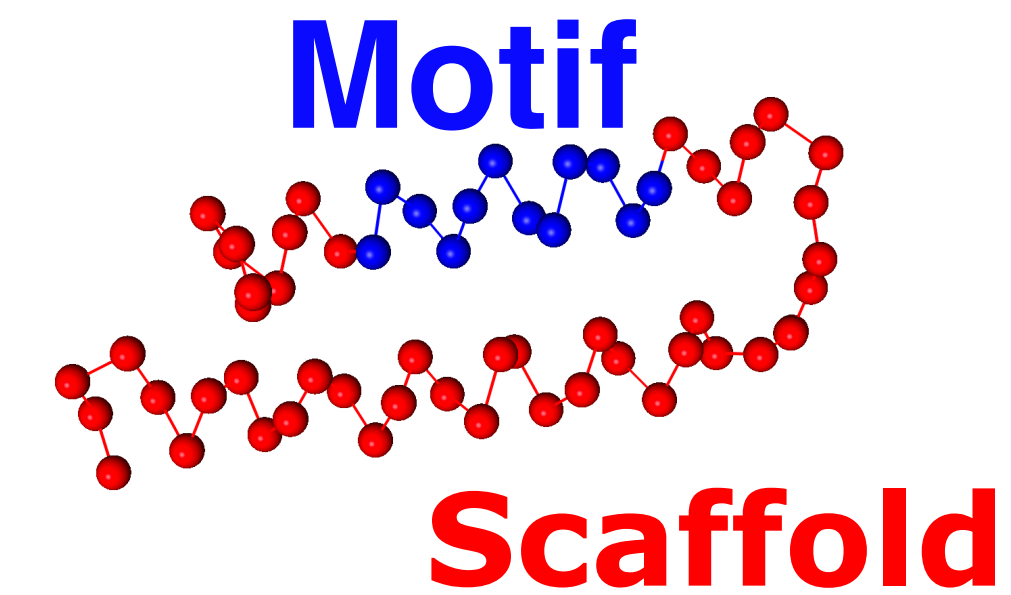
**Result:** With a good enough  $p_{\theta}(x)$ ,  
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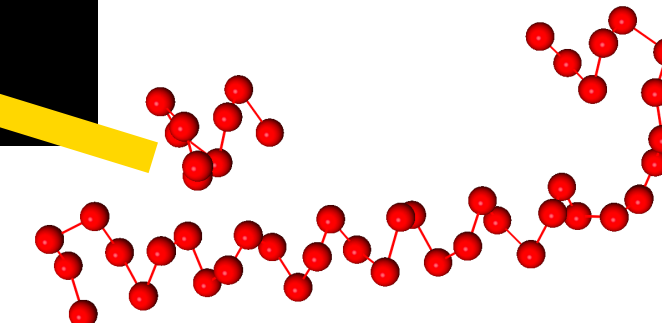


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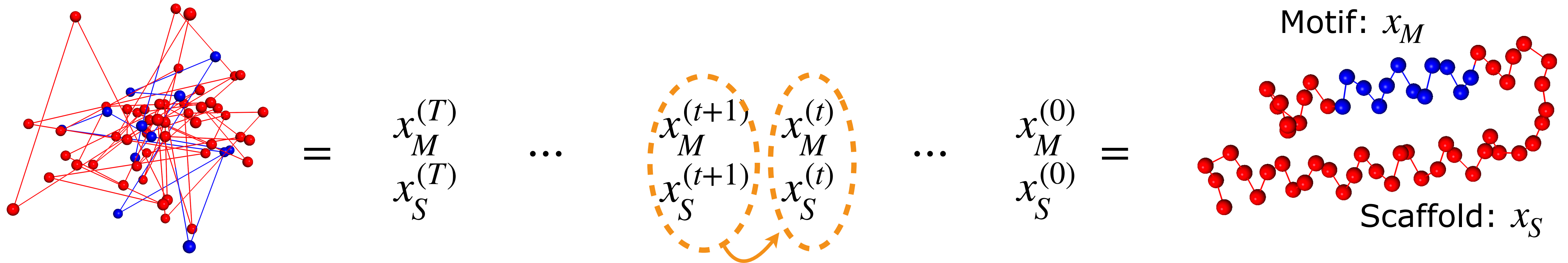
**Challenge:** How to sample  $x_S \sim p_{\theta}(x_S | x_M^*)$



# Inpainting with Diffusion Models

Unconditional Sampling:

$$x^{(t)} \sim p_{\theta}(x^{(t)} | x^{(t+1)})$$





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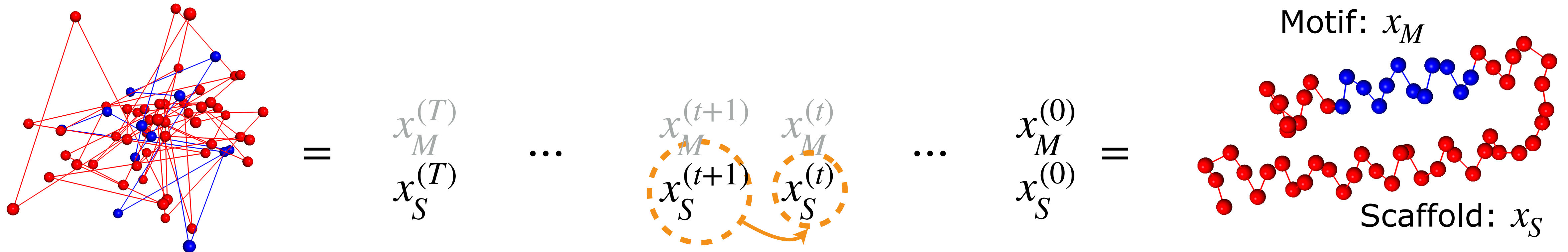
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- Exact but intractable!



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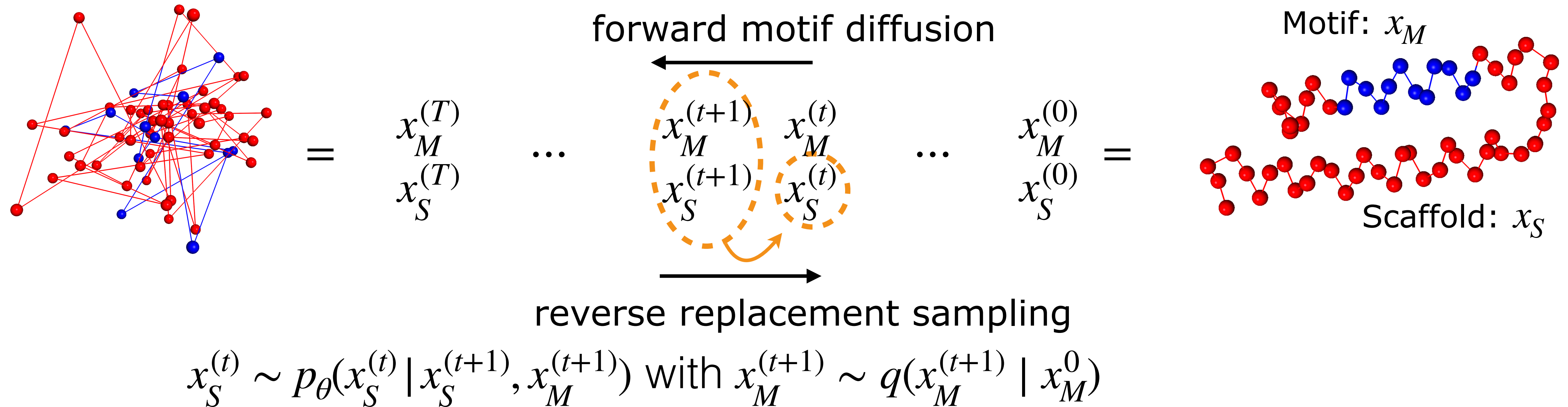
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Tractable alternative: Replacement approach [Song 2021, Meng 2021]





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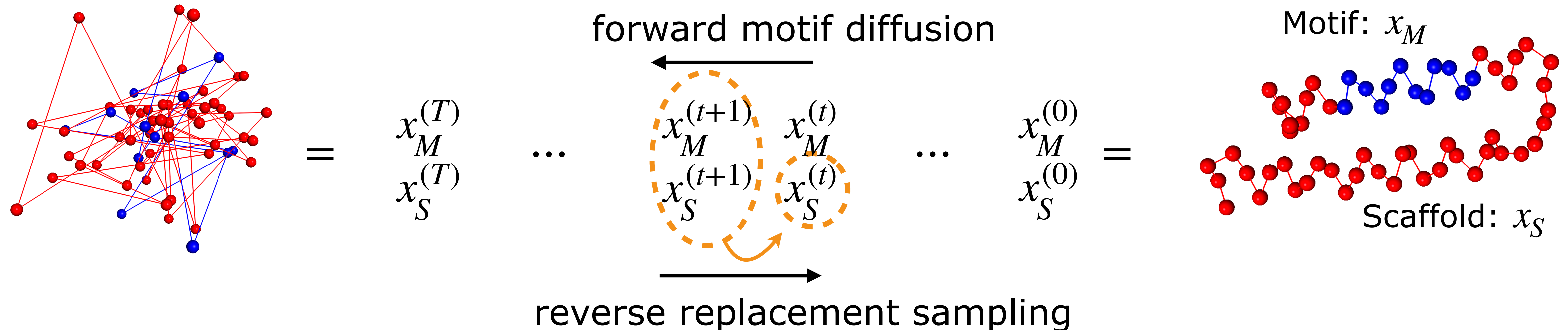
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- Introduces approximation error and leads to chain breaks!

# Improved inpainting with SMCDiff

Replacement Method:

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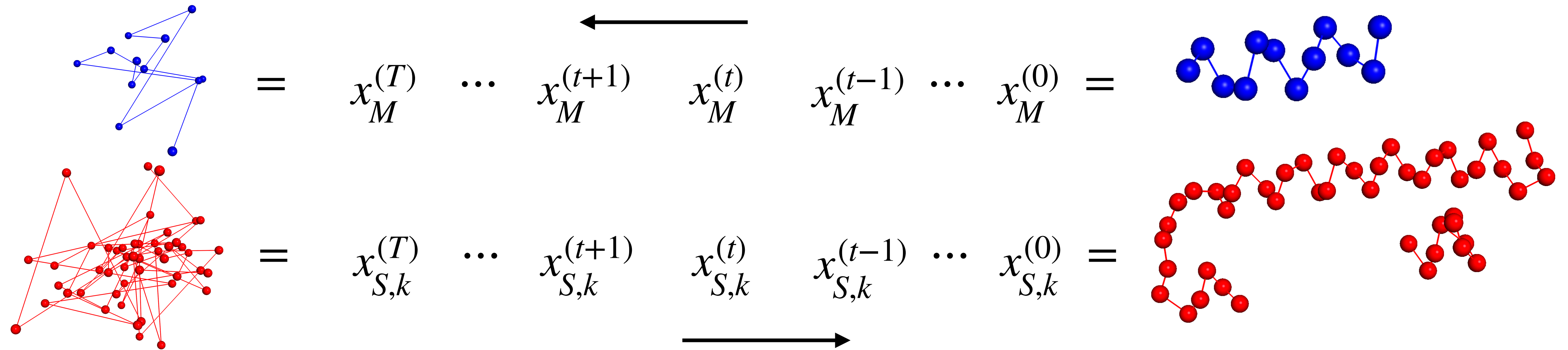
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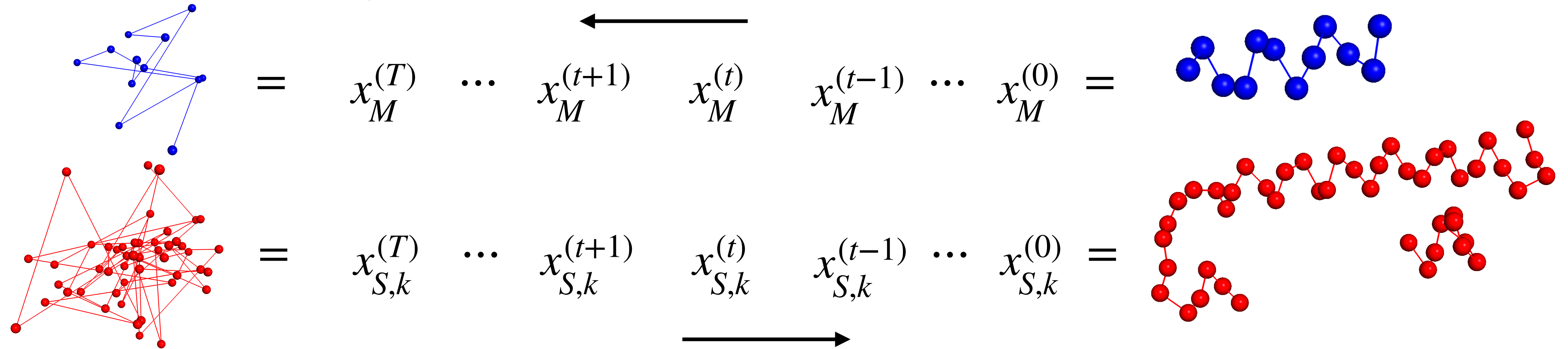
# Improved inpainting with SMCDiff





# Improved inpainting with SMCDiff

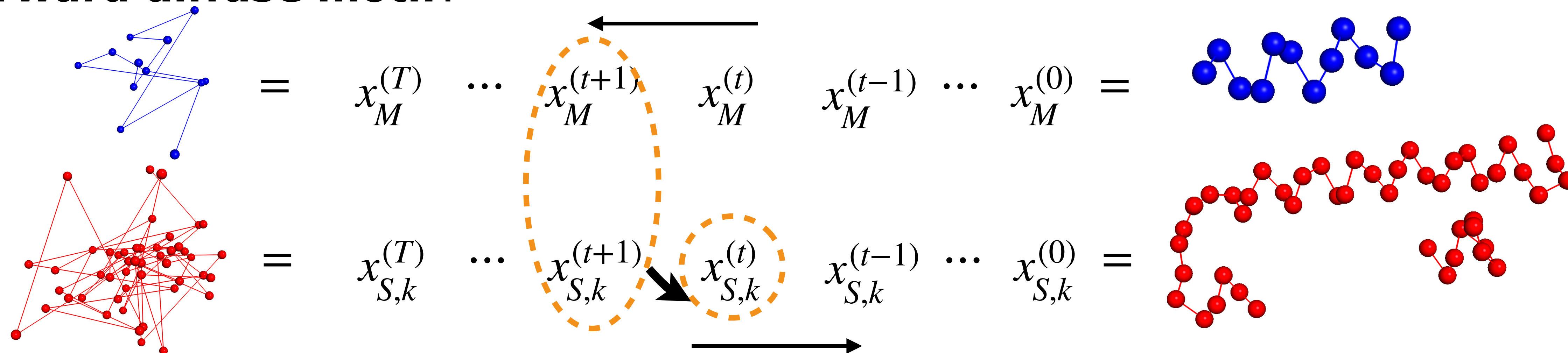
- Forward diffuse motif:



- Reverse diffuse K weighted scaffold "particles"

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## Forward diffuse motif:



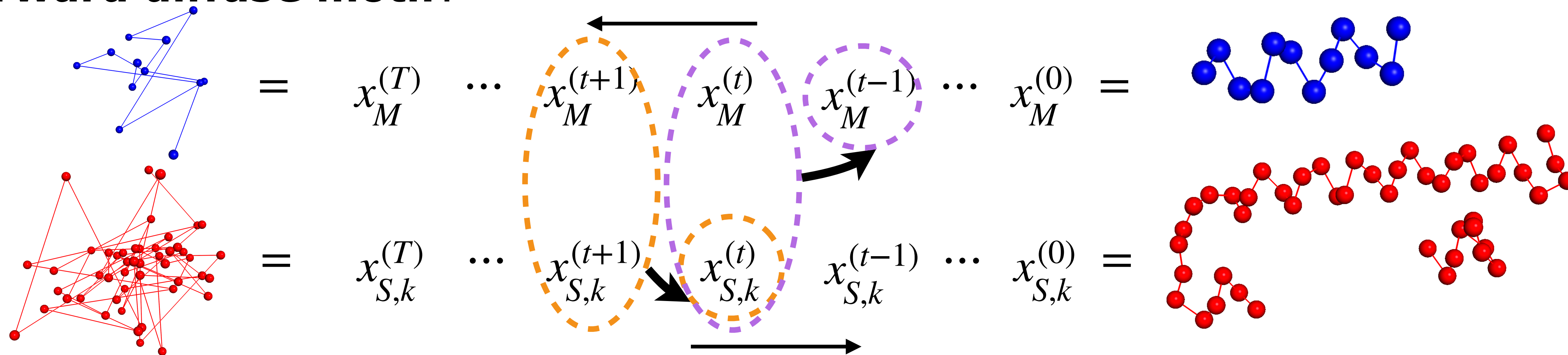
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# Improved inpainting with SMCDiff

## Forward diffuse motif:



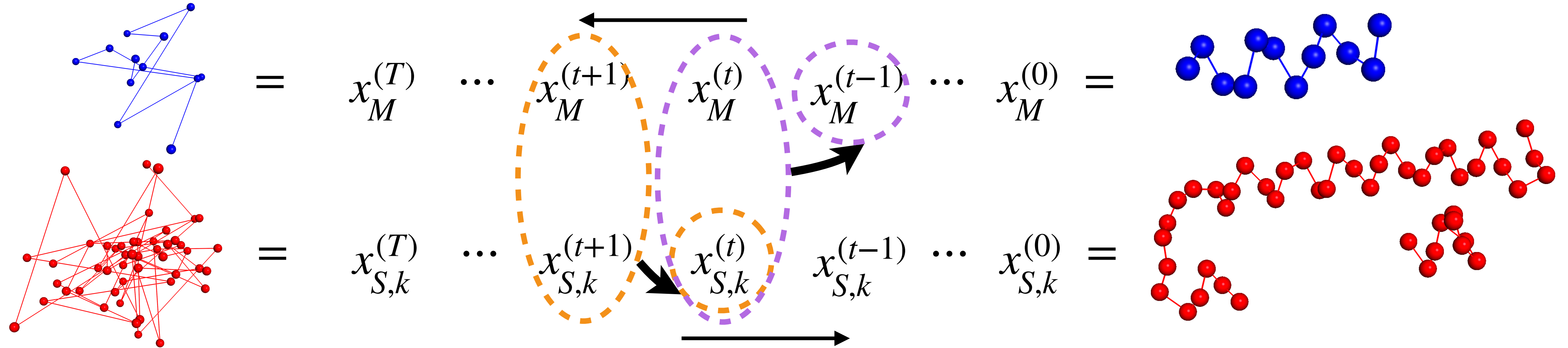
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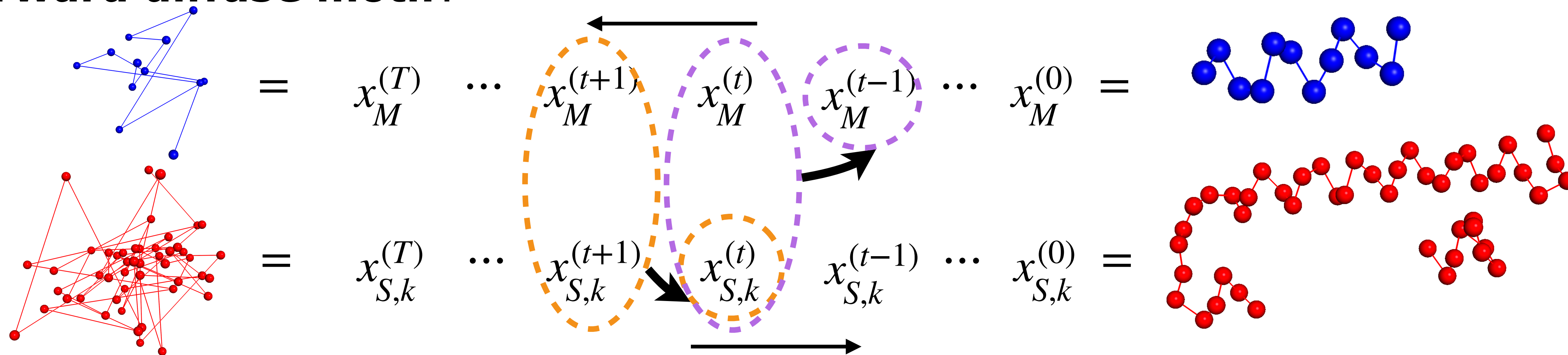
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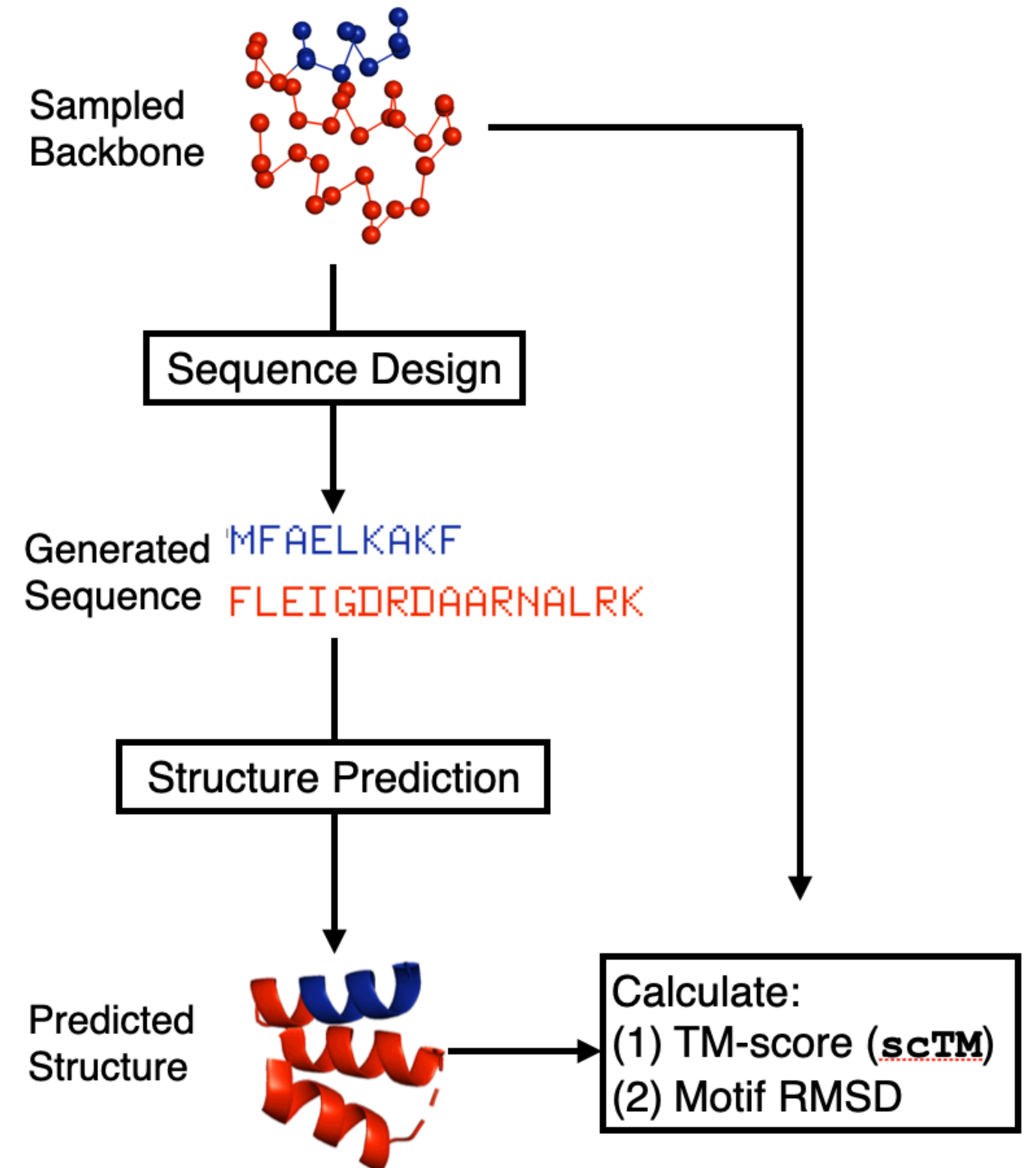
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**Proposition (informal):** If  $p_{\theta}(x^{(0)}) = q(x^{(0)})$ , then  $x_{S,k}^{(0)} \xrightarrow[K \rightarrow \infty]{d} q(x_S^{(0)} | x_M^{(0)})$ .

# Motif self-consistency

- Similar evaluation as before.
- Also calculate **Motif RMSD**
  - Achieving motif RMSD < 1.0Å is imperative in motif-scaffolding.
- Criterion for successful scaffolding:
  - motif RMSD < 1.0Å
  - **scTM** > 0.5

## Self-consistency evaluation

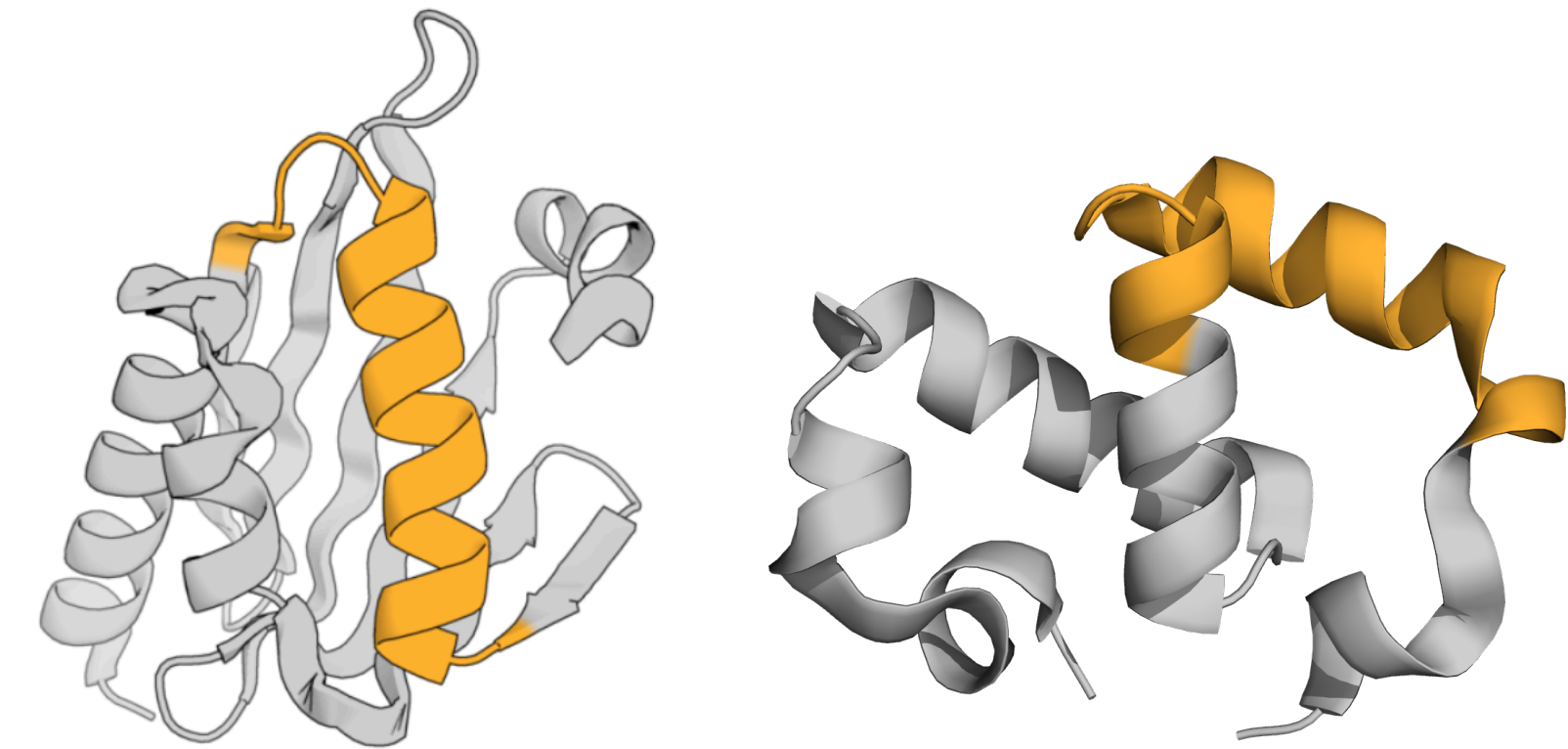




# Motif-scaffolding case-studies

## Evaluation Setup

- Pick motifs from structures in PDB
- Know at least one solution exists



	5trv	6exz
Base Motif Length	21 res.	20 res.
Length	118 res.	72 res.

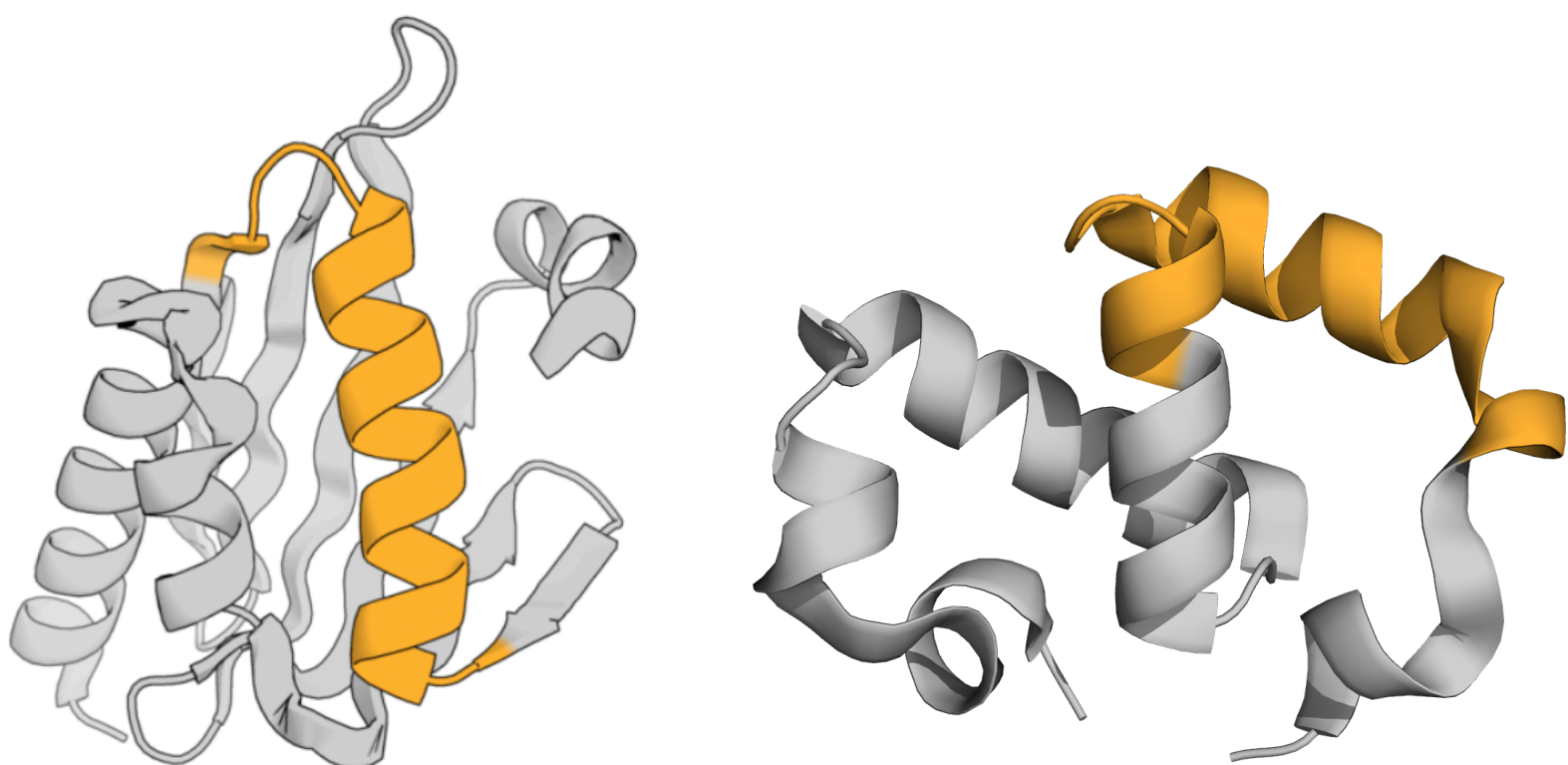
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- Provide variable length “pads” of native scaffold



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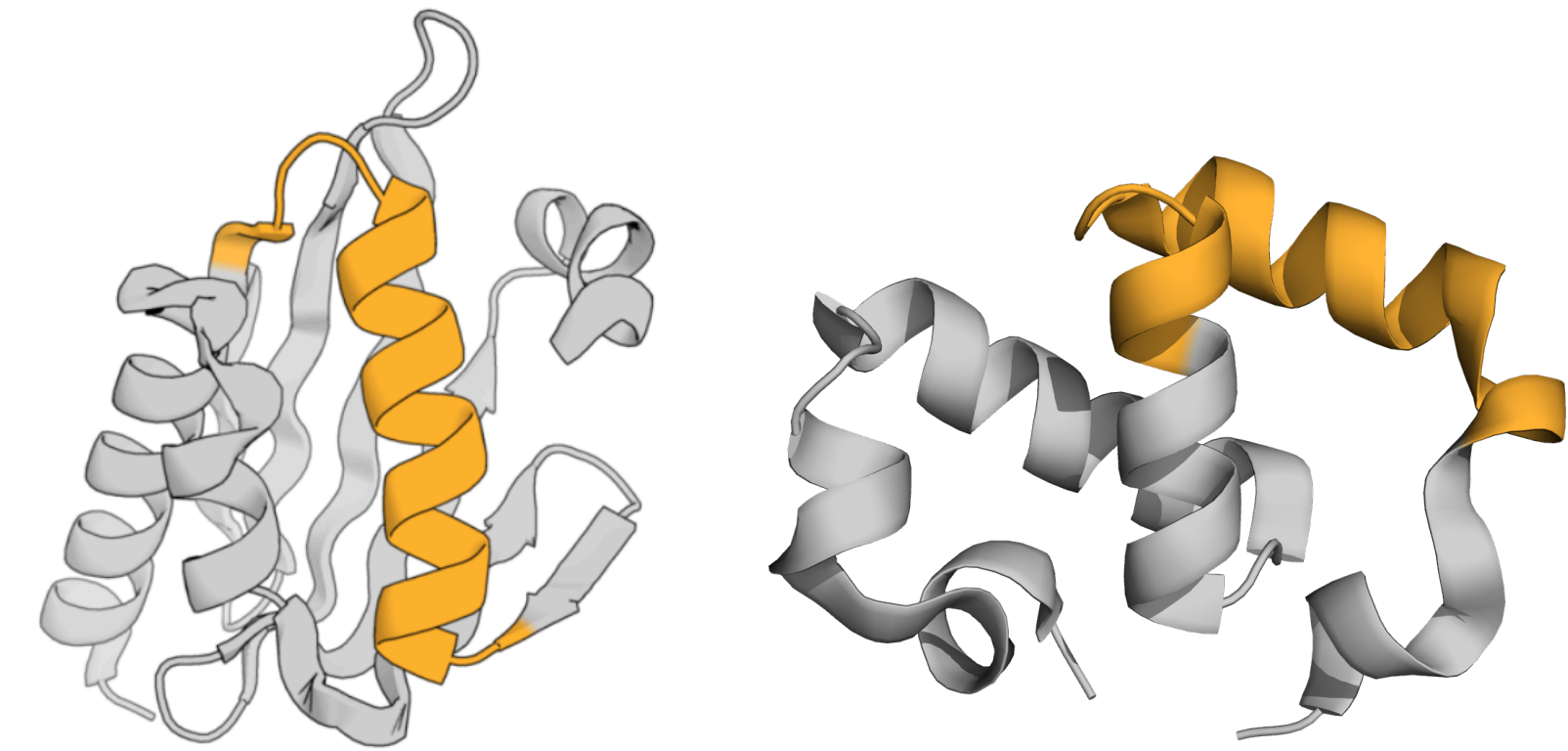
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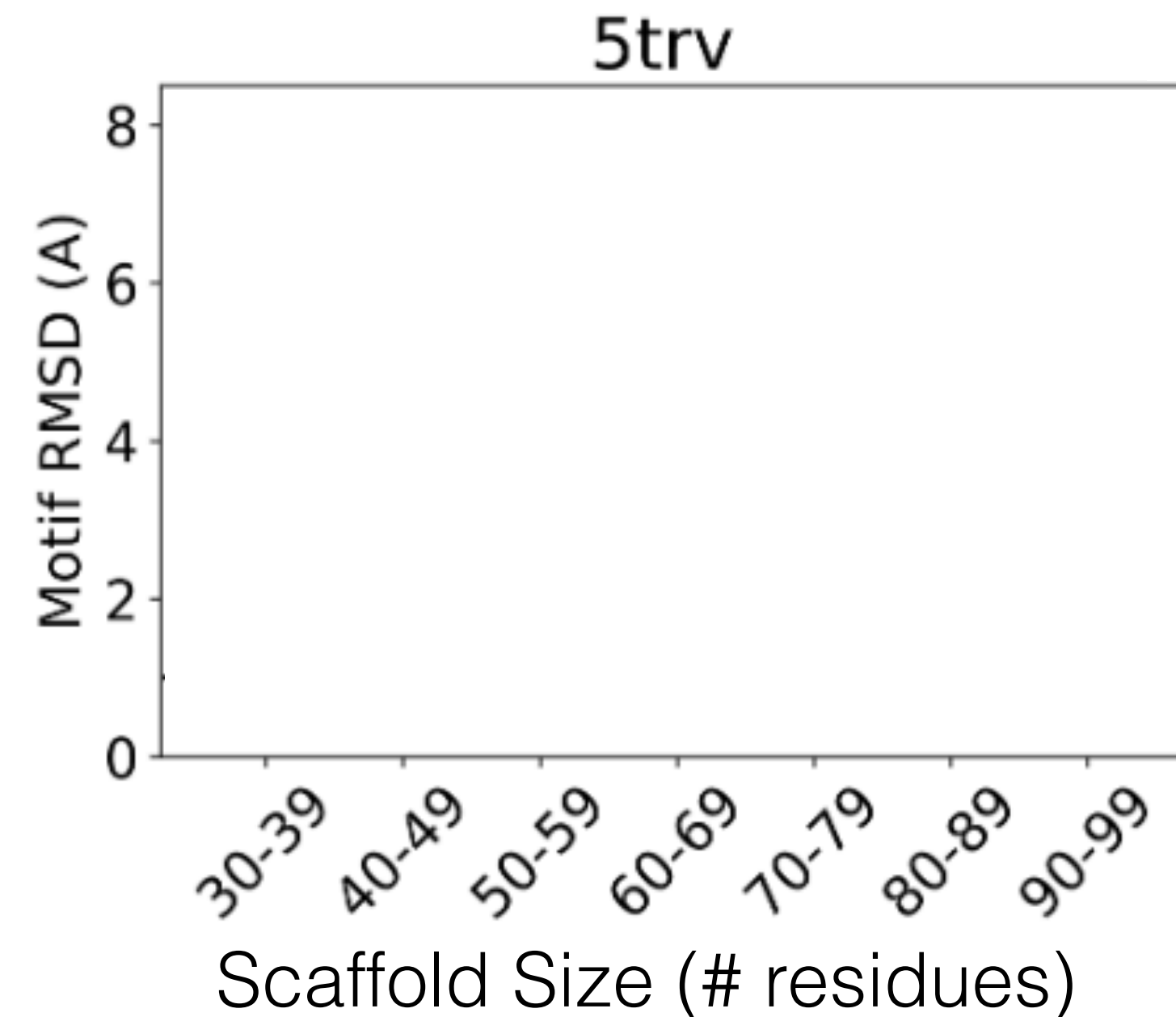
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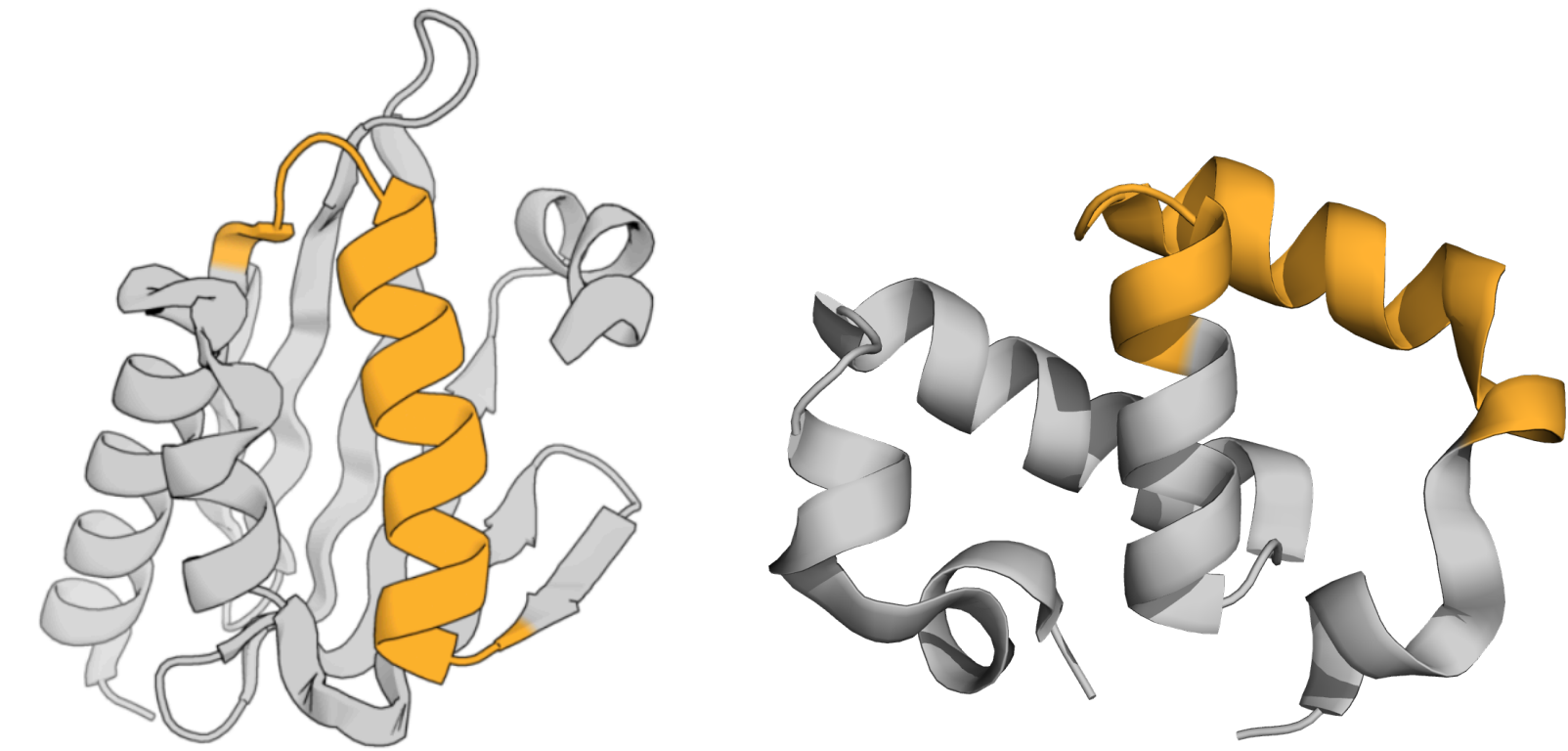
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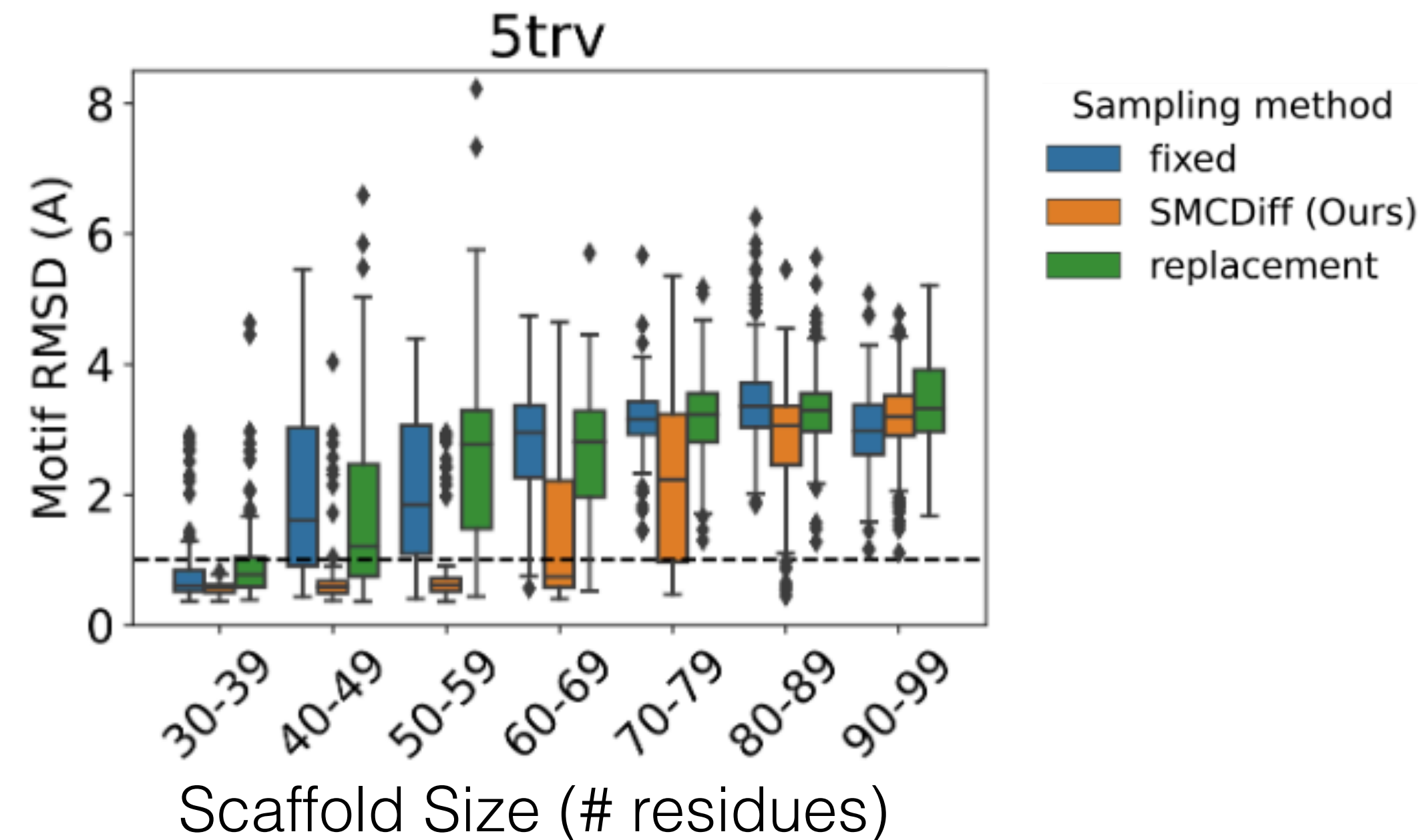
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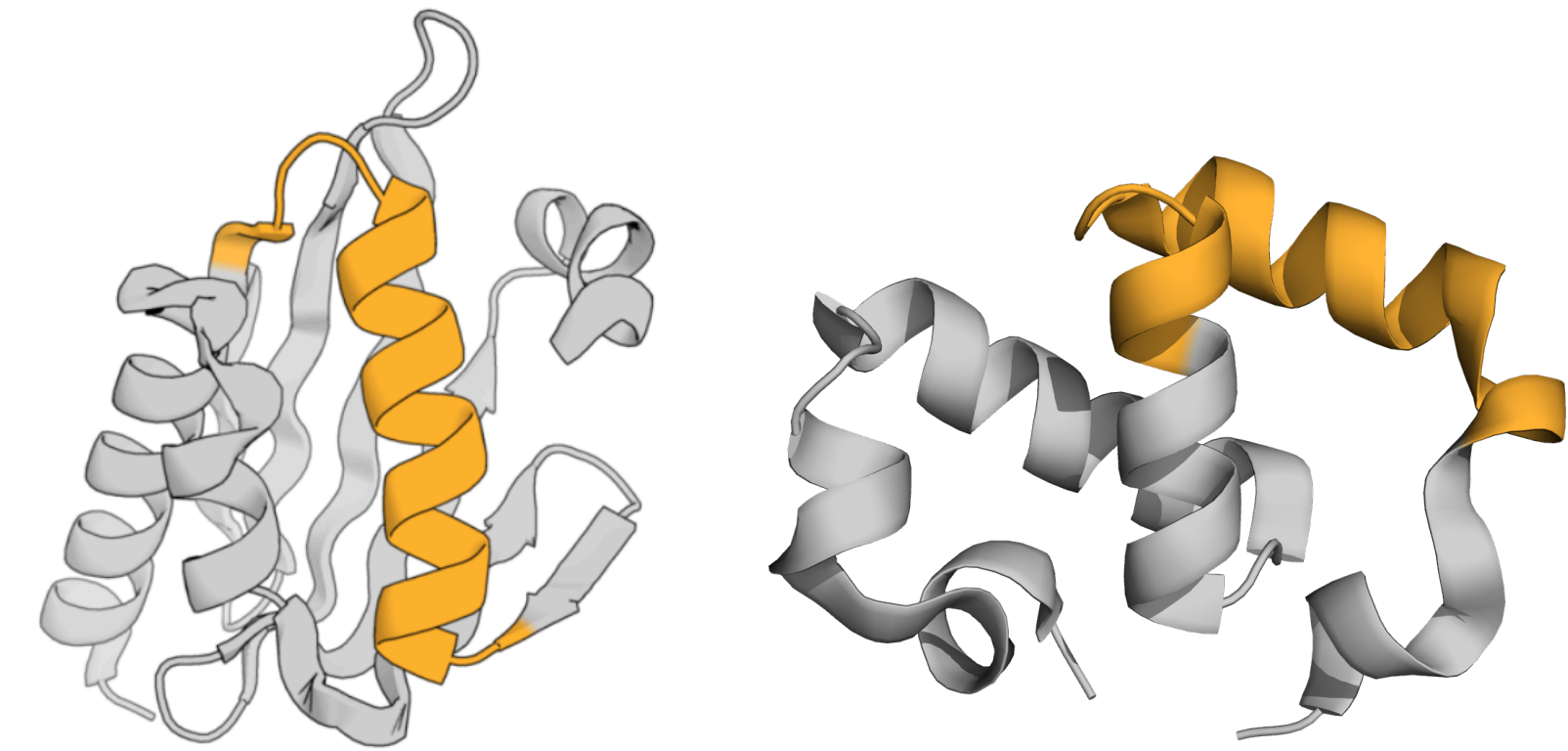
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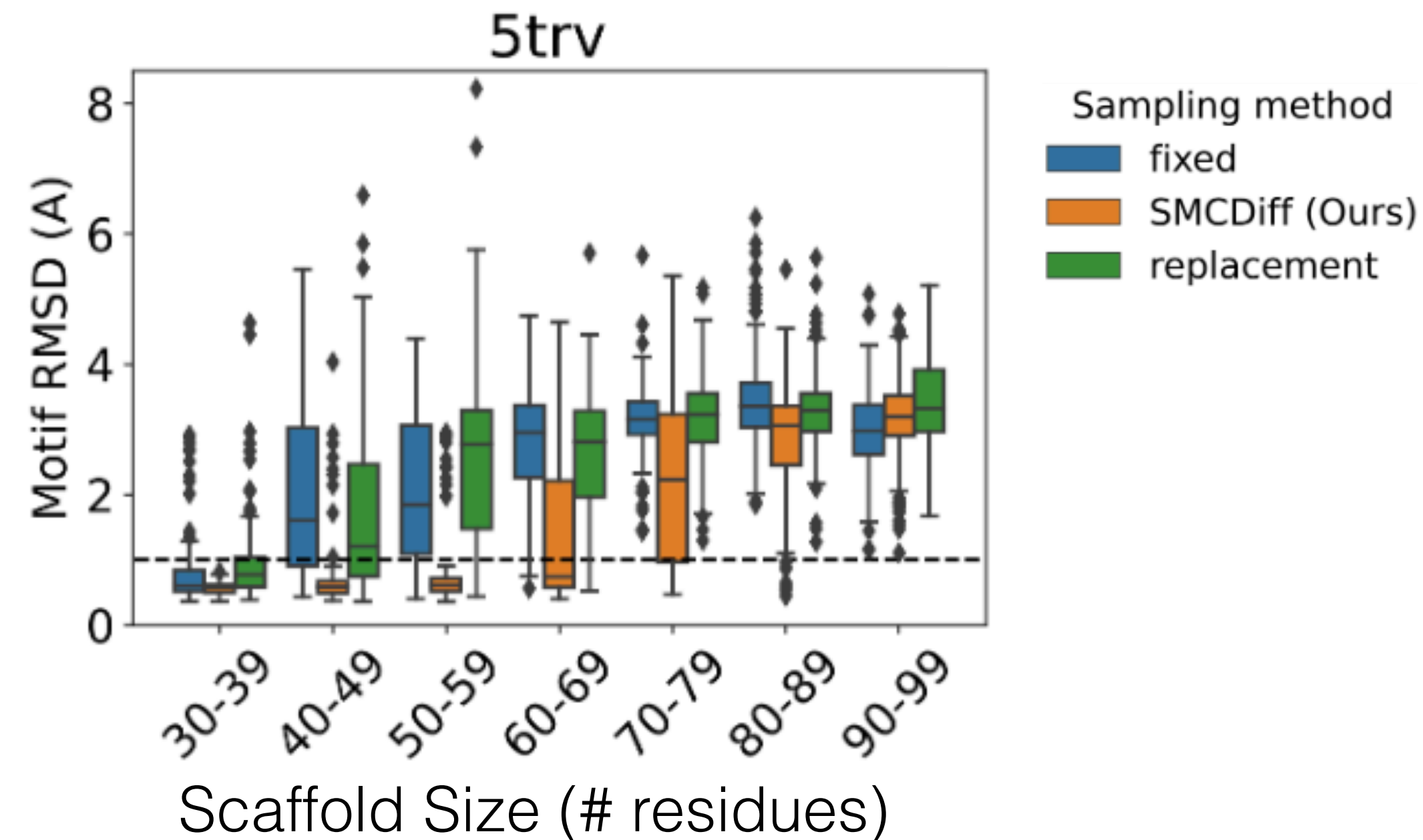
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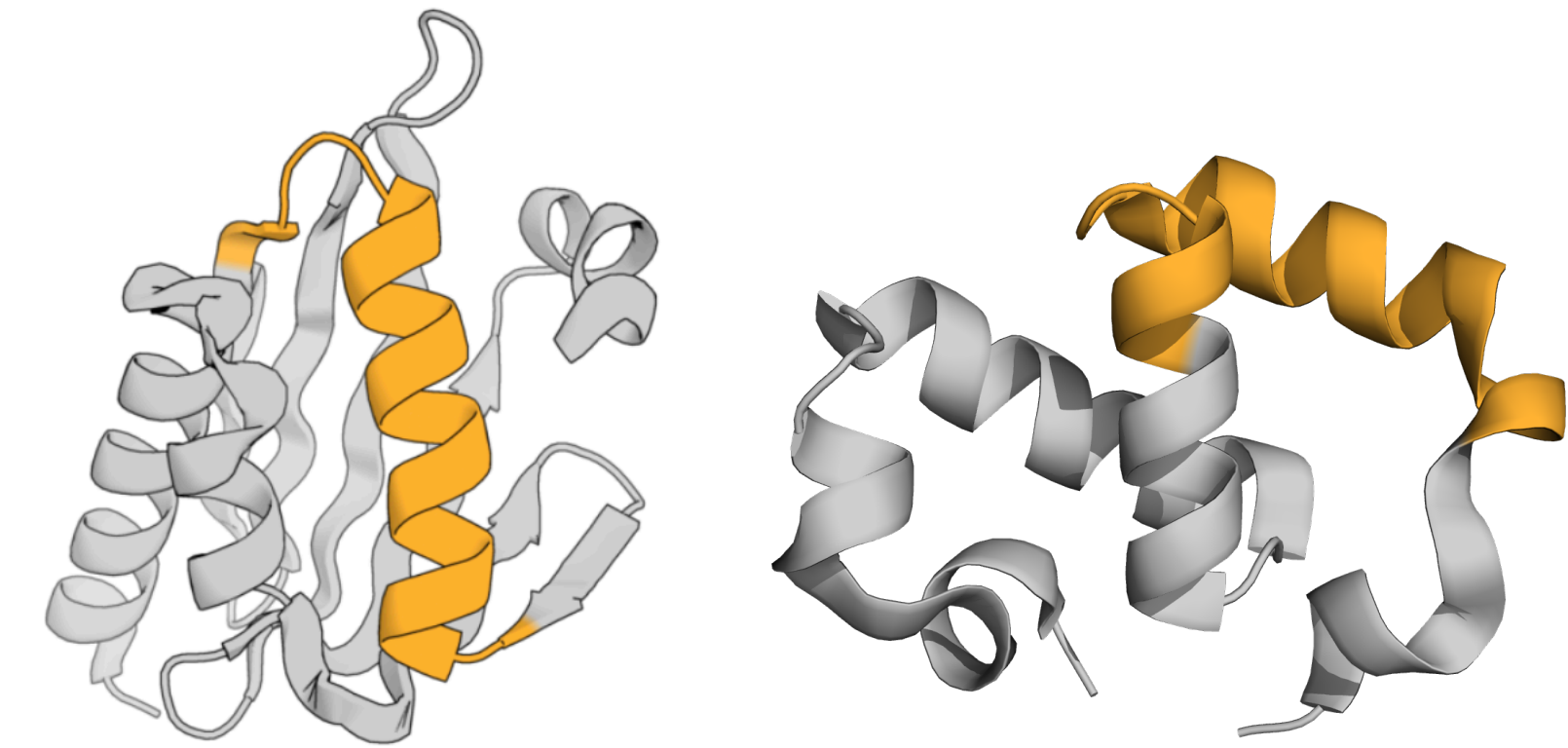
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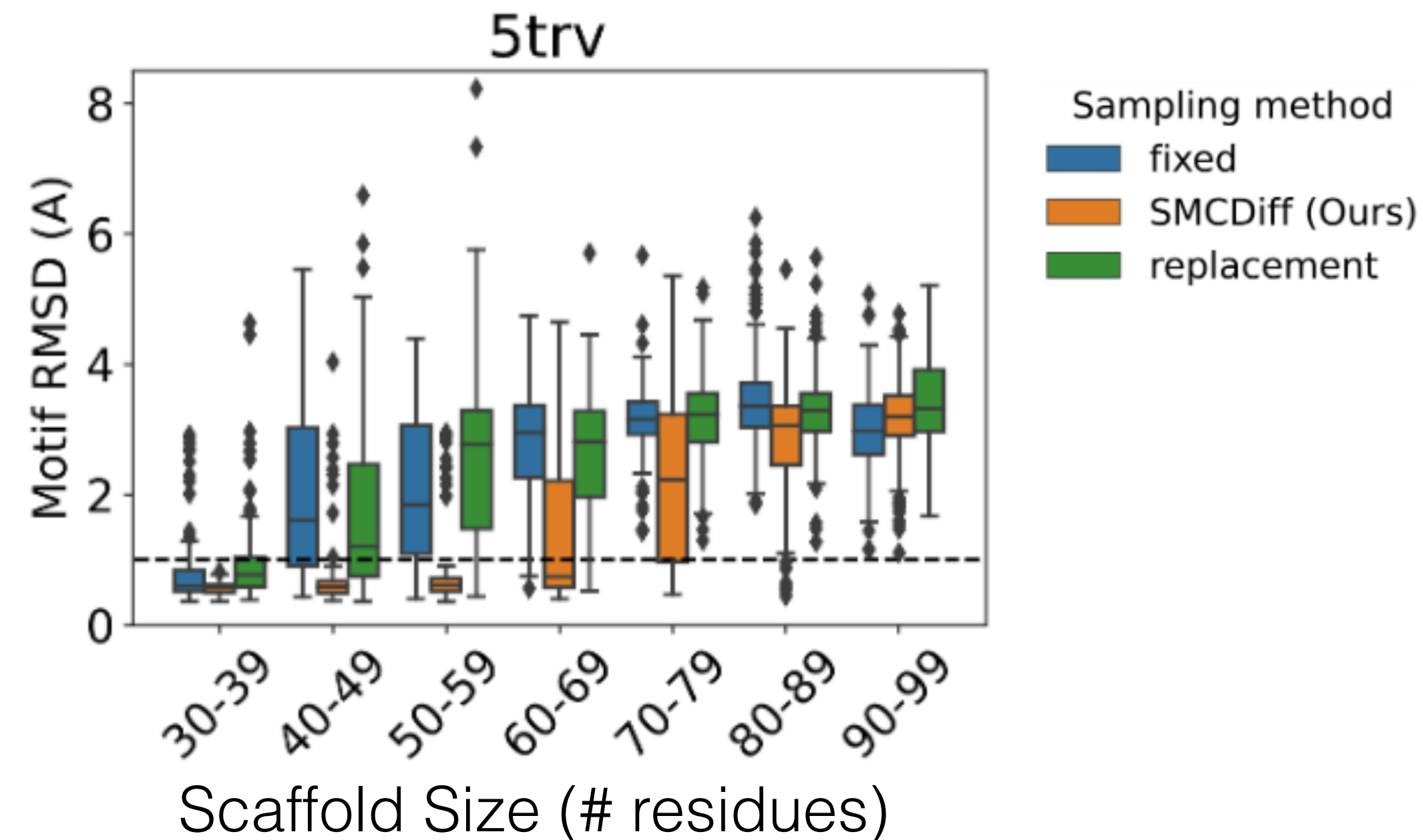
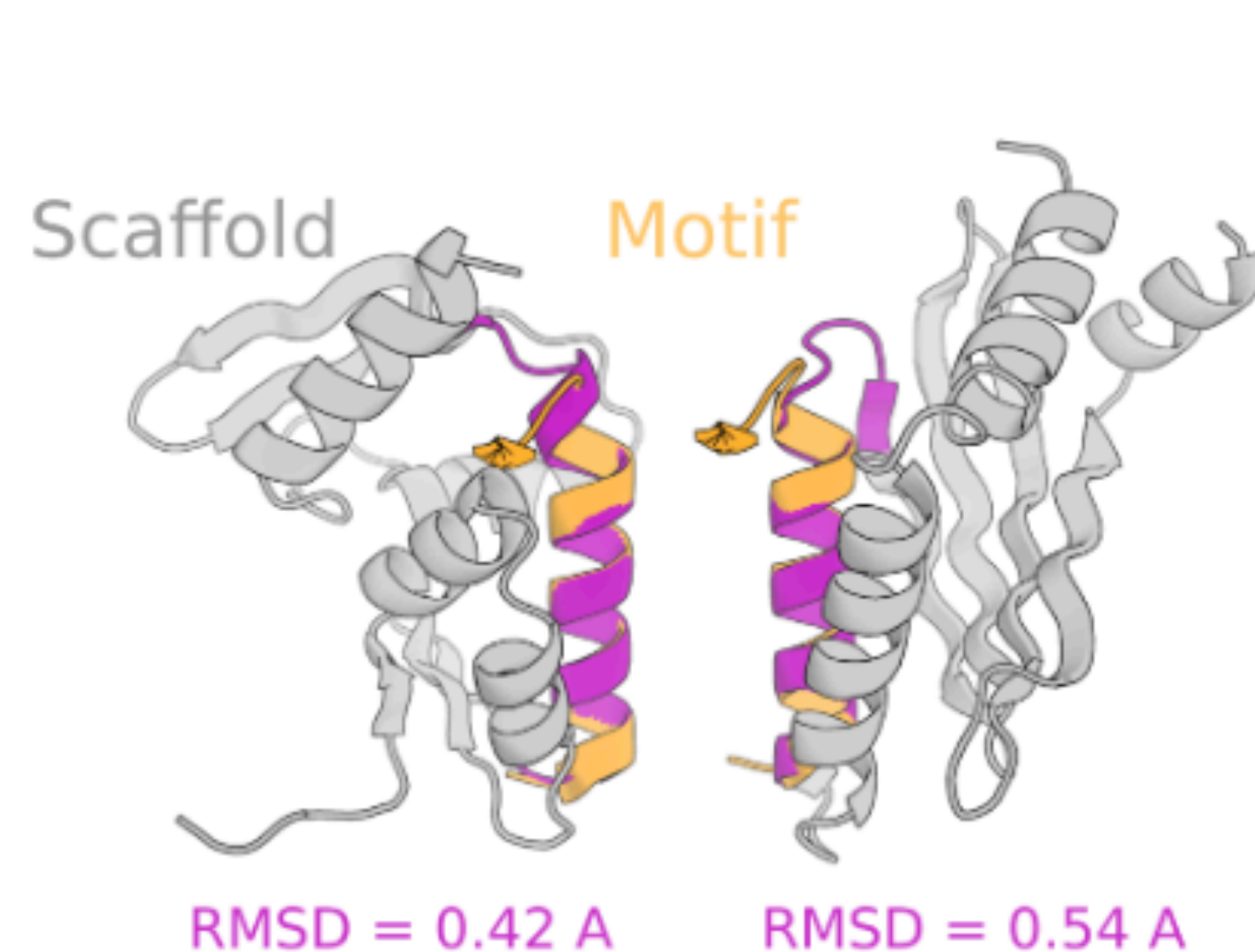
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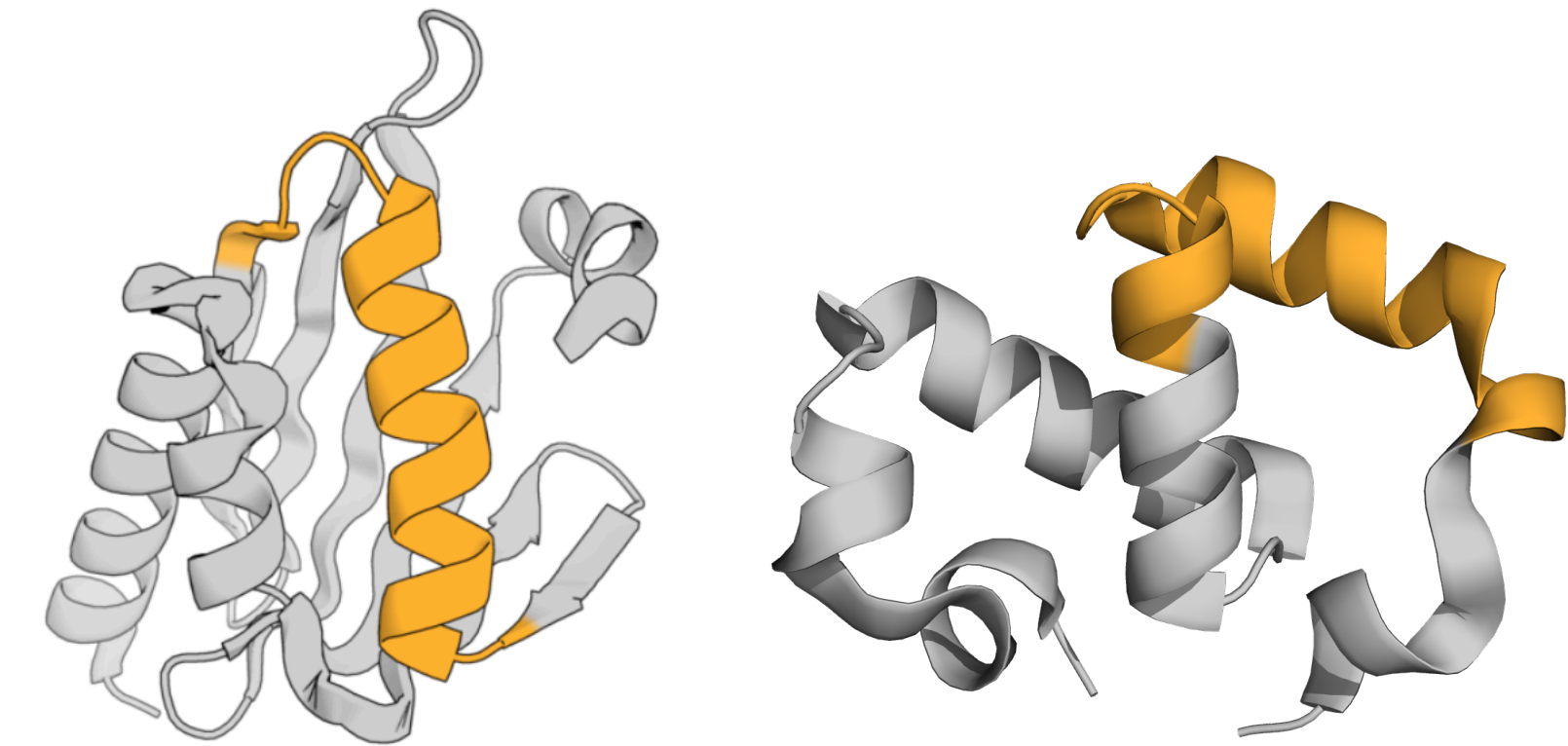
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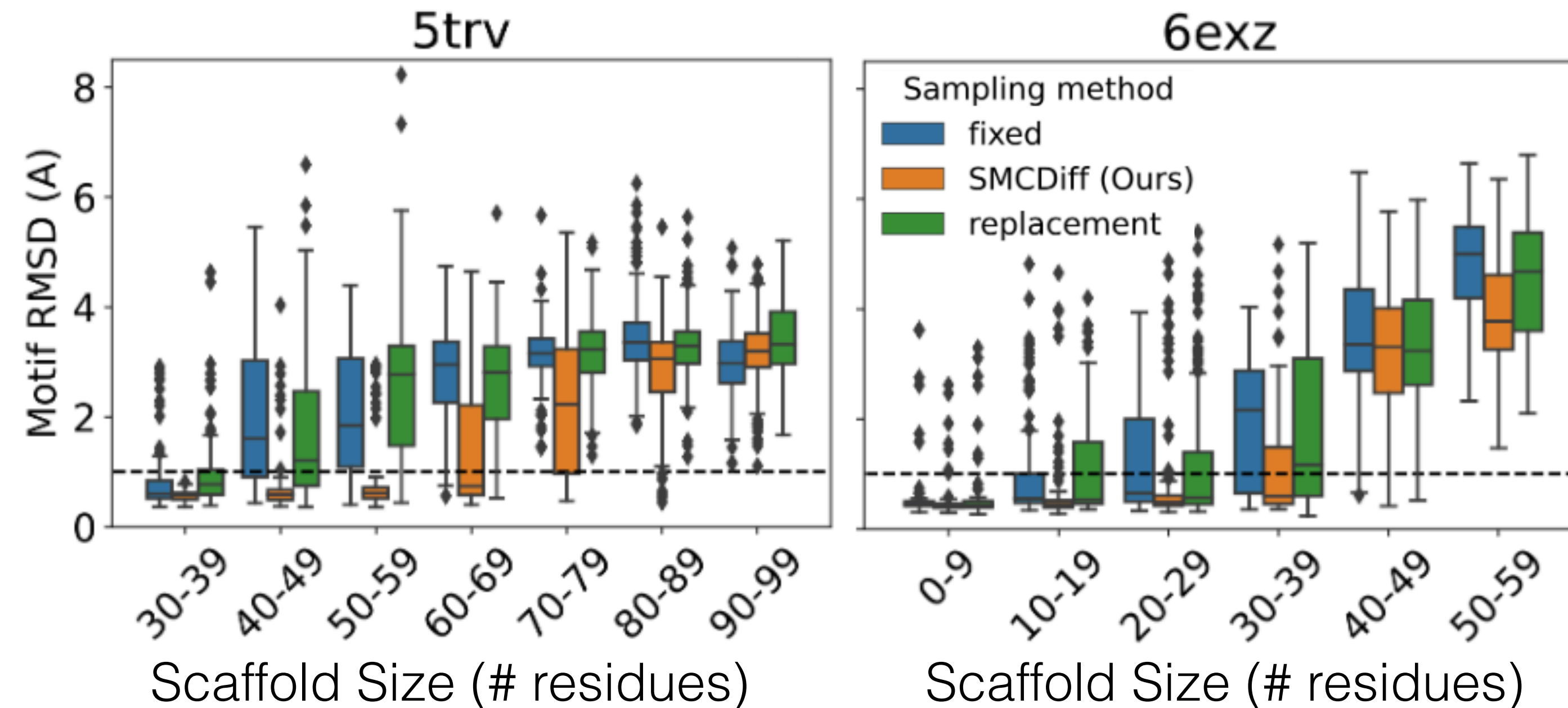
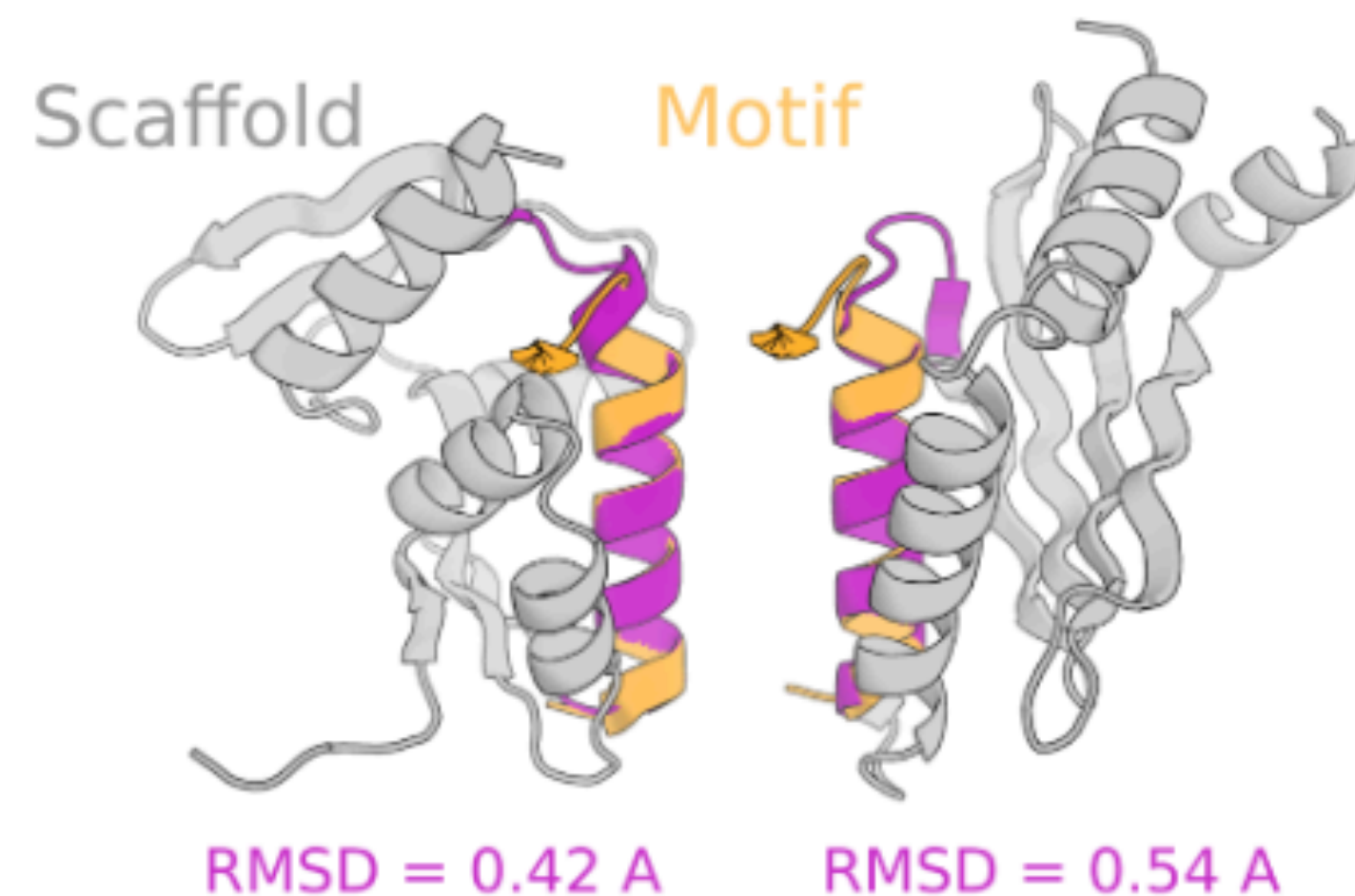
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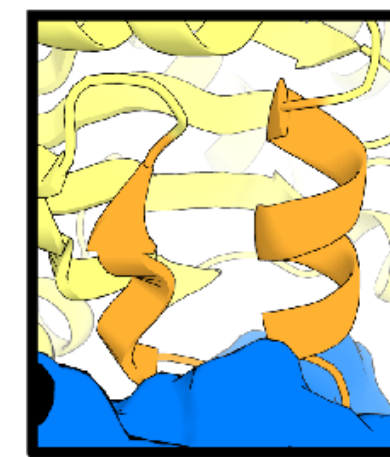
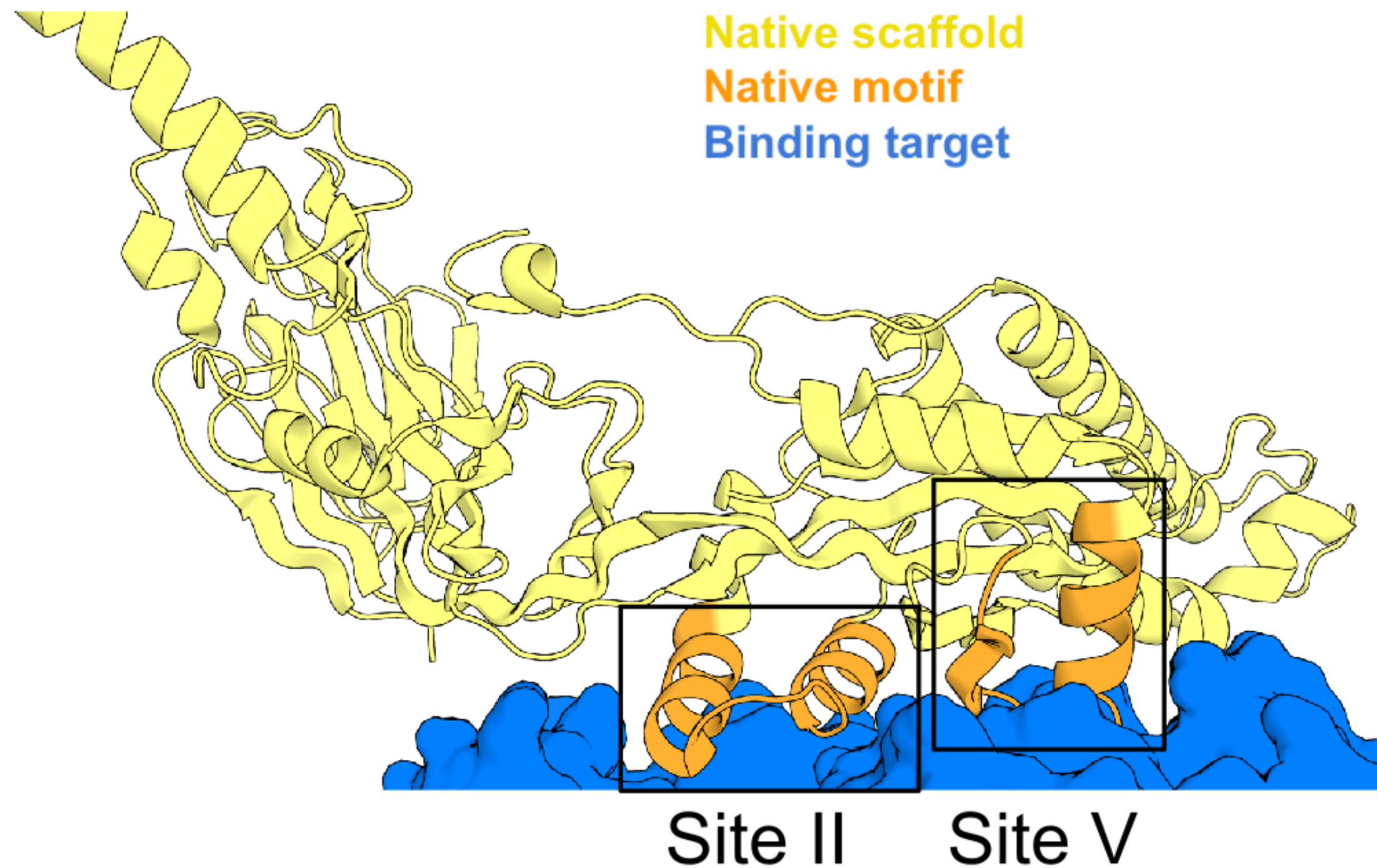
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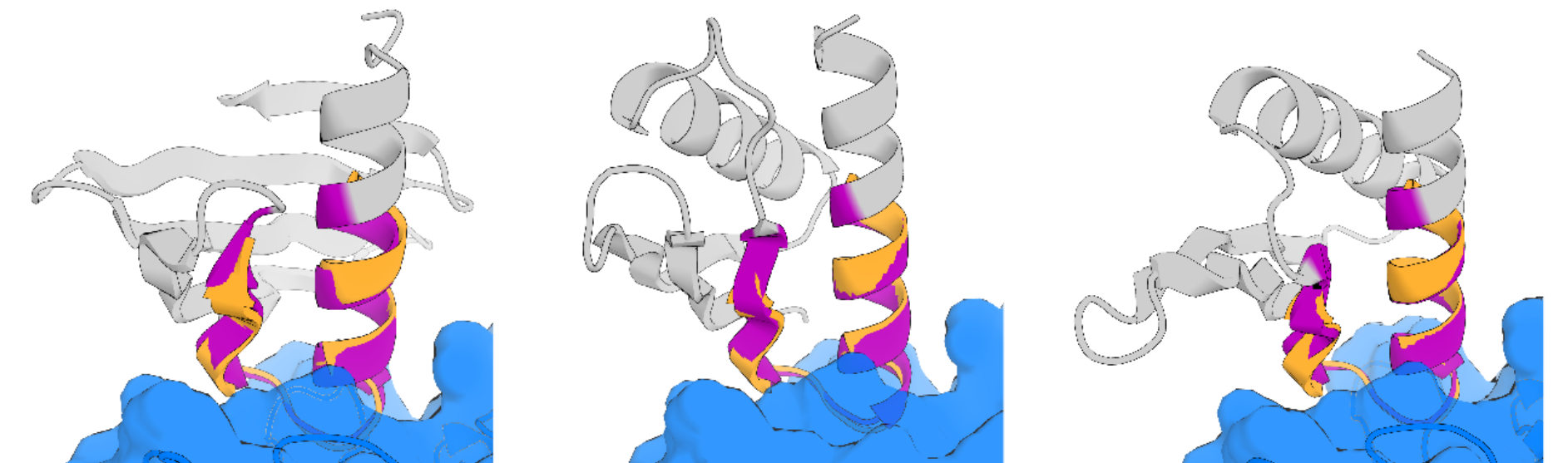
# Failure case: RSV

Respiratory syncytial virus (RSV) neutralizing antibody binds site II and V.

- RSV known to be difficult to scaffold. Only recently [Wang+2022] successfully scaffolded Site V.



Site V



- SMCDiff fails to scaffold RSV.



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**Non-generative motif-scaffolding is “state of the art”, but has limitations:**

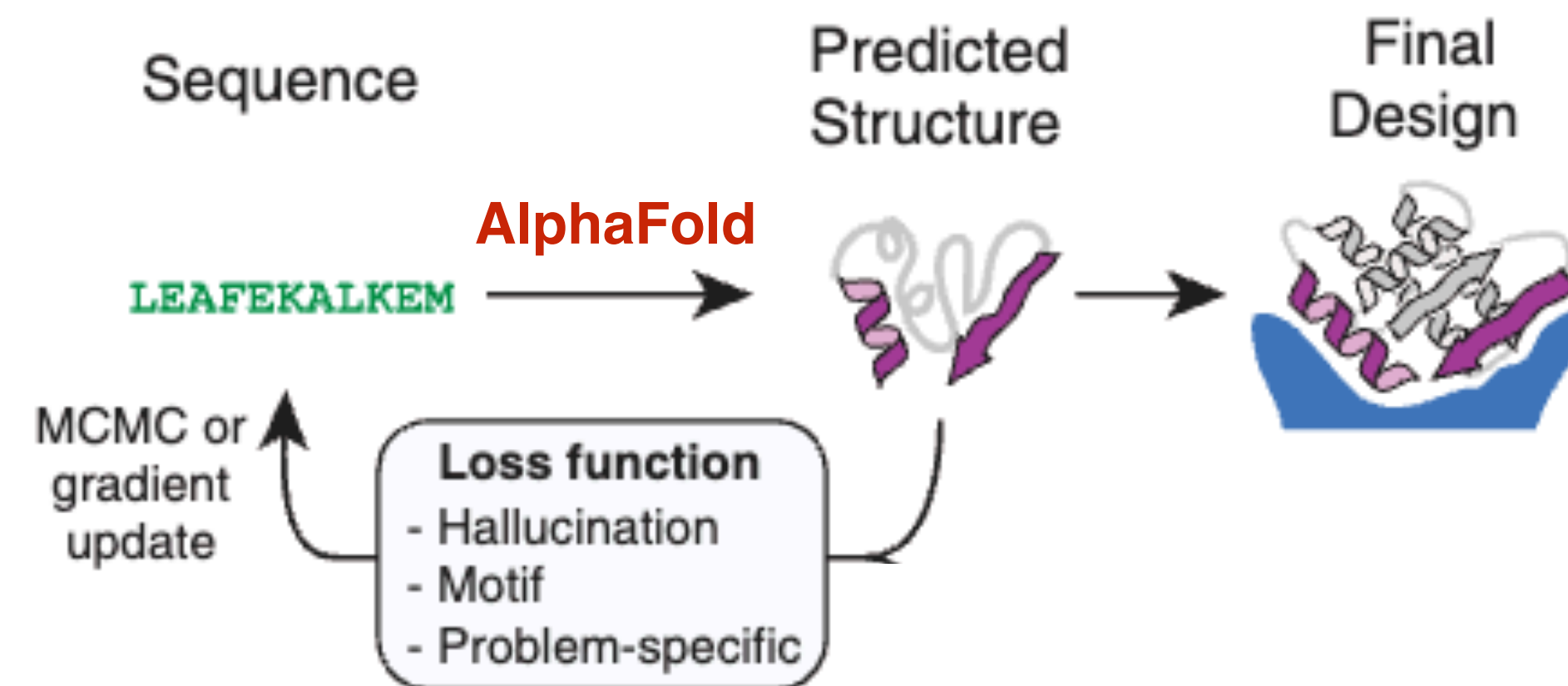


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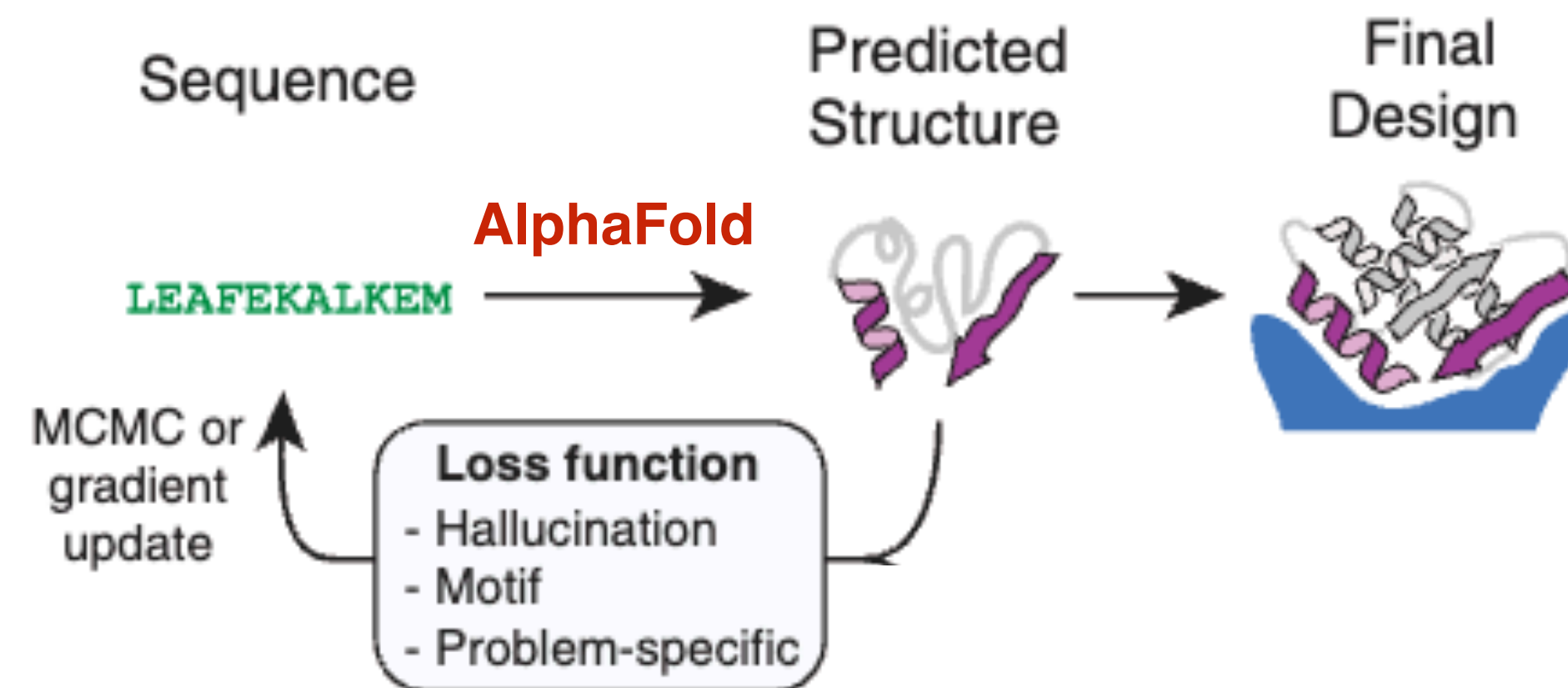


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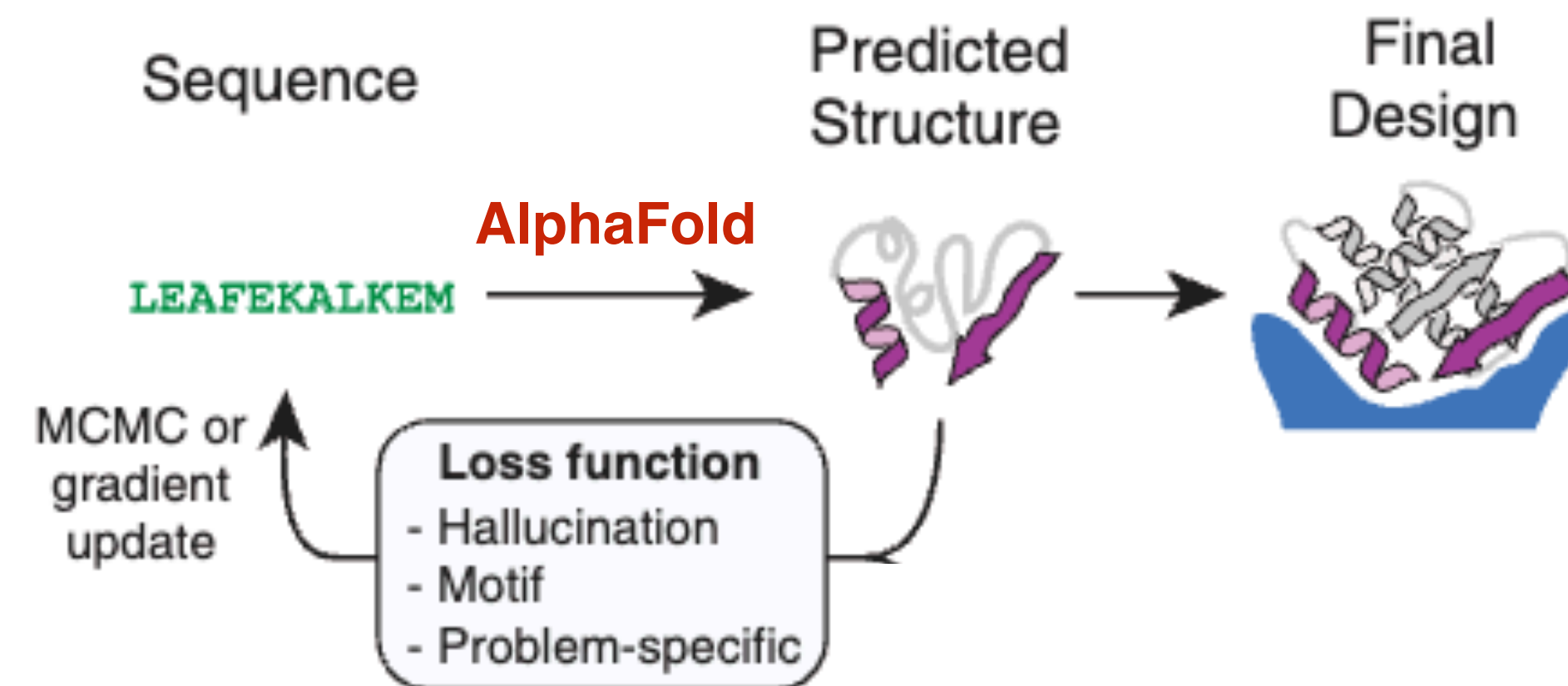


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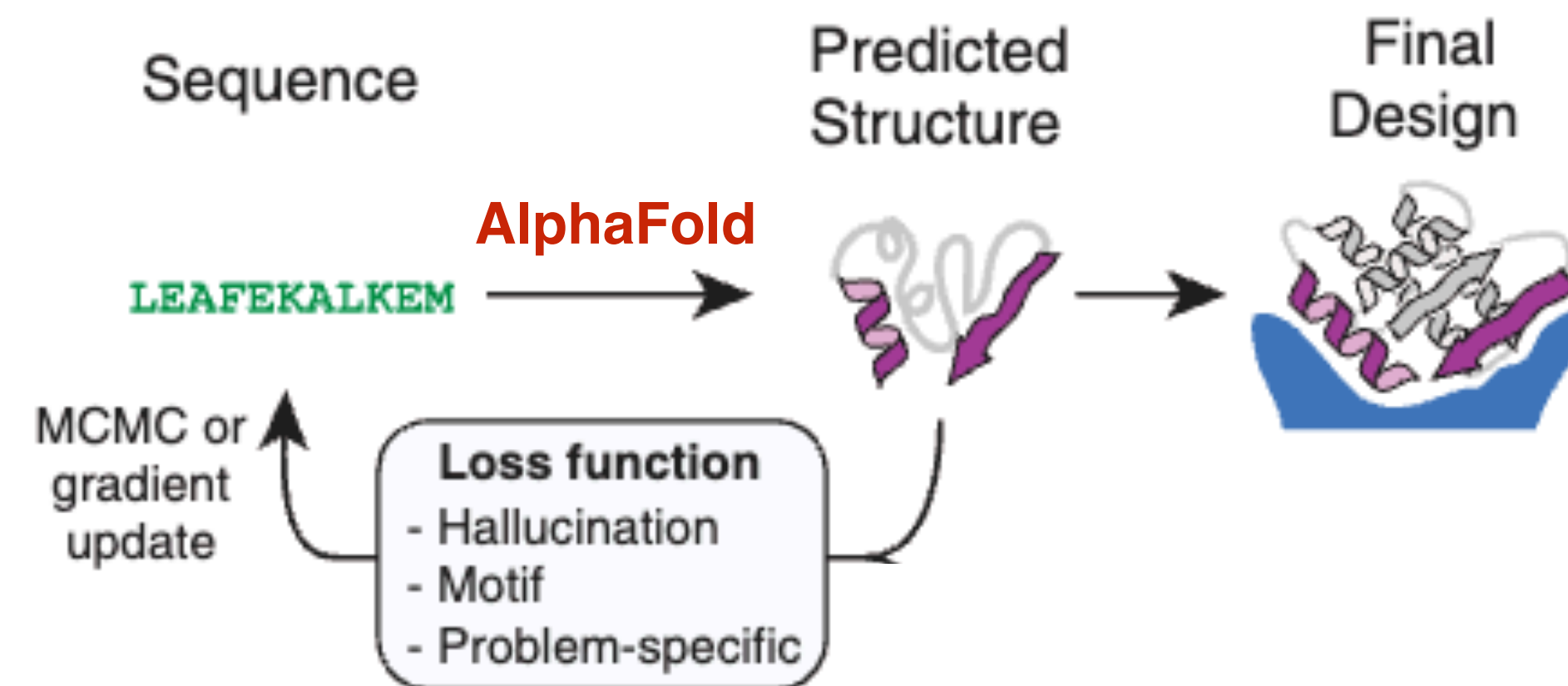


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**Generative models have made recent strides:**



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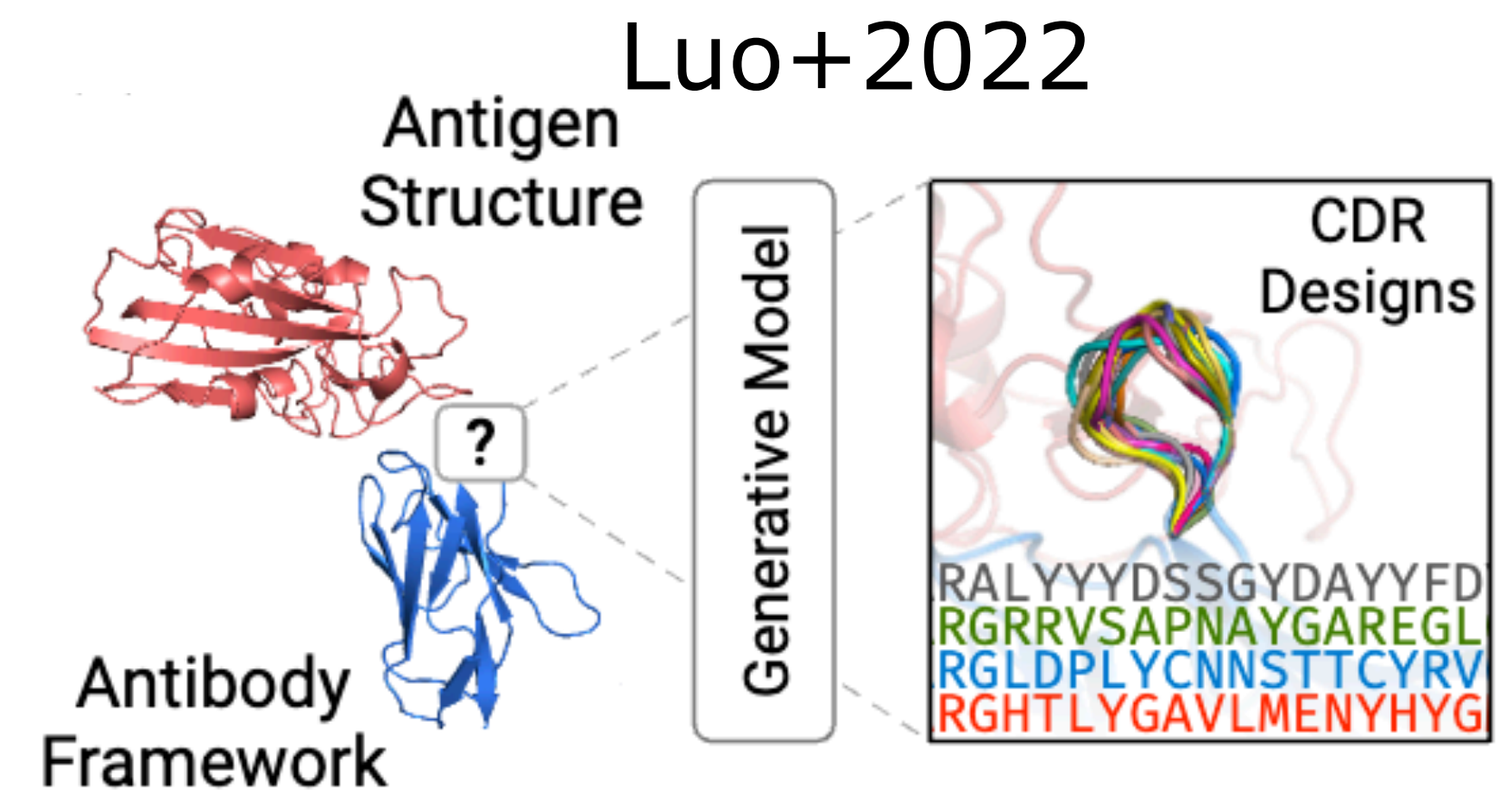
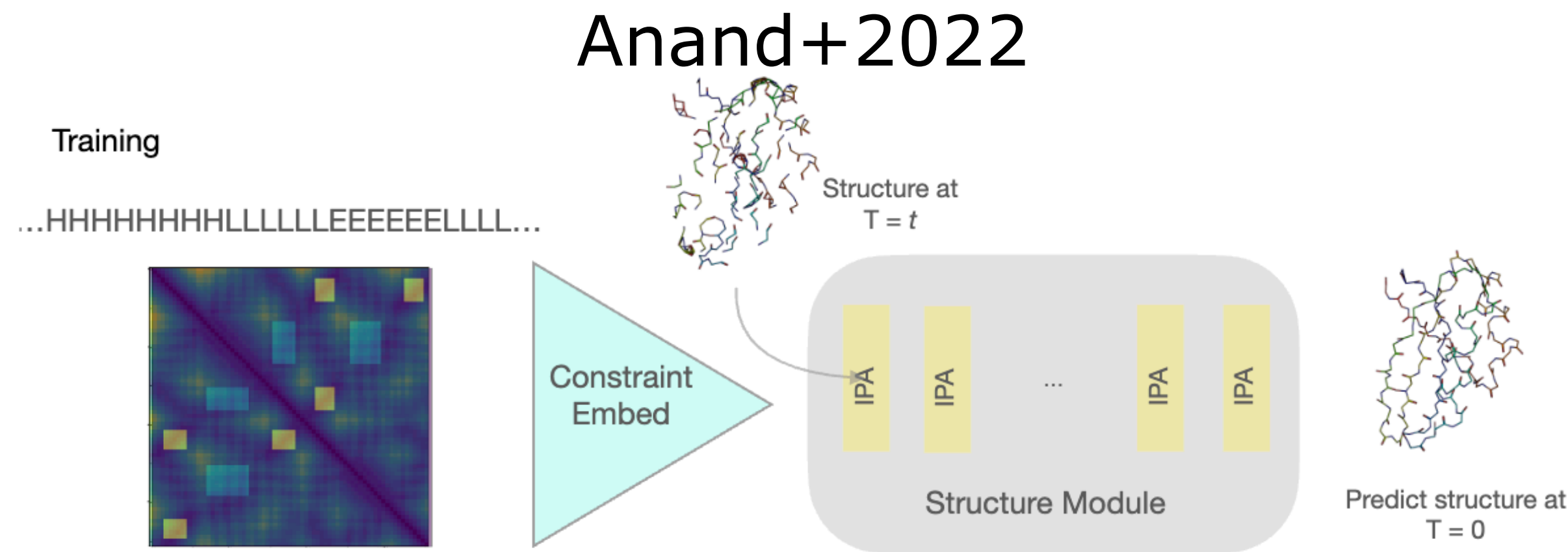
- Modeling distance matrices as images [Lin+2021, Anand+2017, Lee+2022]
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## Generative models have made recent strides:

- Modeling distance matrices as images [Lin+2021, Anand+2017, Lee+2022]
  - Rely on non-differentiable “folding” as second step
- Concurrent work on diffusion in 3D [Anand+2022, Luo+2022]
  - No demonstrations of “unconditional” sampling



# Related work on motif-scaffolding & generative modeling



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## **ProtDiff + SMCDiff Advantages:**

- Unconditional sampling of diverse backbones
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## **ProtDiff Limitations:**

- Doesn't yet extend beyond motifs in train set
- Requires pre-specifying scaffold length and motif placement



# Conclusions

Diffusion models enable a probabilistic approach to scaffolding motifs

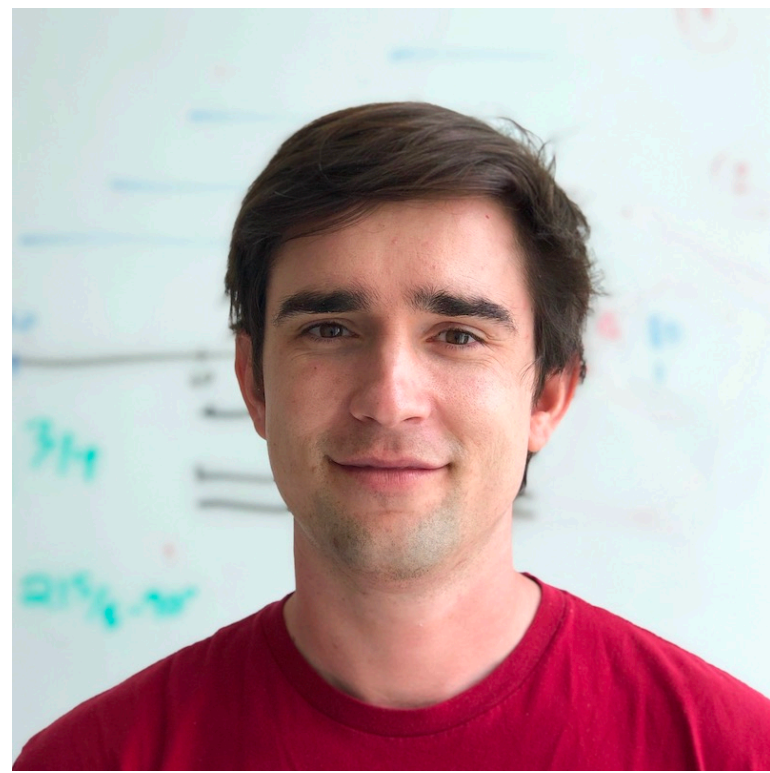
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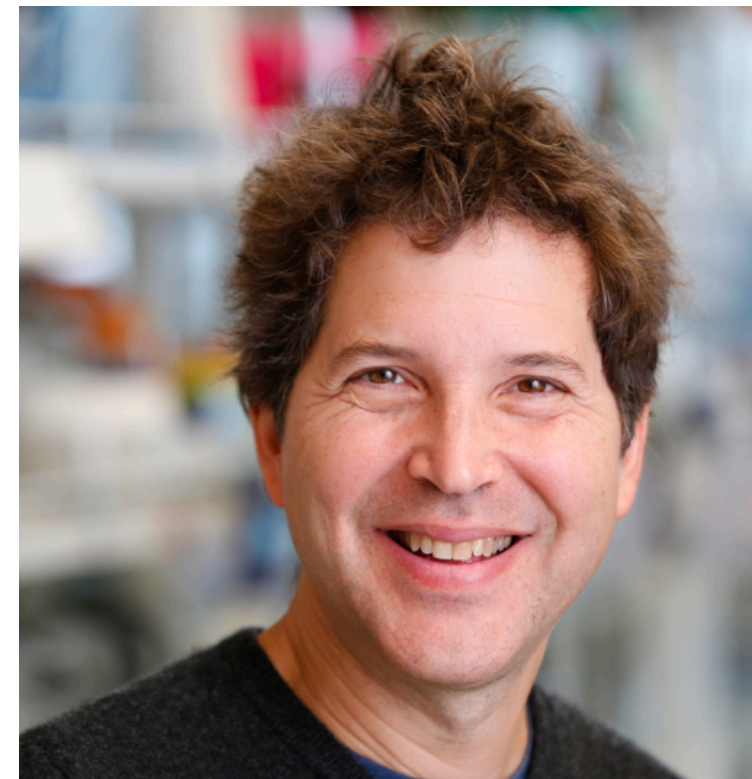
Collaborators



Doug Tischler



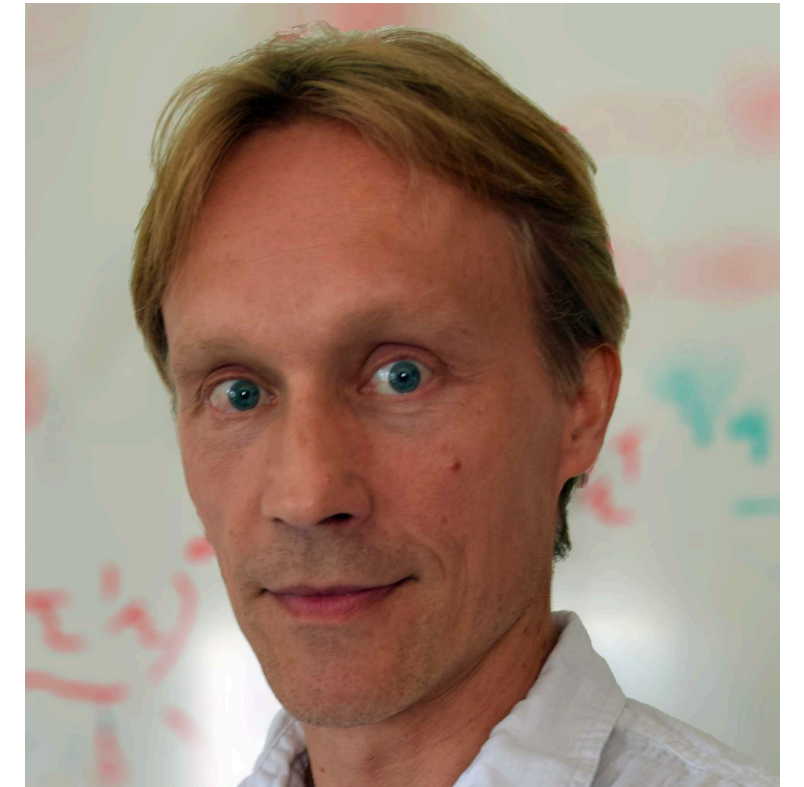
Tamara Broderick



David Baker



Regina Barzilay



Tommi Jaakkola

**Primary Reference:** "Diffusion probabilistic modeling of protein backbones in 3D for the motif-scaffolding problem." *arXiv preprint arXiv:2206.04119*

**Contact:** Brian Trippe ([blt2114@columbia.edu](mailto:blt2114@columbia.edu)), Jason Yim ([jyim@mit.edu](mailto:jyim@mit.edu))



# References

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